

# Examining power of parentage analysis for the Alaska Hatchery Research Program (AHRP)

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Anchorage, Alaska

# Why do power analysis?

- How big of an effect can we detect with current field sampling methods?
- Will that achieve program objectives?
  - RFP states  $RRS_{H/N} = 0.5$
  - Does this study need to detect smaller effects?
- How much would we need to sample to detect smaller effects?
- How would we do that?

# Outline

- Background of AHRP
- Parentage and RRS
- Proposed study design
- Simulations
- Power analysis
- Christie et al. 2014 review

# AHRP Research Questions

- 1) What is the genetic structure of pink and chum in PWS and SEAK?
- 2) What is the extent and annual variability of straying?
- 3) What is the impact on fitness (productivity) of natural pink and chum stocks due to straying hatchery pink and chum salmon?

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**What is the relative reproductive success (RRS) of hatchery-origin and natural-origin fish?**

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$$\mathbf{RRS} = \frac{\widehat{RS}_{\text{Hatchery}}}{\widehat{RS}_{\text{Natural}}}$$

# Previous research

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## Steelhead

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<sup>1</sup>Department of Zoology, Oregon State University, Corvallis, OR 97331-2914; and <sup>2</sup>Oregon Department of Fish and Wildlife, The Dalles, OR 97059-4364

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1315, 30 May 2014

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### Factors influencing the relative fitness of hatchery and wild spring Chinook salmon (*Oncorhynchus tshawytscha*) in the Wenatchee River, Washington, USA

Kevin S. Williamson, Andrew R. Murdoch, Todd N. Pearsons, Eric J. Ward, and Michael J. Ford

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Molecular Ecology (2012) 21, 5236–5250

doi: 10.1111/mec.12066

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MAUREEN A. HESS,\* CRAIG D. RABE,† JASON L. VOGEL,‡ JEFF J. STEPHENSON,\* DOUG D. NELSON† and SHAWN R. NARUM\*

## Evolutionary Applications

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ORIGINAL ARTICLE

### Reproductive success of captive bred and naturally spawned Chinook salmon colonizing newly accessible habitat

Joseph H. Anderson,<sup>1,2,\*</sup> Paul L. Faulds,<sup>2</sup> William I. Atlas,<sup>1,4</sup> and Thomas P. Quinn<sup>1</sup>

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**Keywords:** conservation, dams, hatchery, natural selection, pedigree, reproduction, sexual selection

**Abstract**

Captively reared animals can provide an immediate demographic boost in reintroduction programs, but may also reduce the fitness of colonizing populations. Construction of a fish passage facility at Landsburg Diversion Dam on the Cedar River, WA, USA, provided a unique opportunity to explore this trade-off. We thoroughly sampled adult Chinook salmon (*Oncorhynchus tshawytscha*) at the onset of colonization (2003–2009), constructed a pedigree from genotypes at 10 microsatellite loci, and calculated reproductive success (RS) as the total number of returning adult offspring. Hatchery males were consistently but not significantly less productive than naturally spawned males (range in relative RS 0.29–0.90), but the pattern for females varied between years. The sex ratio was heavily biased toward males; therefore, inclusion of the hatchery males increased the risk of a genetic fitness cost with little demographic benefit. Measurements of natural selection indicated that larger salmon had higher RS than smaller fish. Fish that arrived early to the spawning grounds tended to be more productive than later fish, although in some years, RS was maximized at intermediate dates. Our results underscore the importance of natural and sexual selection in promoting adapta-

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## Coho

### Changes in run timing and natural smolt production in a naturally spawning coho salmon (*Oncorhynchus kisutch*) population after 60 years of intensive hatchery supplementation

Michael J. Ford, Howard Fuss, Brant Boeltis, Eric LaHood, Jeffrey Hard, and Jason Miller

#### MOLECULAR ECOLOGY

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**Abstract**

Supplementation of wild salmonids with captive-bred fish is a common practice for both commercial and conservation purposes. However, evidence for lower fitness of captive-reared fish relative to wild fish has accumulated in recent years, diminishing the apparent effectiveness of supplementation as a management tool. To date, the mechanisms responsible for these fitness declines remain unknown. In this study, we showed with molecular parentage analysis that hatchery coho salmon (*Oncorhynchus kisutch*) had lower reproductive success than wild fish once they reproduced in the wild. This effect was more pronounced in males than in same-aged females. Hatchery spawned fish that were released as unfed fry (age 0), as well as hatchery fish raised for one year in the hatchery (released as smolts, age 1), both experienced lower lifetime reproductive success (RS) than wild fish. However, the subset of hatchery males that returned as 2-year olds (jacks) did not exhibit the same fitness decrease as males that returned as 3-year olds. Thus, we report three lines of evidence pointing to the absence of sexual selection in the hatchery as a contributing mechanism for fitness declines of hatchery fish in the wild: (i) hatchery fish released as unfed fry that survived to adulthood still had low RS relative to wild fish, (ii) age-3 male hatchery fish consistently showed a lower relative RS than female hatchery fish (suggesting a role for sexual selection), and (iii) age-2 jacks, which use a sneaker mating strategy, did not show the same declines as 3-year olds, which compete differently for females (again, implicating sexual selection).

**Keywords:** captive breeding, parentage analysis, reproductive success, salmonids, sexual selection, supplementation

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15 JULY 2014

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### Factors influencing the relative fitness of hatchery and wild spring Chinook salmon (*Oncorhynchus tshawytscha*) in the Wenatchee River, Washington, USA

Kevin S. Williamson, Andrew R. Murdoch, Todd N. Pearsons, Eric J. Ward, and Michael J. Ford

### MOLECULAR ECOLOGY

Molecular Ecology (2012) 21, 526–529

doi: 10.1111/mec.12026

### Supportive breeding boosts natural population abundance with minimal negative impacts on fitness of a wild population of Chinook salmon

MAUREEN A. HESS,\* CRAIG D. RABE,† JASON L. YOGEL,‡ JEFF J. STEPHENSON,\* DOUG D. NELSON† and SHAWN R. NARUM\*

### Evolutionary Applications

Evolutionary Applications ISSN 1524-4711

ORIGINAL ARTICLE

### Reproductive success of captively bred and naturally spawned Chinook salmon colonizing newly accessible habitat

Joseph H. Anderson,<sup>1,2</sup> Paul L. Fauds,<sup>1</sup> William I. Atlas<sup>1,4</sup> and Thomas P. Quinn<sup>1</sup>

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<sup>4</sup>Present address: Department of Biological Sciences, Simon Fraser University Burnaby, B.C. Canada

Keywords: conservation, demography, hatchery, natural selection, pedigree, reintroduction, sexual selection

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doi:10.1111/j.1524-4711.2012.02271.x

[Article]

## Coho

### Changes in run timing and natural smolt production in a naturally spawning coho salmon (*Oncorhynchus kisutch*) population after 60 years of intensive hatchery supplementation

Michael J. Ford, Howard Fuss, Brant Boelts, Eric LaHood, Jeffrey Hard, and Jason Miller

### MOLECULAR ECOLOGY

Molecular Ecology (2011) 20, 1860–1869

doi: 10.1111/j.1365-2108.2011.02686.x

### Reduced reproductive success of hatchery coho salmon in the wild: insights into most likely mechanisms

VÉRONIQUE THÉRIAULT,\* GREGORY R. MOYER,\*\* LAURA S. JACKSON,† MICHAEL S. BLOUNT† and MICHAEL A. BANKS\*

<sup>\*</sup>Coastal Oregon Marine Experiment Station, Hatfield Marine Science Center, Department of Fisheries and Wildlife, Oregon State University, 2030 SE Marine Science Drive, Newport, OR 97365, USA; <sup>†</sup>Oregon Department of Fish and Wildlife, 4152 N. Umpqua Highway, Reedsport, OR 97570, USA; <sup>‡</sup>Department of Zoology, 3025 Cordell Hall, Oregon State University, Corvallis, OR 97331, USA

Abstract

Supplementation of wild salmonids with captive-bred fish is a common practice for both commercial and conservation purposes. However, evidence for lower fitness of captive-reared fish relative to wild fish has accumulated in recent years, diminishing the apparent effectiveness of supplementation as a management tool. To date, the mechanisms responsible for these fitness declines remain unknown. In this study, we showed with molecular parentage analysis that hatchery coho salmon (*Oncorhynchus kisutch*) had lower reproductive success than wild fish once they reproduced in the wild. This effect was more pronounced in males than in same-aged females. Hatchery spawned fish that were released as unfed fry (age 0), as well as hatchery fish raised for one year in the hatchery (reared as smolts, age 1), both experienced lower lifetime reproductive success (RS) than wild fish. However, the subset of hatchery males that returned as 2-year olds (jacks) did not exhibit the same fitness decrease as males that returned as 3-year olds. Thus, we report three lines of evidence pointing to the absence of sexual selection in the hatchery as a contributing mechanism for fitness declines of hatchery fish in the wild: (i) hatchery fish released as unfed fry that survived to adulthood still had low RS relative to wild fish, (ii) age-3 male hatchery fish consistently showed a lower relative RS than female hatchery fish (suggesting a role for sexual selection), and (iii) age-2 jacks, which use a sneaker mating strategy, did not show the same declines as 3-year olds, which compete differently for females (again, implicating sexual selection).

Keywords: captive breeding, parentage analysis, reproductive success, salmonids, sexual selection, supplementation

Received 20 January 2010; revision received 14 January 2011; accepted 18 January 2011

## Chum

### Reproductive behavior and relative reproductive success of natural- and hatchery-origin Hood Canal summer chum salmon (*Oncorhynchus keta*)

Barry A. Berejikian, Donald M. Van Doornik, Julie A. Scheurer, and Richard Bush

Abstract: Estimates of the relative fitness of hatchery- and natural-origin salmon can help determine the value of hatchery stocks in contributing to recovery efforts. This study compared the ability to fry reproductive success of natural-origin summer chum salmon (*Oncorhynchus keta*) with that of first- to third-generation hatchery-origin salmon in an experiment that included four replicate broodstock groups. Hatchery- and natural-origin chum salmon exhibited similar reproductive success. Hatchery and natural-origin males obtained similar access to nesting females, and females of both types exhibited similar breeding behaviors and durations. Male body size was positively correlated with access to nesting females and reproductive success. The estimates of relative reproductive success (hatchery/natural = 0.83) in this study were similar to those in other studies of other anadromous salmonids in which the hatchery population was founded from the local natural population and much higher than those in studies that evaluated the lifetime relative reproductive success of monokal hatchery populations.

2543

781

# Previous research

Steelhead

Chinook

Coho

- Also lots of Atlantic salmon

- No pinks, few chum

- Different hatchery culture

- Smaller releases

- None in Alaska

- Sample ↑ prop. adults/offspring

**Differential reproductive success of sympatric, naturally spawning hatchery and wild steelhead trout (*Oncorhynchus mykiss*) through the adult stage**

Jennifer E. Morgan, Paul B. Bestgen, and Thomas P. Quinn

**MOLECULAR ECOLOGY**

Molecular Ecology (2011) 20, 1860–1869 doi: 10.1111/j.1365-294X.2011.02856.x

**Reduced reproductive success of hatchery coho salmon in the wild: insights into most likely mechanisms**

VÉRONIQUE THÉRIAULT,<sup>1</sup> GREGORY R. MOYER,<sup>1\*</sup> LAURA S. JACKSON,<sup>1</sup> MICHAEL S. BLOUNT,<sup>1</sup> and MICHAEL A. BANKS<sup>2</sup>

**Genetic Effects of Captive Breeding Cause a Rapid, Cumulative Fitness Decline in the Wild**

Hitoshi Araki,<sup>1</sup> Becky Cooper, Michael S. Blouin

**Effective population size of steelhead trout: influence of variance in reproductive success, hatchery programs, and genetic compensation between life-history forms**

Hitoshi Araki,<sup>1</sup> Robert M. Waples,<sup>1</sup> Michael S. Blouin<sup>1</sup>

**biology letters**

Carry-over effect of captive breeding reduces fitness of wild-born descendants

Hitoshi Araki<sup>1,2</sup>, Becky Cooper and Michael S. Blouin

**Transactions of the American Fisheries Society**

**Disinherited: Reproductive Success of Steelhead in a Hatchery Supplement Program**

Erison A. Bertoni,<sup>1</sup> Richard L. Carmichael,<sup>2</sup> Richard A. Fisher,<sup>3</sup> Eric J. Ward,<sup>4</sup> Paul Moran<sup>5</sup>

**Genetic adaptation to captivity can occur in a single generation**

Mark R. Christie<sup>1</sup>, Melanie L. Marline<sup>1</sup>, Rod A. French<sup>2</sup>, and Michael S. Blouin<sup>1</sup>

**Use of Parentage Analysis to Determine Reproductive Success of Hatchery-Origin Spring Chinook Salmon Outplanted in the Wild**

Robert Spathek<sup>1,2</sup> and Geoff FitzGerald<sup>3</sup>

**MOLECULAR ECOLOGY**

Molecular Ecology (2012) 21, 5236–5250 doi: 10.1111/jee.12306

**Factors influencing the relative fitness of hatchery and wild spring Chinook salmon (*Oncorhynchus tshawytscha*) in the Wenatchee River, Washington, USA**

Maureen A. Hess,<sup>1</sup> Craig D. Raef,<sup>1</sup> Jason L. Vogel,<sup>2</sup> Jeff J. Stephenson,<sup>1</sup> Doug D. Nelson<sup>1</sup> and Shawn R. Narum<sup>1</sup>

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Maureen A. Hess,<sup>1</sup> Craig D. Raef,<sup>1</sup> Jason L. Vogel,<sup>2</sup> Jeff J. Stephenson,<sup>1</sup> Doug D. Nelson<sup>1</sup> and Shawn R. Narum<sup>1</sup>

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**Abstract**

**Chum**

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Barry A. Berejikian, Donald M. Van Doornik, Julie A. Scheurer, and Richard Bush

# Outline

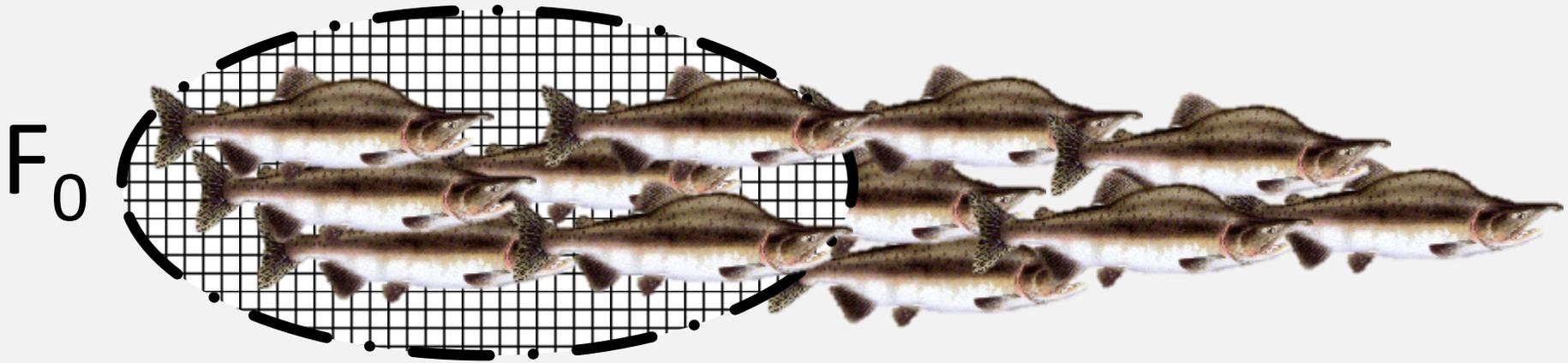
- Background of AHRP
- **Parentage and RRS**
- Proposed study design
- Simulations
- Power analysis
- Christie et al. 2014 review

# How to measure RS?

$F_0$



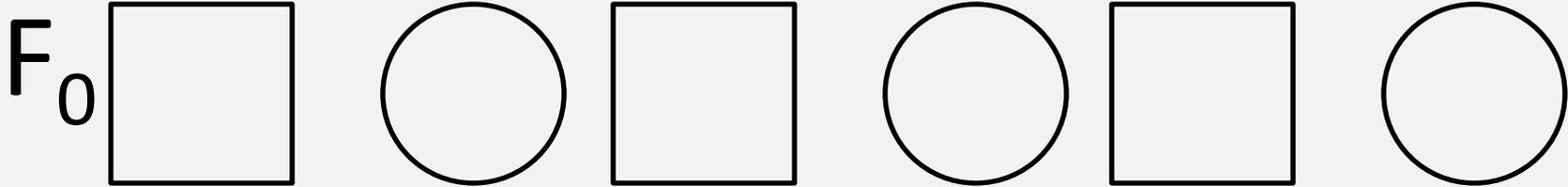
# How to measure RS?



# How to measure RS?



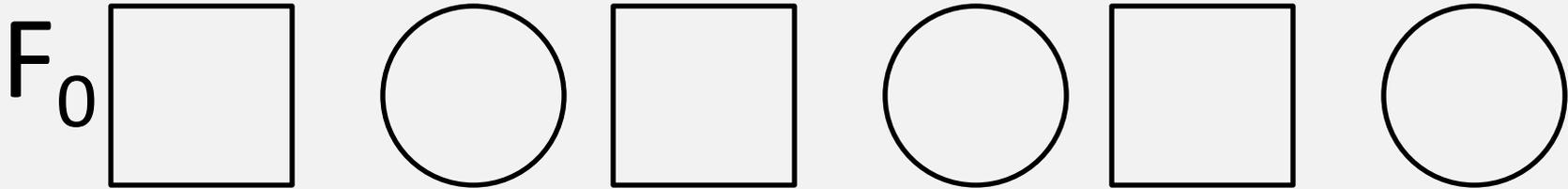
# How to measure RS?



Male

Female

# How to measure RS?

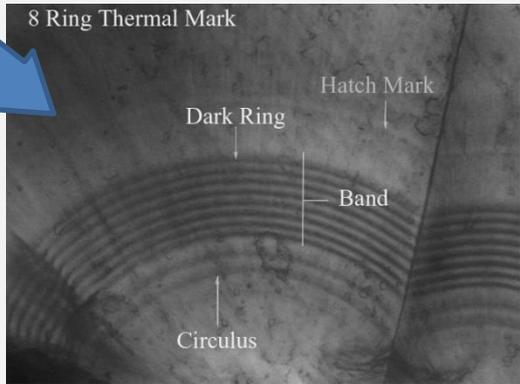
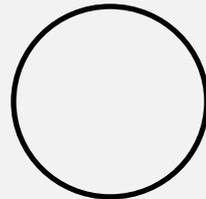
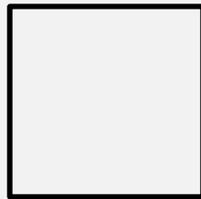
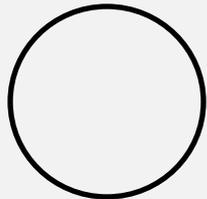
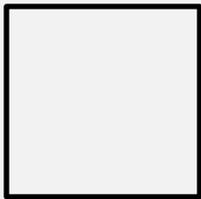
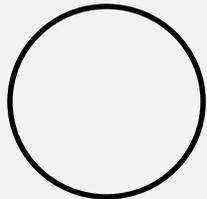
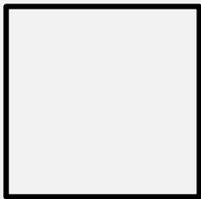


Male

Female

# How to measure RS?

F<sub>0</sub>

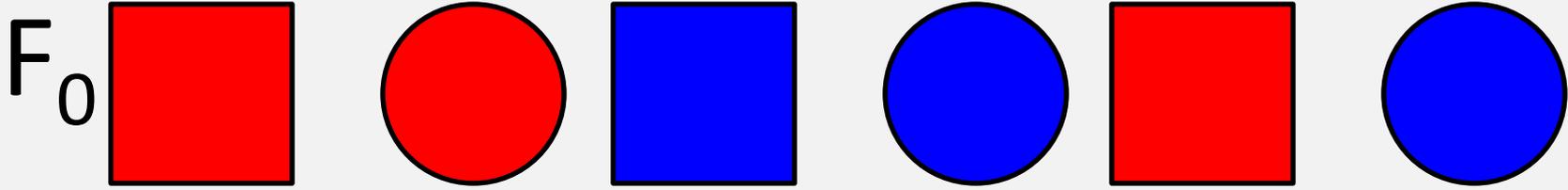


Hatchery-origin

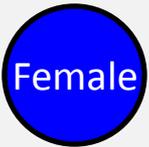
Male

Female

# How to measure RS?

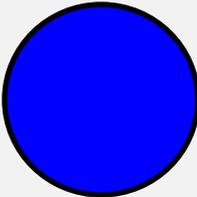
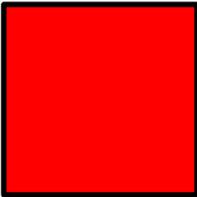
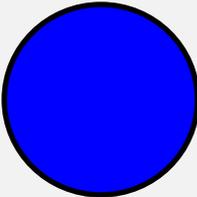
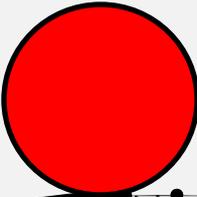
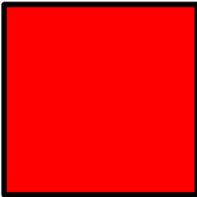


**Natural**      **Hatchery**

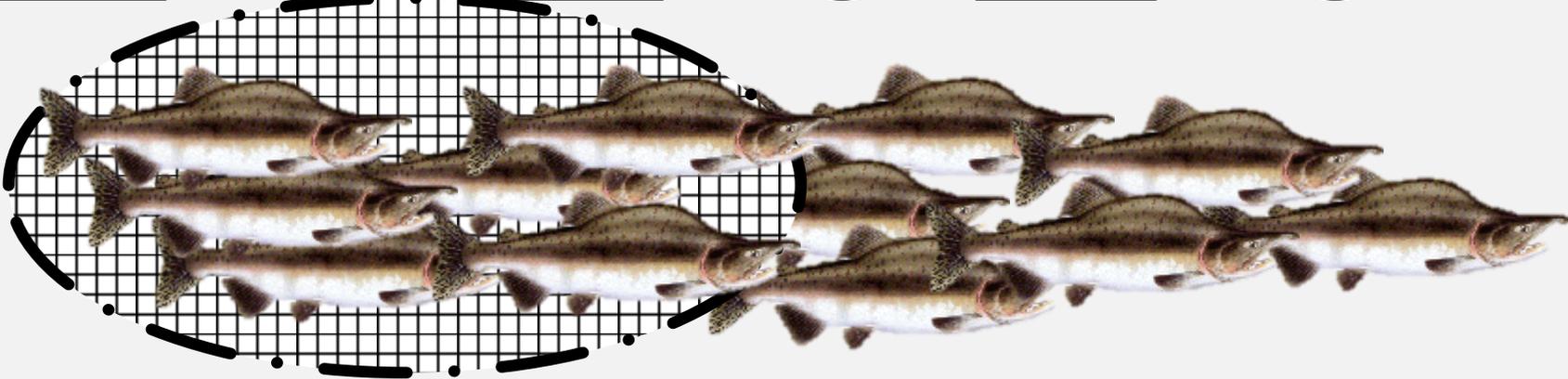


# How to measure RS?

$F_0$



$F_1$

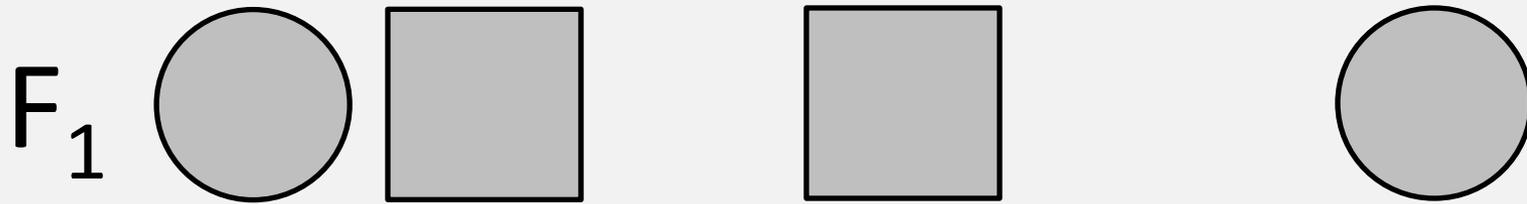
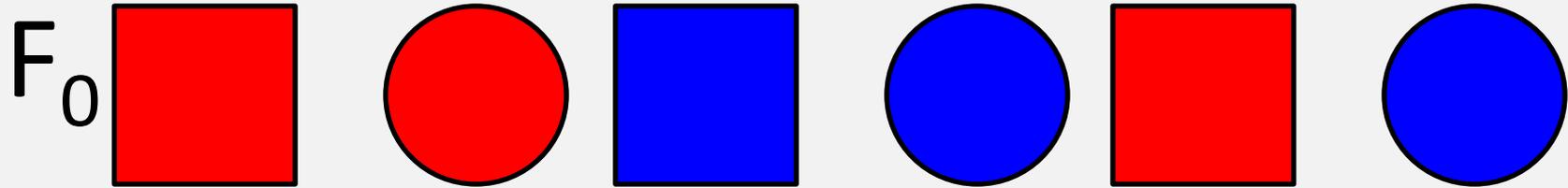


Natural

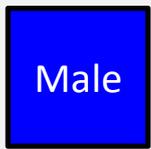
Hatchery



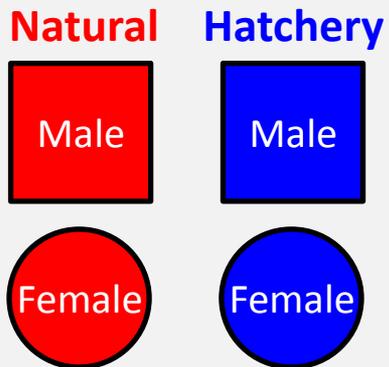
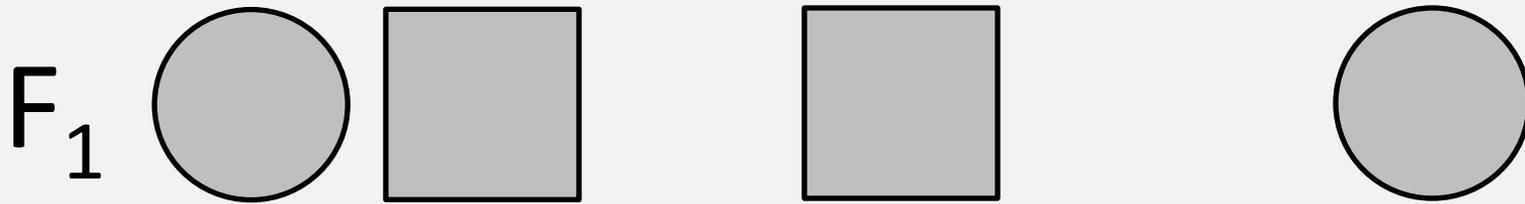
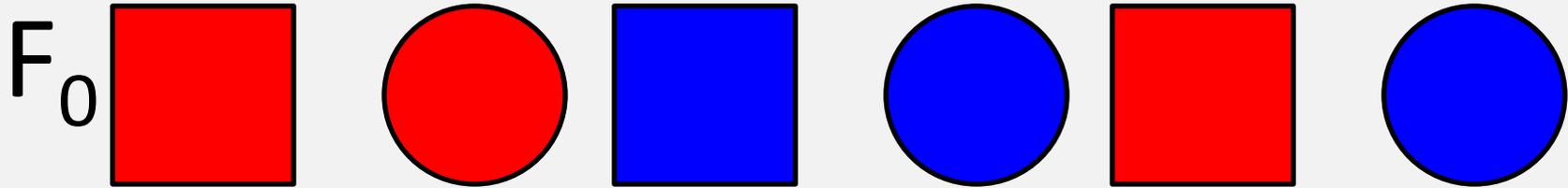
# How to measure RS?



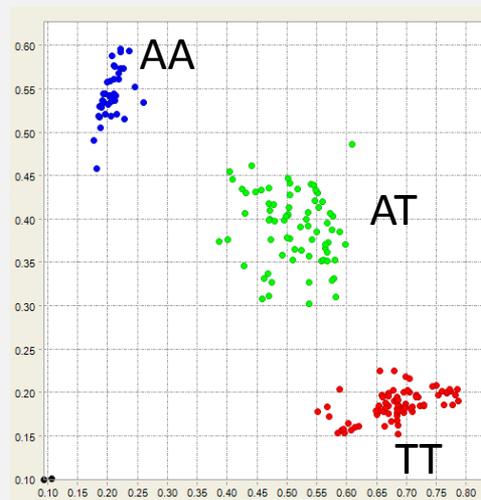
Natural Hatchery



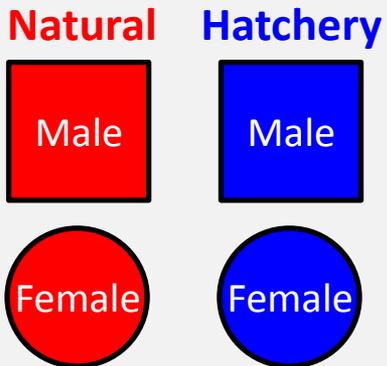
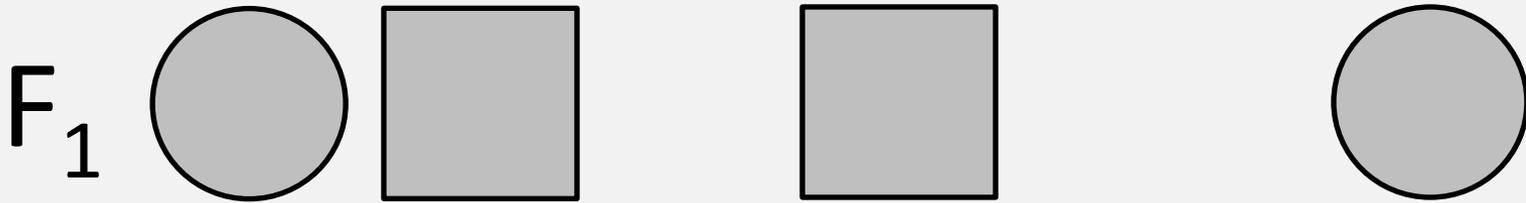
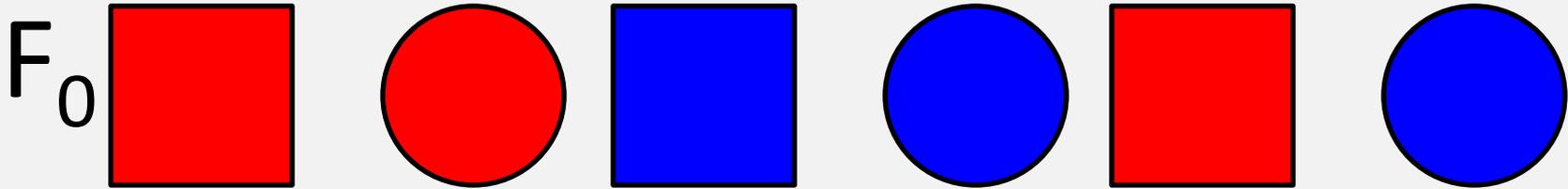
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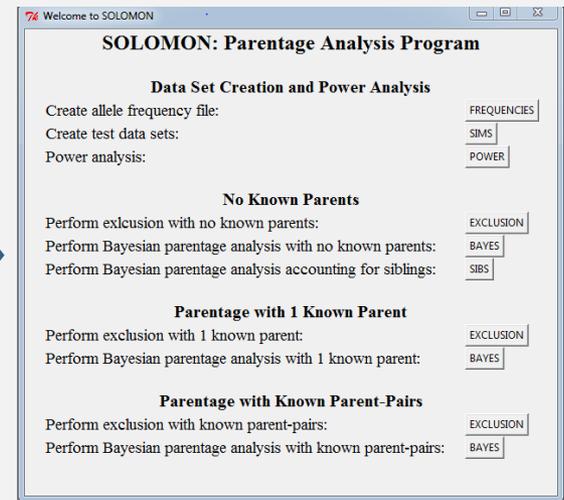
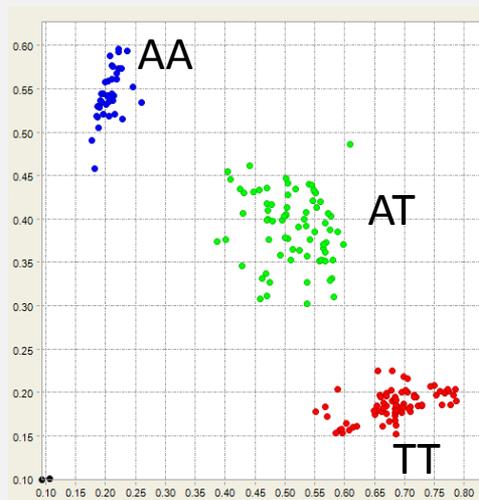
96  
SNPs



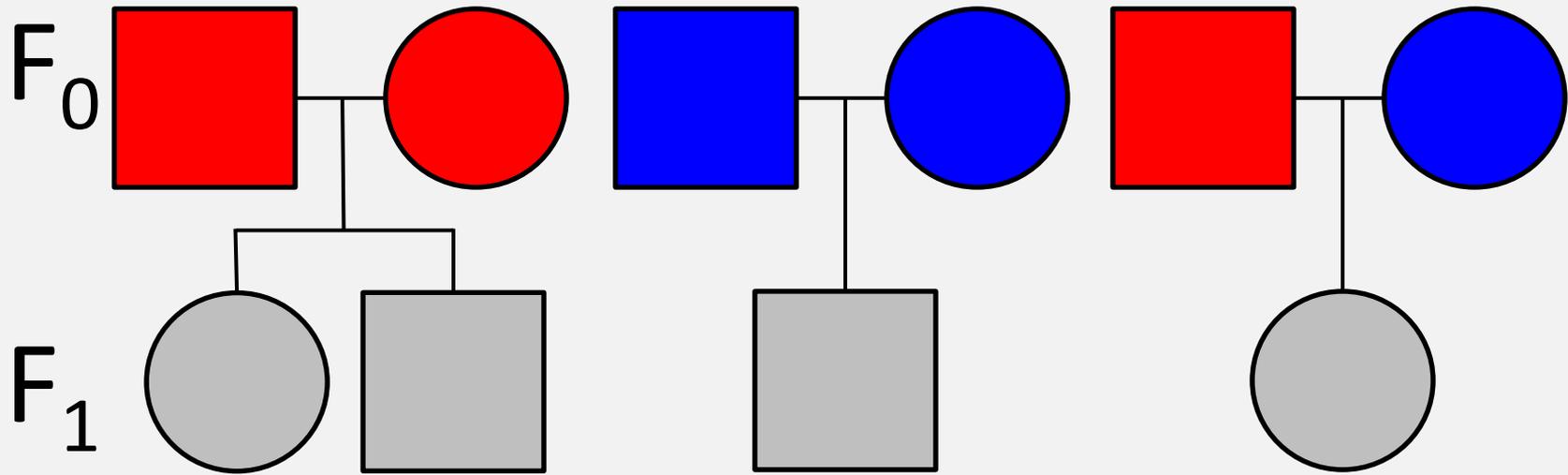
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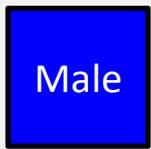
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SNPs



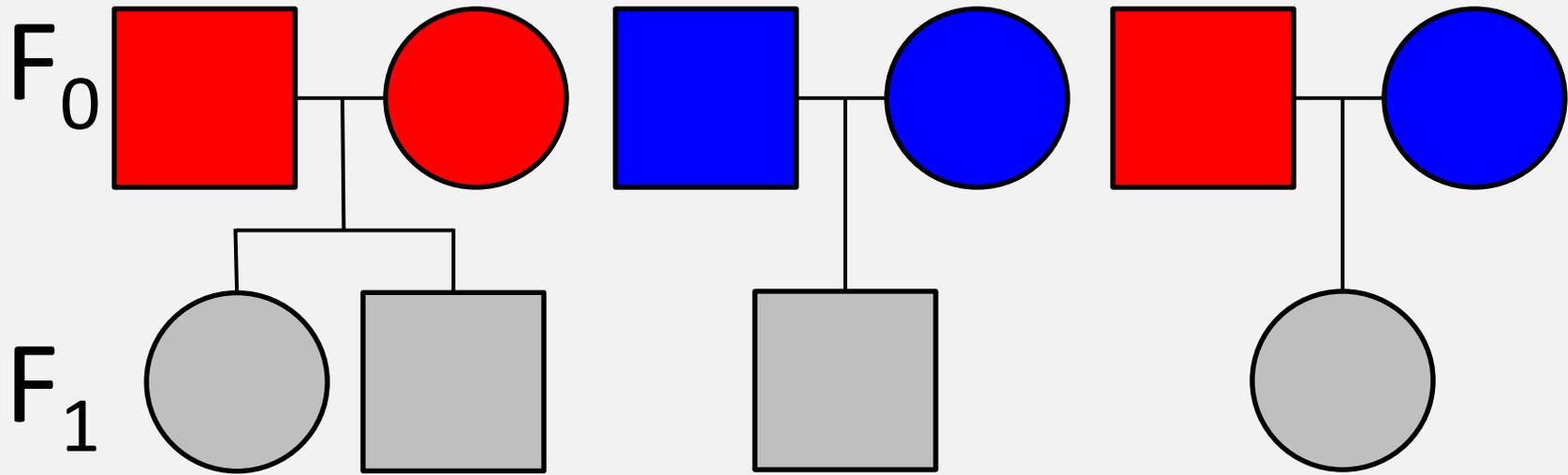
# How to measure RS?



Natural Hatchery

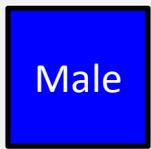


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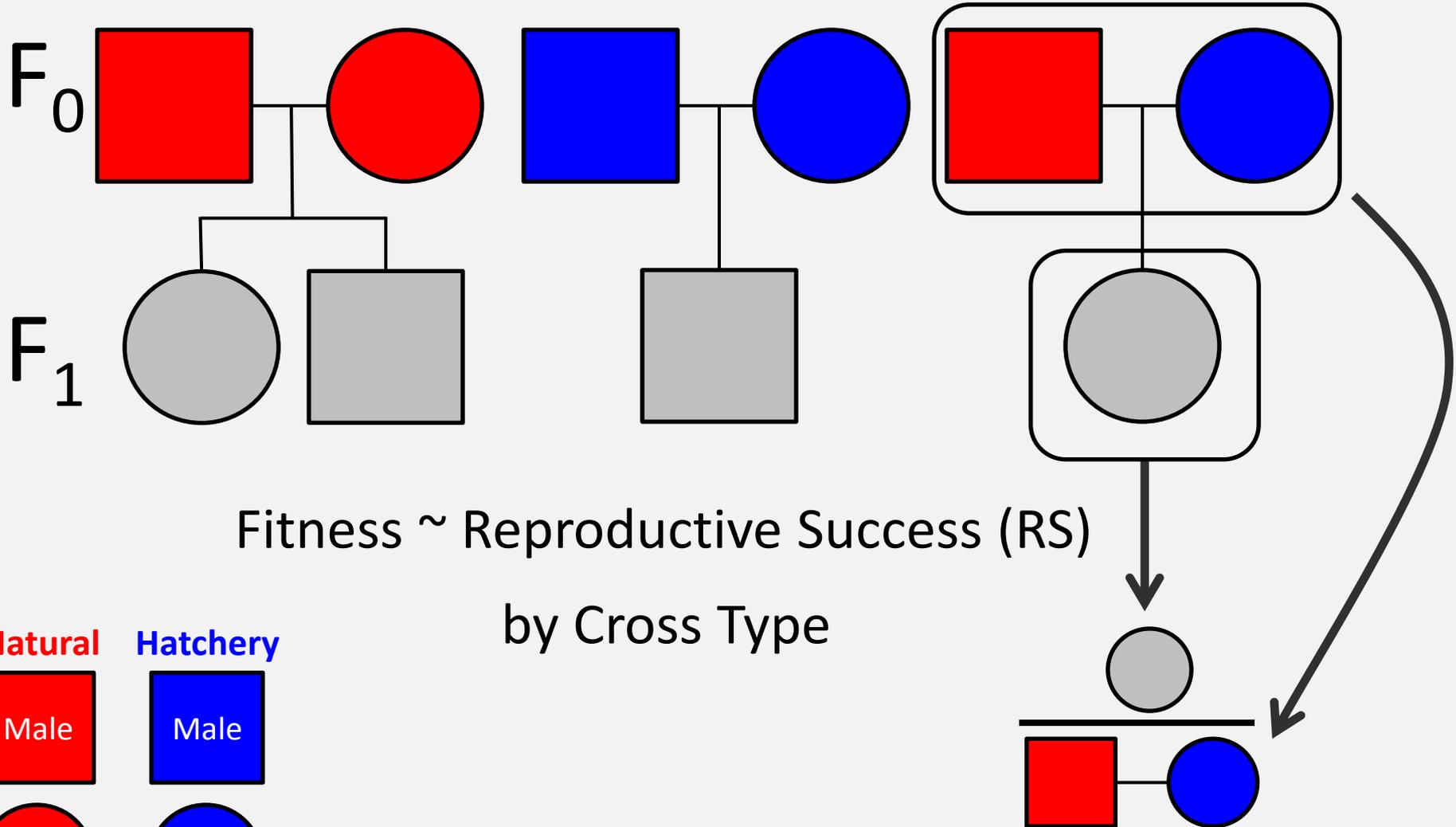


Fitness  $\sim$  Reproductive Success (RS)

Natural Hatchery

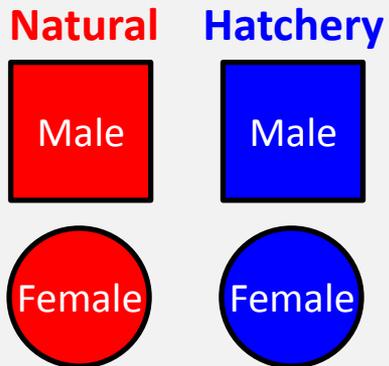


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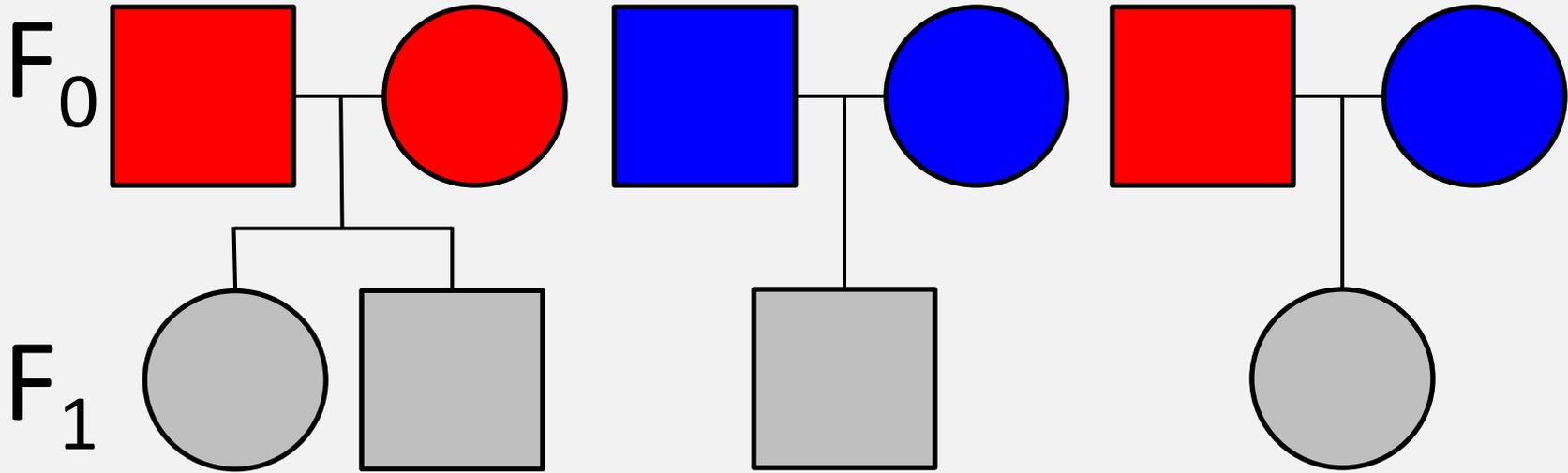


Fitness  $\sim$  Reproductive Success (RS)

by Cross Type



# How to measure RS?

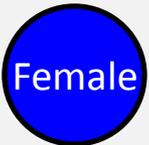


$$RS_{N/N} = 2$$

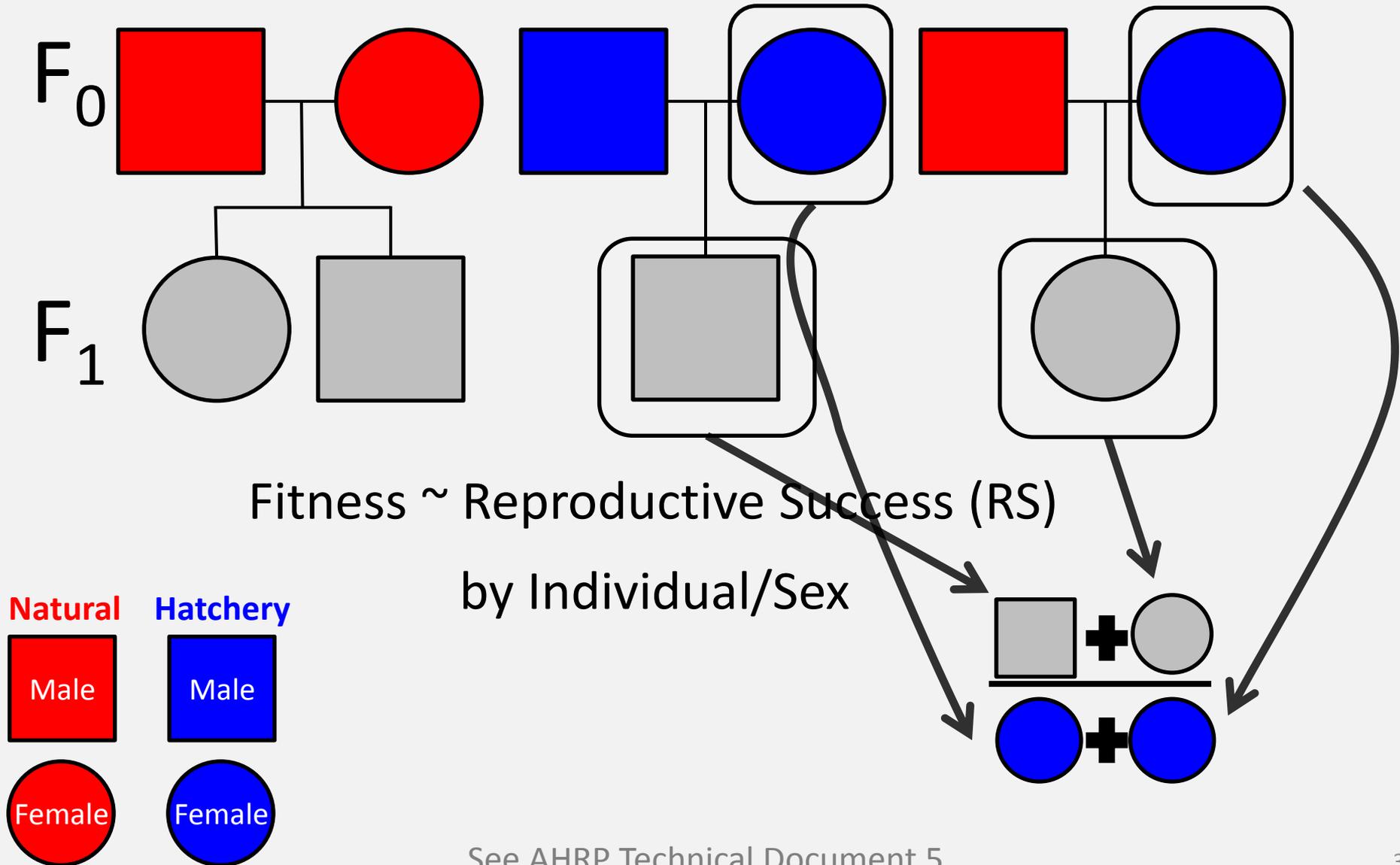
$$RS_{H/H} = 1$$

$$RS_{H/N} = 1$$

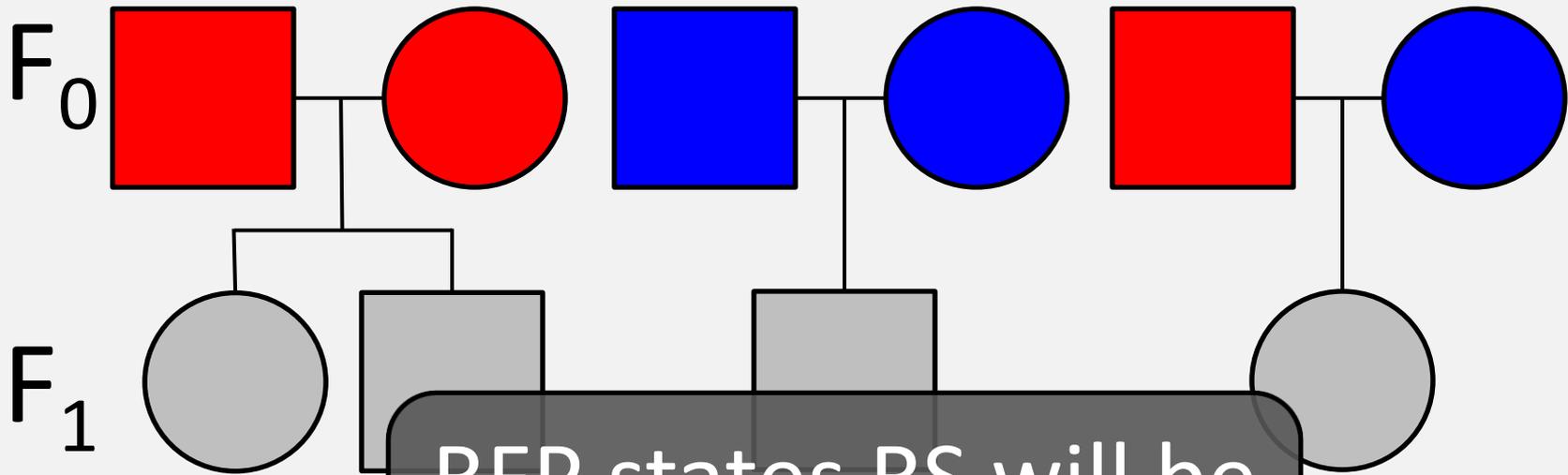
Natural Hatchery



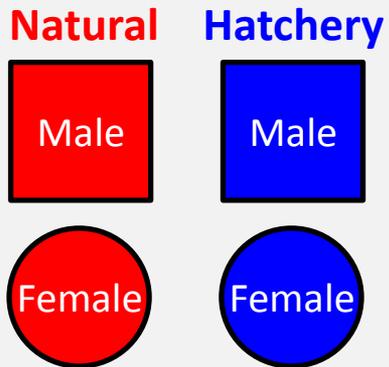
# How to measure RS?



# How to measure RS?



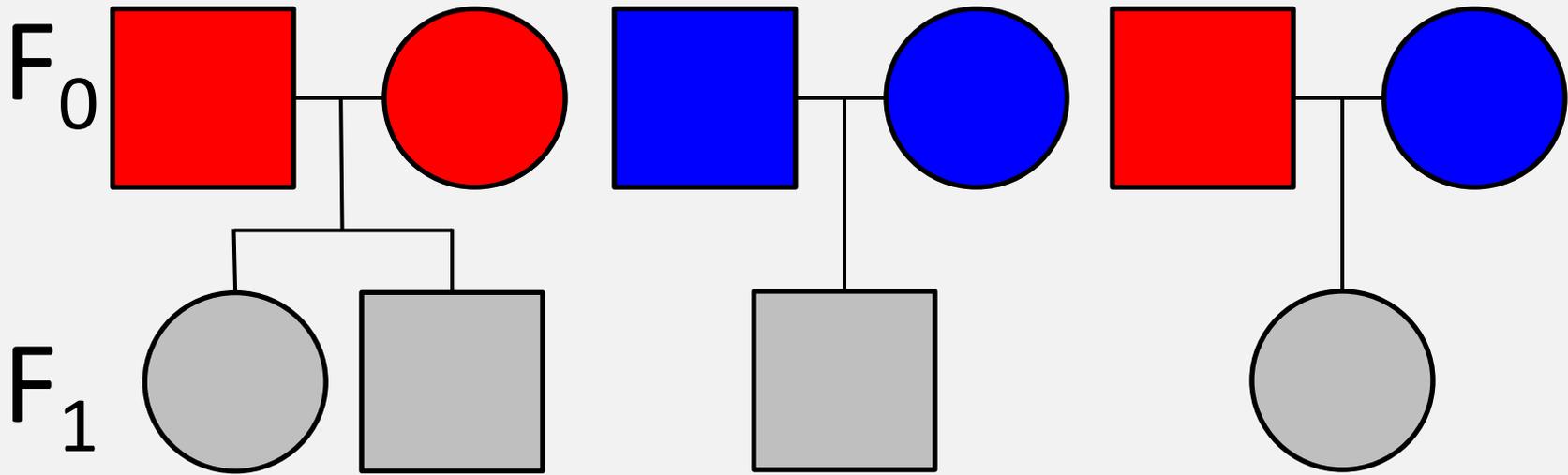
RFP states RS will be determined by sex = 1



$RS_{N \text{ Male}} = 1.5$

$RS_{H \text{ Male}} = 1$

# How to measure RS?



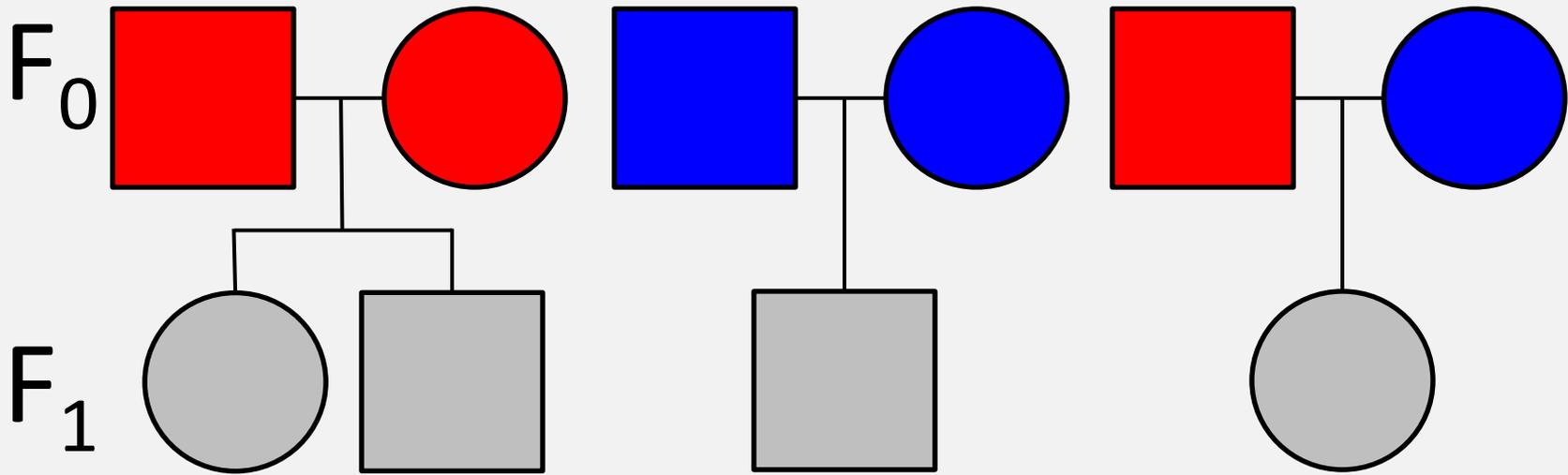
Fitness  $\sim$  Reproductive Success (RS)

Natural Hatchery

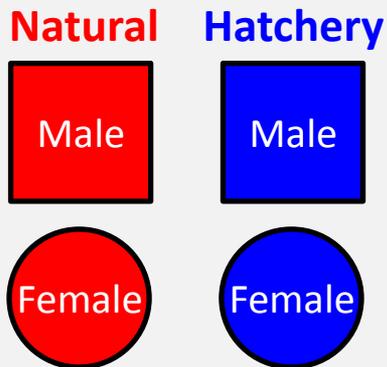


Relative Reproductive Success (RRS)

# How to measure RS?



Fitness  $\sim$  Reproductive Success (RS)



Relative Reproductive Success (RRS)

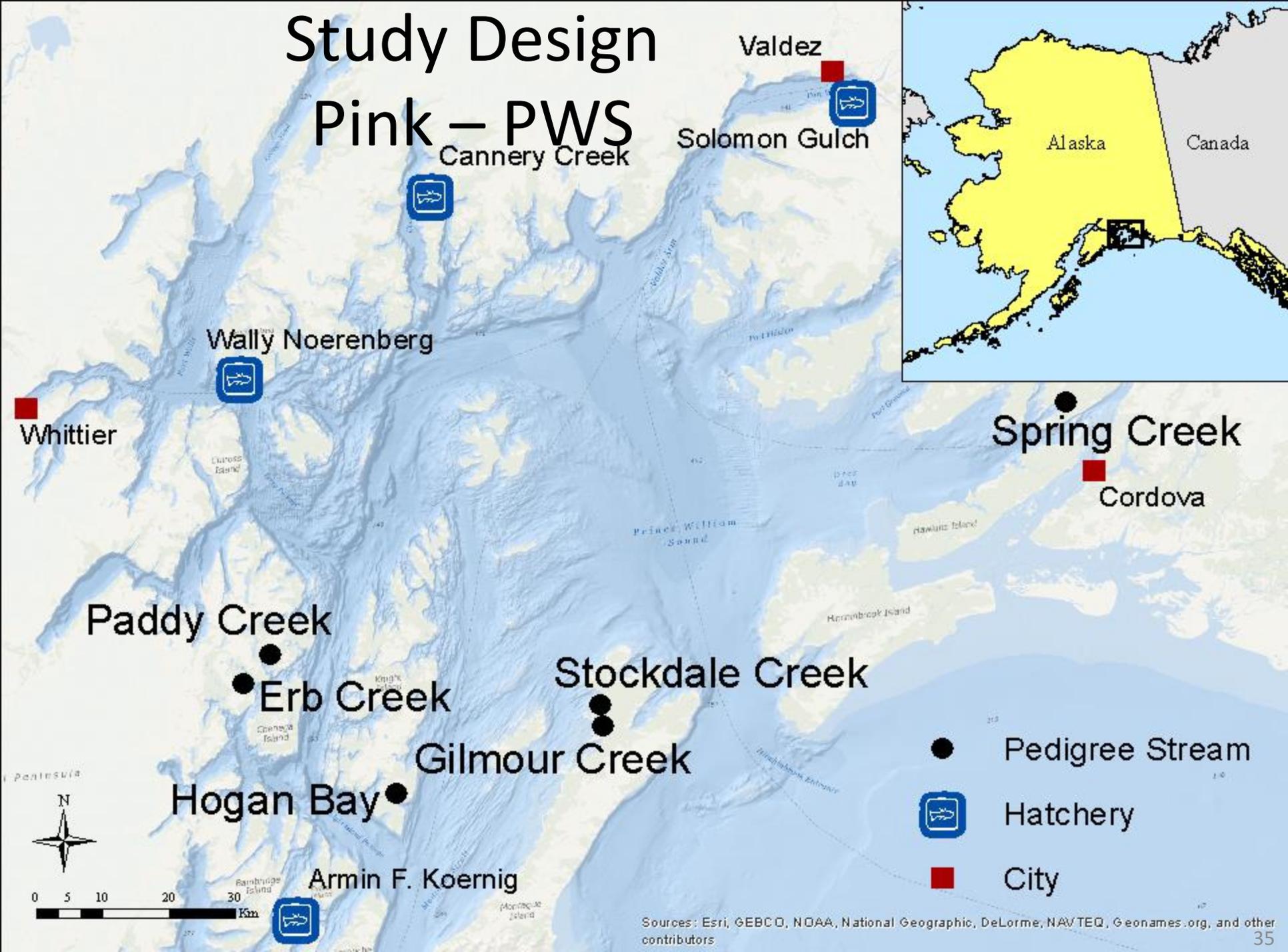
$$RRS = \frac{\widehat{RS}_{\text{Hatchery}}}{\widehat{RS}_{\text{Natural}}}$$

# Outline

- Background of AHRP
- Parentage and RRS
- **Proposed study design**
- Simulations
- Power analysis
- Christie et al. 2014 review

# Study Design

## Pink – PWS



Valdez

Solomon Gulch

Cannery Creek

Wally Noerenberg

Whittier

Spring Creek

Cordova

Paddy Creek

Erb Creek

Stockdale Creek

Gilmour Creek

Hogan Bay

Armin F. Koernig

Alaska

Canada

● Pedigree Stream

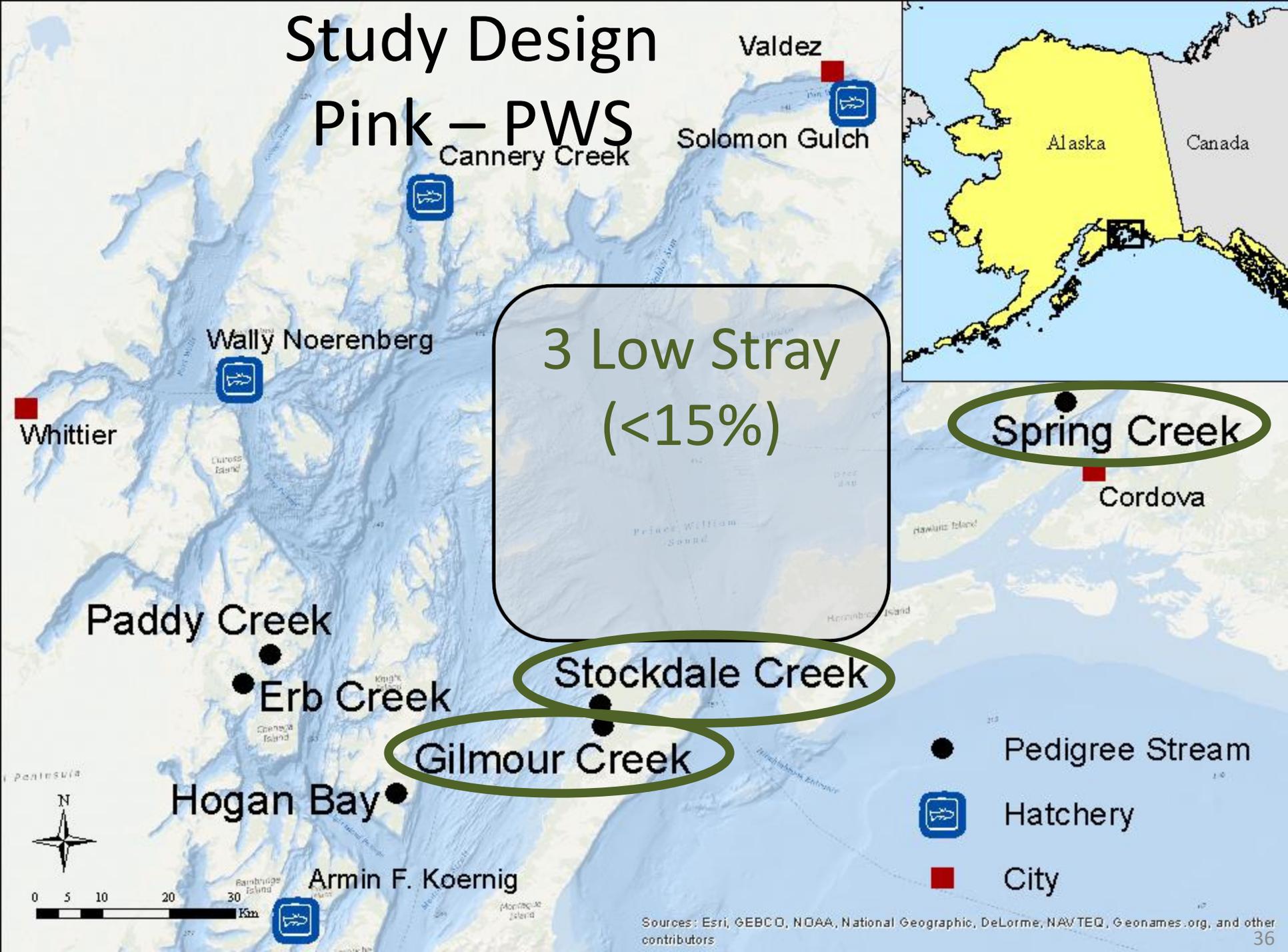
▣ Hatchery

■ City

Sources: Esri, GEBCO, NOAA, National Geographic, DeLorme, NAVTEQ, Geonames.org, and other contributors

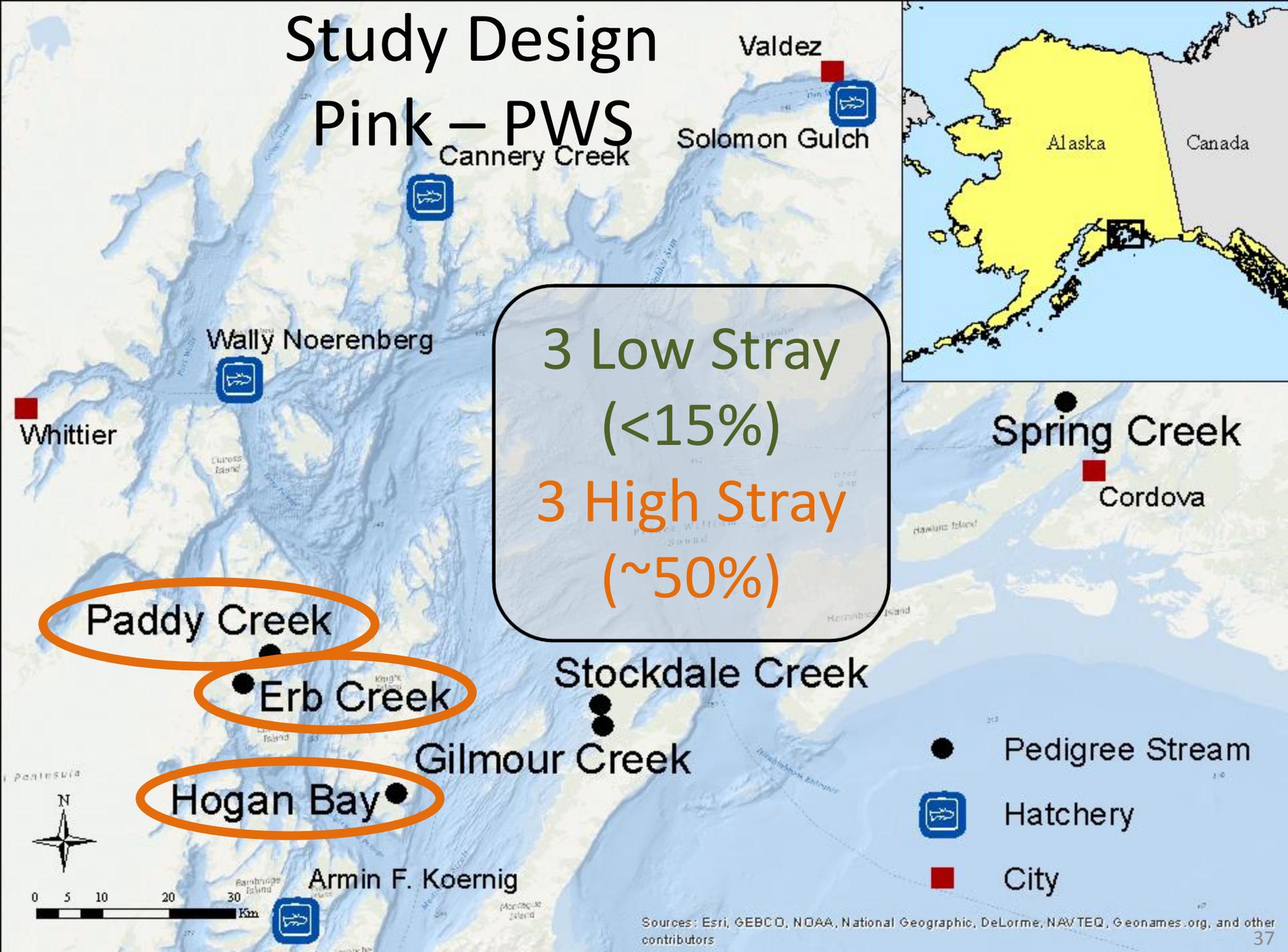
# Study Design

## Pink – PWS



# Study Design

## Pink – PWS



3 Low Stray  
(<15%)

3 High Stray  
(~50%)

# Study Design

## Pink – PWS

Cannery Creek

Valdez

Solomon Gulch

Wally Noerenberg

Whittier

Paddy Creek

Erb Creek

Hogan Bay

Armin F. Koernig

### Selection

- Run size
- Stray rate
- Logistics

Spring Creek

Cordova

Stockdale Creek

Gilmour Creek

● Pedigree Stream



■ Hatchery



■ City



0 5 10 20 30 Km

# Study Design

## Pink – PWS

Cannery Creek

Valdez

Solomon Gulch

Wally Noerenberg

Whittier

Sample  
500-1000/yr

Paddy Creek

Erb Creek

Stockdale Creek

Gilmour Creek

Hogan Bay

Armin F. Koernig

Spring Creek

Cordova

● Pedigree Stream

☐ Hatchery

■ City



0 5 10 20 30 Km

Sources: Esri, GEBCO, NOAA, National Geographic, DeLorme, NAVTEQ, Geonames.org, and other contributors

# Study Design

## Pink – PWS

Cannery Creek

Valdez

Solomon Gulch

Wally Noerenberg

Whittier

Sample  
500-1000/yr  
Escapement  
~3000/yr

Paddy Creek

Erb Creek

Stockdale Creek

Gilmour Creek

Hogan Bay

Armin F. Koernig

Spring Creek

Cordova

● Pedigree Stream



Hatchery



City



0 5 10 20 30 Km



Sources: Esri, GEBCO, NOAA, National Geographic, DeLorme, NAVTEQ, Geonames.org, and other contributors

# Study Design

## Pink – PWS

Cannery Creek

Valdez

Solomon Gulch

Wally Noerenberg

Whittier

Assume

$$RRS_{H/N} \leq 0.5$$

Paddy Creek

Erb Creek

Stockdale Creek

Gilmour Creek

Hogan Bay

Armin F. Koernig

Spring Creek

Cordova

● Pedigree Stream

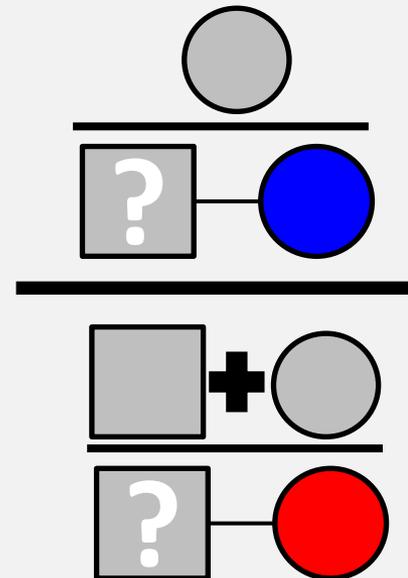
☐ Hatchery

■ City

Sources: Esri, GEBCO, NOAA, National Geographic, DeLorme, NAVTEQ, Geonames.org, and other contributors

# RFP Guidelines

- Sampling
  - High stray
    - 500 adults / 3000 escapement = 1/6
  - Low stray
    - 1000 adults / 3000 escapement = 1/3
- RRS measured by sex



# Considerations

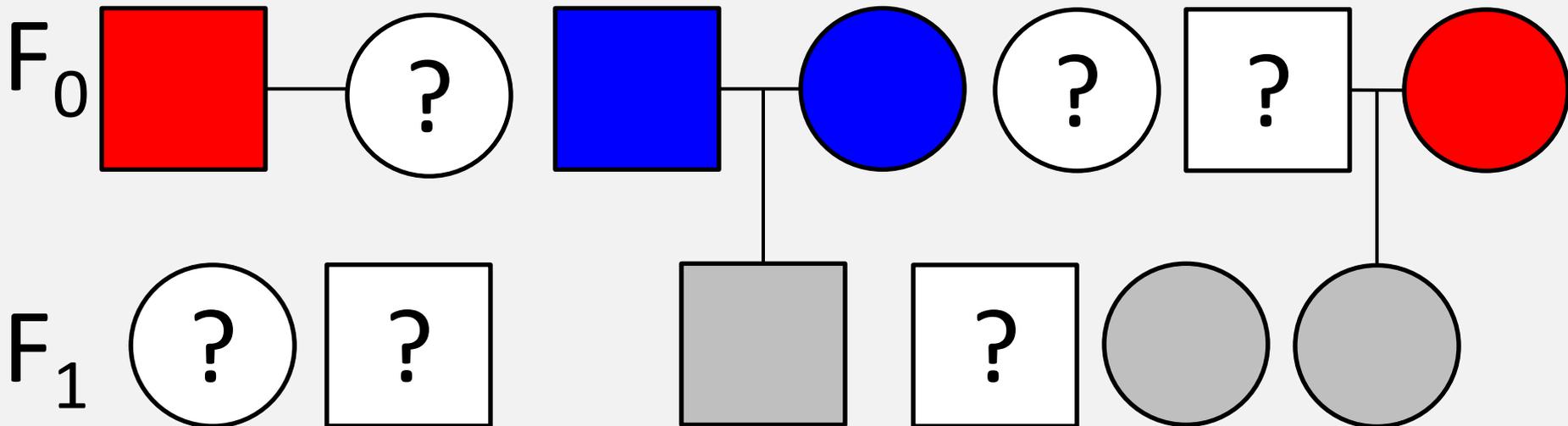
**Missing  
F<sub>0</sub> parents** + **Missing  
F<sub>1</sub> offspring** = **?**

**-Araki and Blouin 2005**

# Considerations

**Missing**  
**F<sub>0</sub> parents** + **Missing**  
**F<sub>1</sub> offspring** = **?**

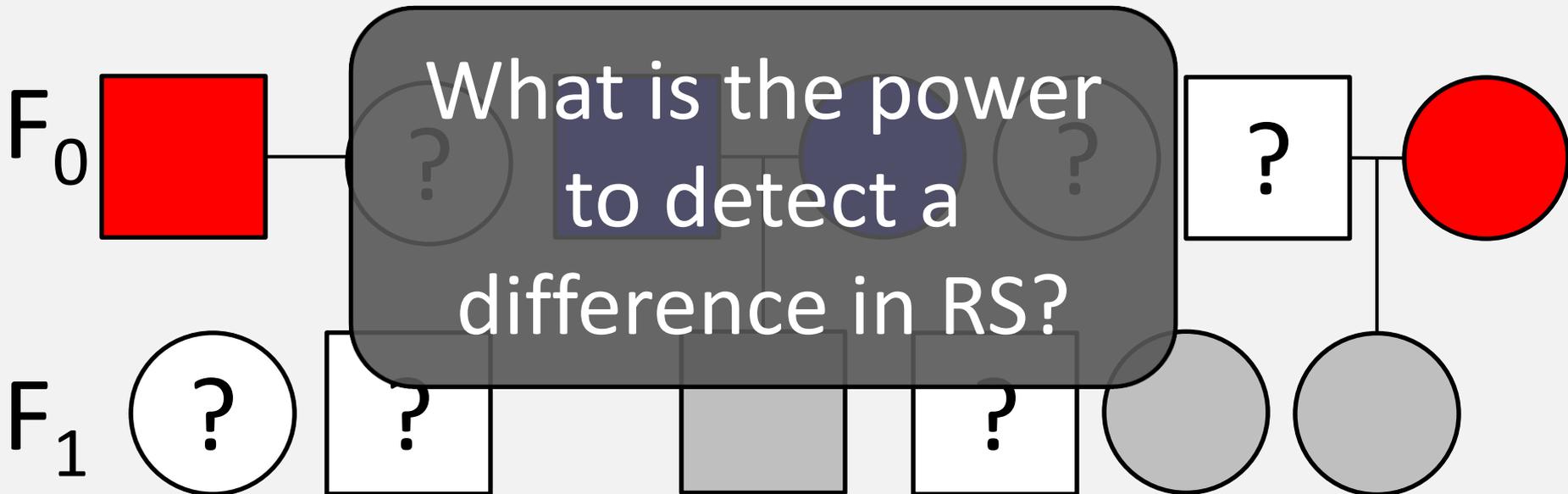
-Araki and Blouin 2005



# Considerations

**Missing**  
**F<sub>0</sub> parents** + **Missing**  
**F<sub>1</sub> offspring** = **?**

-Araki and Blouin 2005



**BREAK: Questions so far?**

# Outline

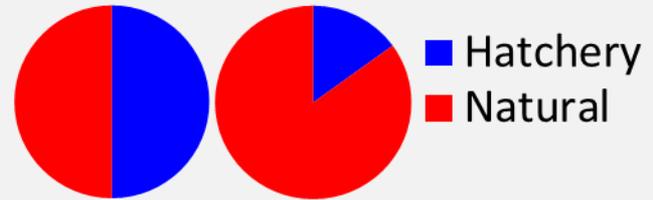
- Background of AHRP
- Parentage and RRS
- Proposed study design
- **Simulations**
- Power analysis
- Christie et al. 2014 review

# Simulation design

- Stray rate

- ❖ % hatchery-origin in stream

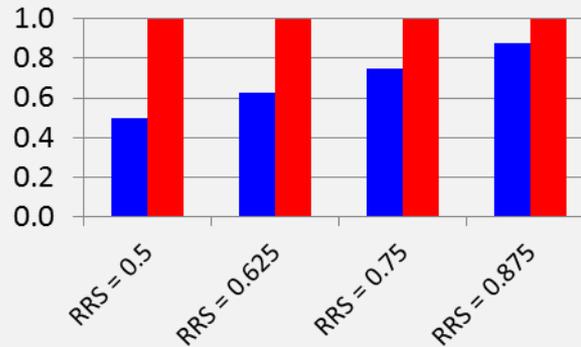
- high (50%), low (15%)



- Effect size

- ❖  $RRS_{H/H \text{ to } N/N}$

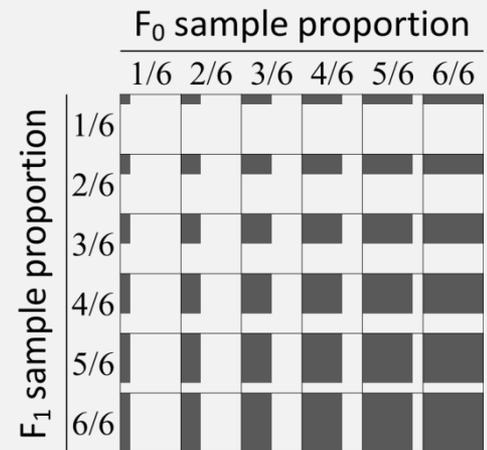
- 0.5 to 0.875



- Proportion of population sampled

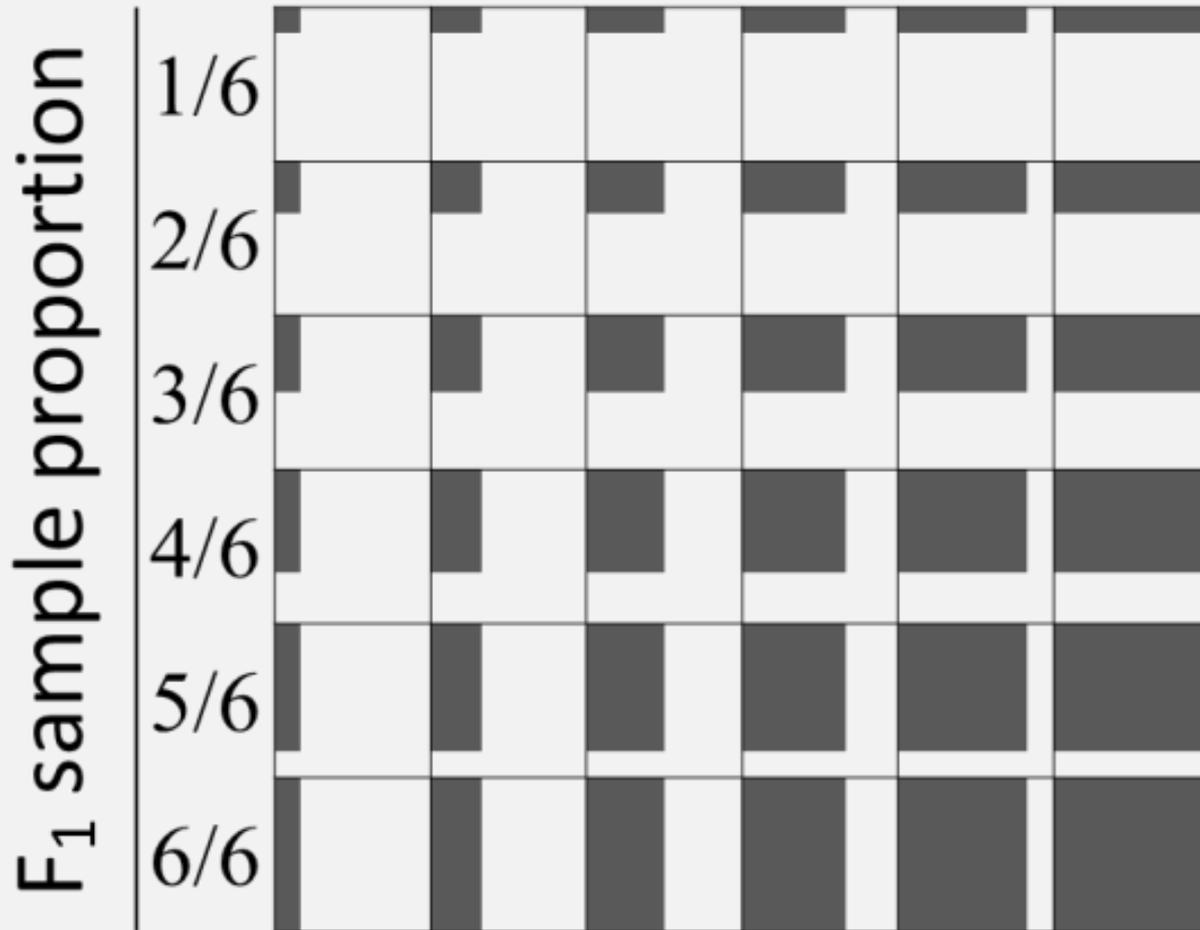
- ❖ Adults and offspring

- 1/6 to 1



# F<sub>0</sub> sample proportion

1/6 2/6 3/6 4/6 5/6 6/6

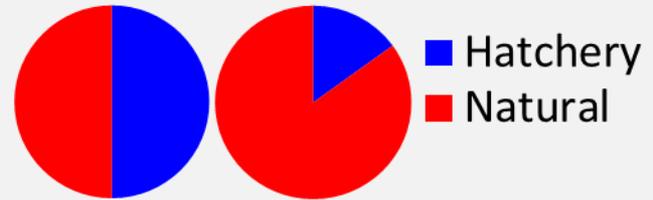


# Simulation design

- Stray rate

- ❖ % hatchery-origin in stream

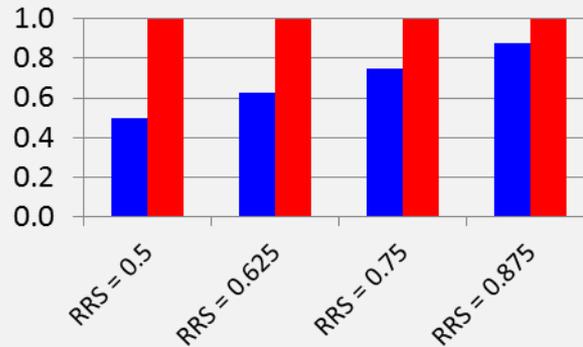
- high (50%), low (15%)



- Effect size

- ❖  $RRS_{H/H \text{ to } N/N}$

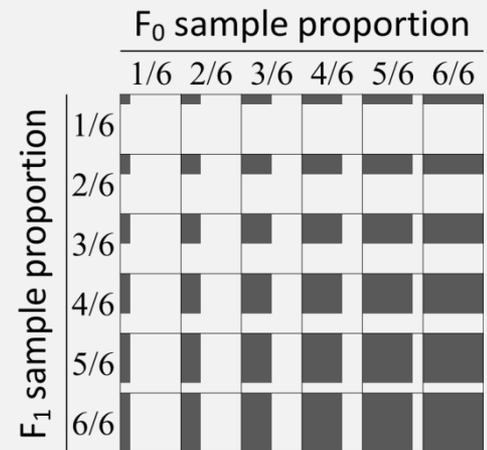
- 0.5 to 0.875



- Proportion of population sampled

- ❖ Adults and offspring

- 1/6 to 1



288 simulated data sets for parentage

**Hatchery**

**Natural**

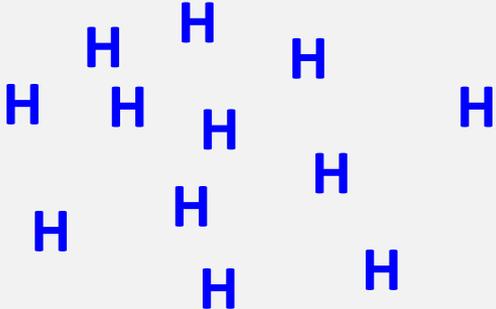
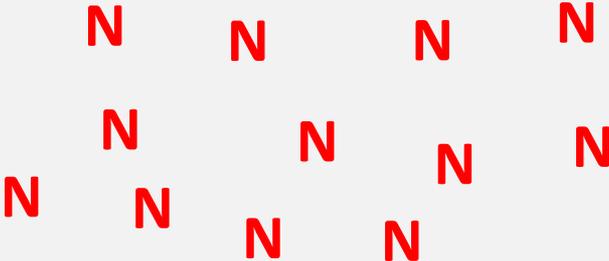
# Simulation design

# Simulation design

Hatchery

Natural

$F_0$



# Simulation design

Hatchery

Natural

F<sub>0</sub>

N N N N  
N N N N  
N N N N

H H H H  
H H H H  
H H H H

Mating

NxN  
NxN

NxN

NxH

NxH

HxN

NxH

HxN

HxN

HxH

HxH

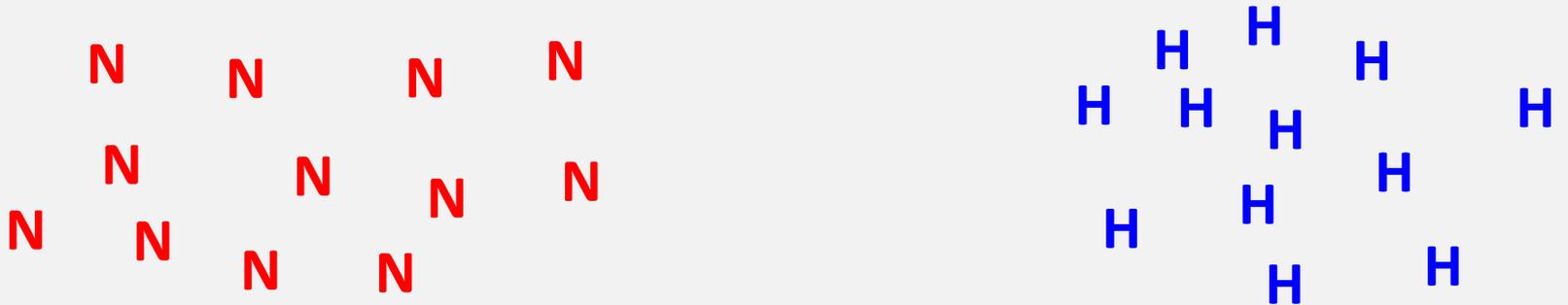
HxH

# Simulation design

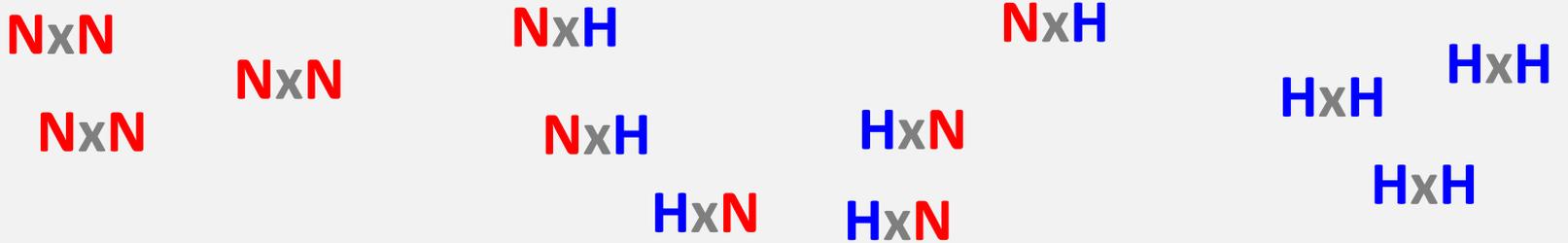
Hatchery

Natural

$F_0$



Mating



#  $F_1$



# Simulation design

Hatchery

Natural

$F_0$

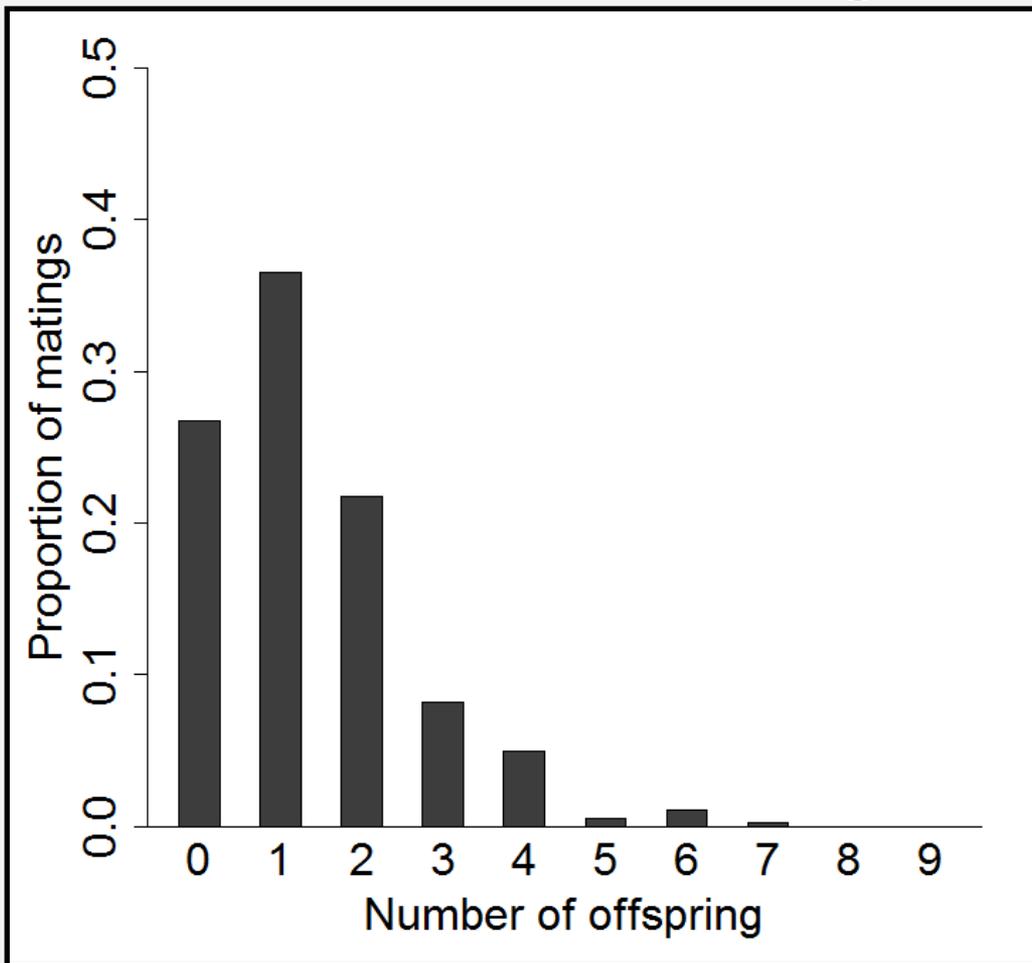
N N N  
N N N

Mating

NxN NxN  
NxN

#  $F_1$

2 1 3



1 0 2  
1 2  
4 3  
4 0  
2

H H H H H  
H H H H  
HxH HxH  
HxH

Hatchery

Natural

# Simulation design

$F_0$

N N N N  
 N N N N  
 N N N N

H H H H  
 H H H H  
 H H H H

Mating

NxN  
 NxN

NxN

NxH

NxH

HxN

NxH

HxN

HxN

HxH

HxH

HxH

#  $F_1$

2

3

1

0

1

4



3

2

2



4



2

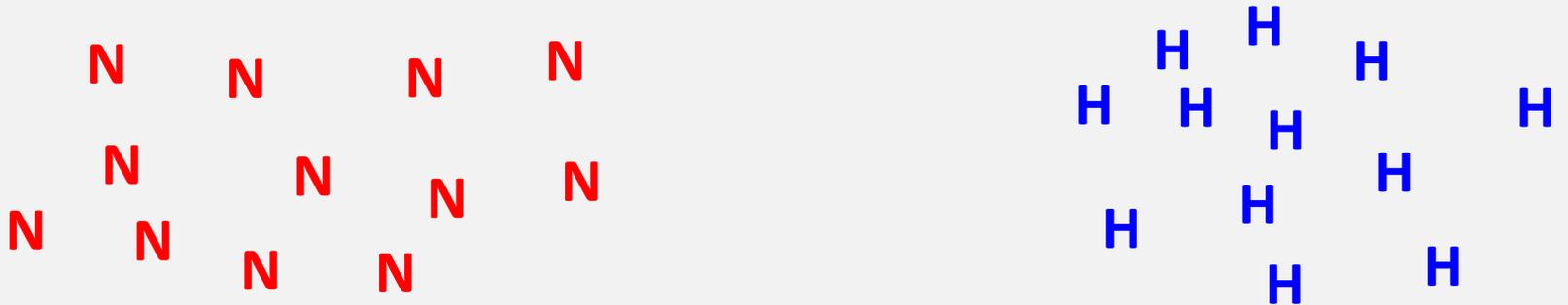
0

# Simulation design

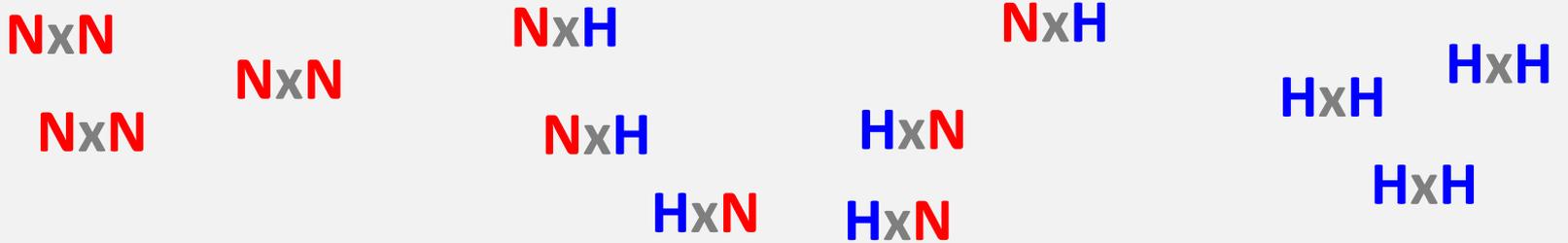
Hatchery

Natural

$F_0$



Mating



#  $F_1$



# Simulation design

Hatchery

Natural

$F_0$

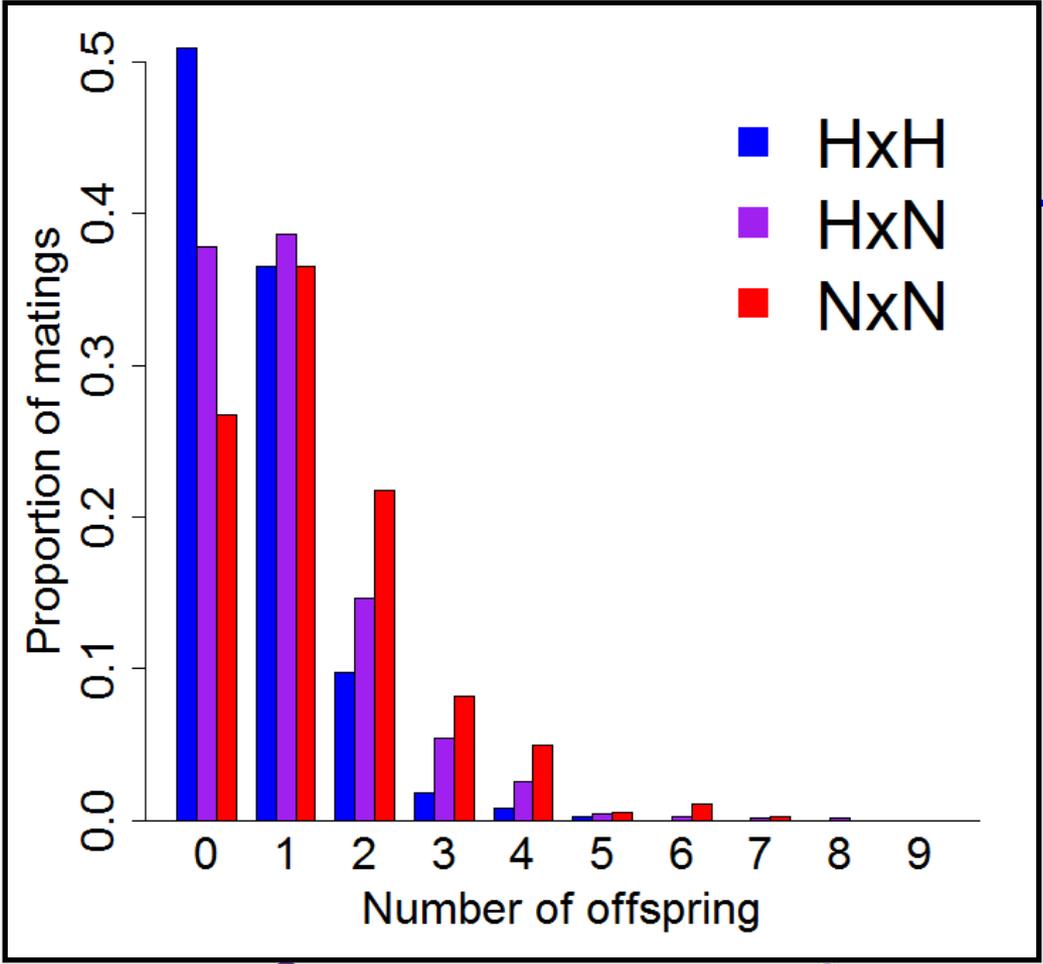
N N N  
N N N

Mating

NxN NxN  
NxN

#  $F_1$

2 1  
3



1

0

1

2

2

2

2

0

1

H H H H H H H H  
H H H H H H H H  
HxH HxH HxH

Hatchery

Natural

# Simulation design

$F_0$

N N N N  
 N N N N  
 N N N N

H H H H  
 H H H H  
 H H H H

Mating

NxN NxN  
 NxN NxN

NxH NxH NxH  
 NxH NxH  
 HxN HxN

NxH HxH HxH  
 HxH HxH

$$RS_{NN} = 2$$

$$RS_{HN} = 1.5$$

$$RS_{HH} = 1$$

#  $F_1$

2 1  
 3

0 2  
 1 2  
 2 2

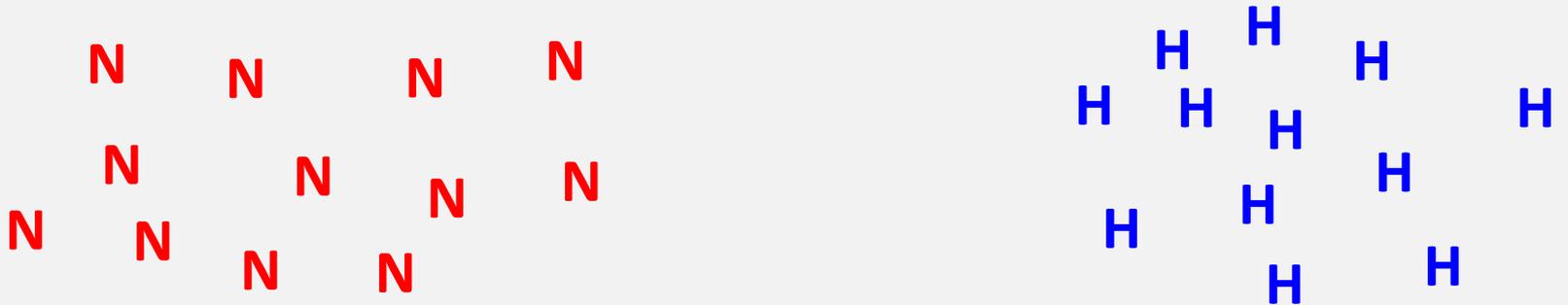
2 0  
 1

# Simulation design

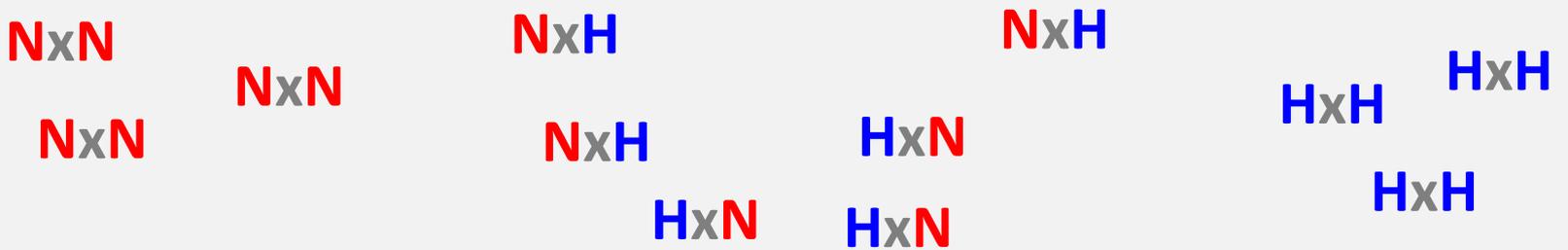
Hatchery

Natural

$F_0$



Mating

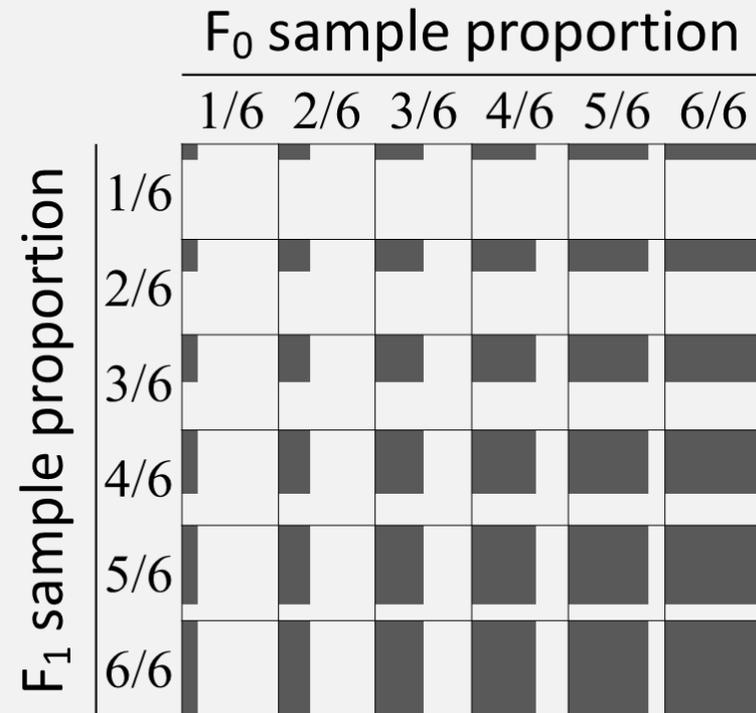


#  $F_1$

	$N \times N$	$H \times N / N \times H$	$H \times H$
RRS	1	0.75	0.5

# Simulation design

- Create  $F_1$  genotypes
- Sample adults
  - 1/6, 2/6, 3/6, 4/6, 5/6, 6/6
- Sample offspring
  - 1/6, 2/6, 3/6, 4/6, 5/6, 6/6



- Incorporate random 0.5% genotype error rate

# Outline

- Background of AHRP
- Parentage and RRS
- Proposed study design
- Simulations
- **Power analysis**
- Christie et al. 2014 review

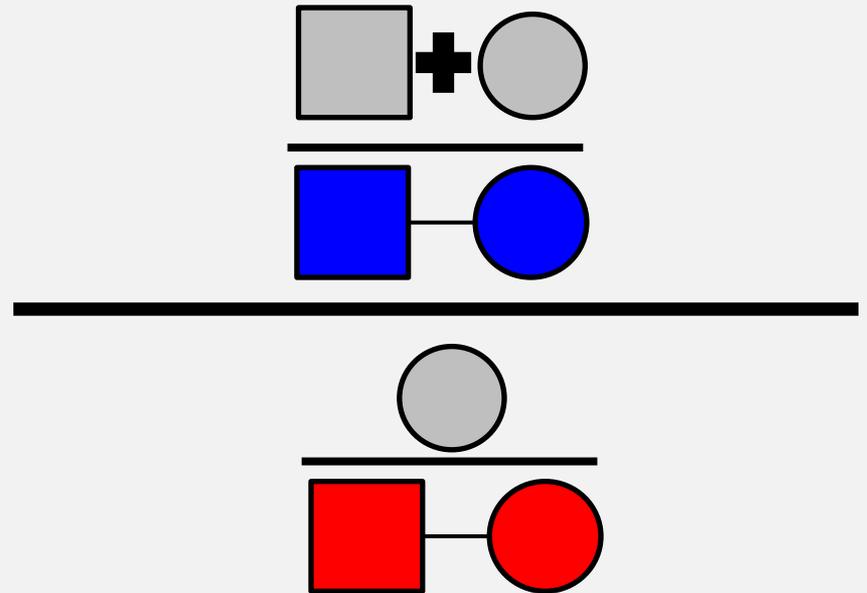
# Relative reproductive success

- Four comparisons
  - By cross type
    - H/H to N/N
    - H/N to N/N

Fitness impact of  
hatchery strays on  
natural fitness

# Relative reproductive success

- Four comparisons
  - By cross type
    - H/H to N/N →
    - H/N to N/N



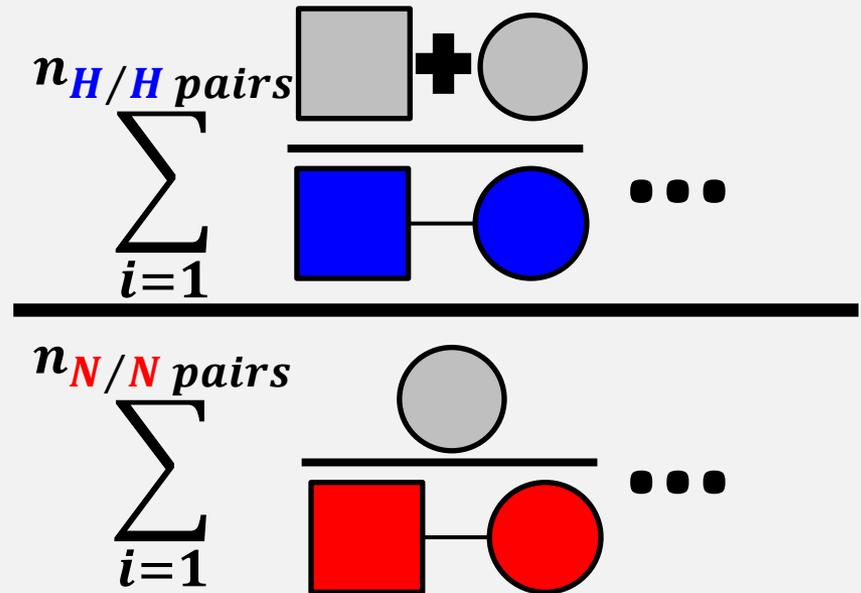
# Relative reproductive success

- Four comparisons

- By cross type

- **H/H** to **N/N** →

- **H/N** to **N/N**



# Relative reproductive success

- Four comparisons

- By cross type

- **H/H** to **N/N** →

- **H/N** to **N/N**

$$\frac{\sum_{i=1}^{n_{H/H} \text{ pairs}} 2; 3; 1; 2 \dots}{\sum_{i=1}^{n_{N/N} \text{ pairs}} 1; 2; 1; 1 \dots}$$

# Relative reproductive success

- Four comparisons

- By cross type

- H/H to N/N →

- H/N to N/N

$$\frac{\sum_{i=1}^{n_{H/H \text{ pairs}}} n_{off i}}{\sum_{i=1}^{n_{N/N \text{ pairs}}} n_{off i}}$$

# Relative reproductive success

- Four comparisons

- By cross type

- **H/H** to **N/N** →

- **H/N** to **N/N**

$$\frac{\sum_{i=1}^{n_{\mathbf{H}/\mathbf{H}} \text{ pairs}} n_{\text{off}_i} / n_{\mathbf{H}/\mathbf{H}} \text{ pairs}}{\sum_{i=1}^{n_{\mathbf{N}/\mathbf{N}} \text{ pairs}} n_{\text{off}_i} / n_{\mathbf{N}/\mathbf{N}} \text{ pairs}}$$

# Relative reproductive success

- Four comparisons

- By cross type

- **H/H** to **N/N**  $\rightarrow RRS_{H/H\text{to}N/N} = 1.3$

- **H/N** to **N/N**

# Relative reproductive success

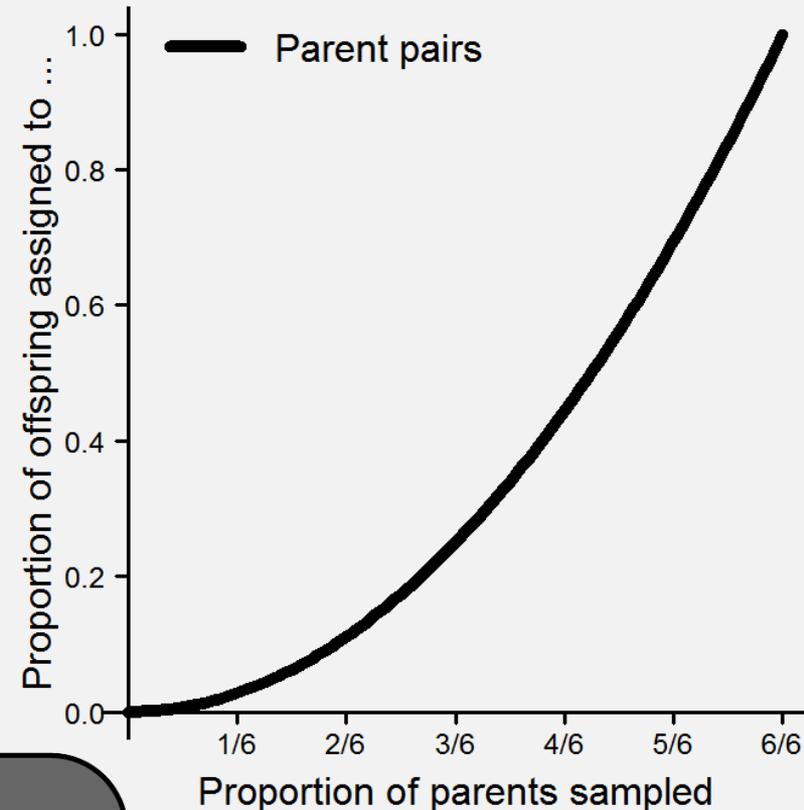
- Four comparisons
  - By cross type
    - H/H to N/N
    - H/N to N/N

## Caveats

- Min # offspring = 1
- Smaller sample size

# Relative reproductive success

- Four comparisons
  - By cross type
    - H/H to N/N
    - H/N to N/N



## Caveats

- Min # offspring = 1
- Smaller sample size

# Relative reproductive success

- Four comparisons
  - By cross type
    - H/H to N/N
    - H/N to N/N
  - Single parent by sex
    - $H_{\text{female}}$  to  $N_{\text{female}}$
    - $H_{\text{male}}$  to  $N_{\text{male}}$

# Relative reproductive success

- Four comparisons

- By cross type

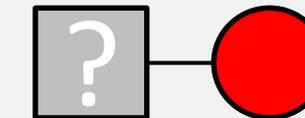
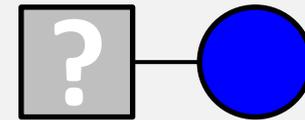
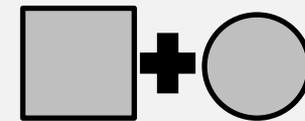
- H/H to N/N

- H/N to N/N

- Single parent by sex

- $H_{\text{female}}$  to  $N_{\text{female}}$  →

- $H_{\text{male}}$  to  $N_{\text{male}}$



# Relative reproductive success

- Four comparisons

- By cross type

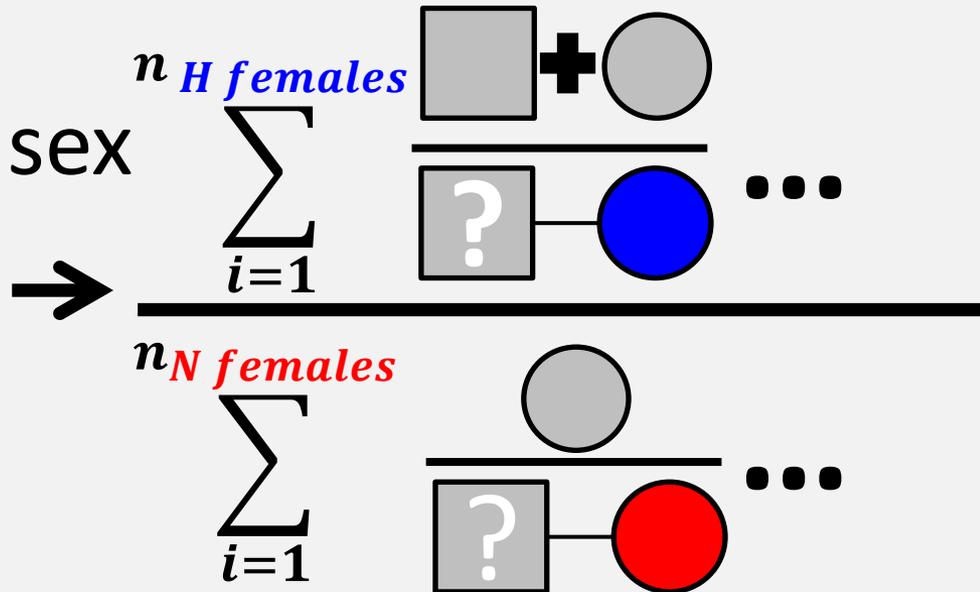
- H/H to N/N

- H/N to N/N

- Single parent by sex

- $H_{\text{female}}$  to  $N_{\text{female}}$

- $H_{\text{male}}$  to  $N_{\text{male}}$



# Relative reproductive success

- Four comparisons

- By cross type

- H/H to N/N

- H/N to N/N

- Single parent by sex

- $H_{\text{female}}$  to  $N_{\text{female}}$

- $H_{\text{male}}$  to  $N_{\text{male}}$

$$\frac{\sum_{i=1}^{n_{H \text{ females}}} 0; 1; 2; 1 \dots}{\sum_{i=1}^{n_{N \text{ females}}} 0; 3; 4; 1 \dots}$$

# Relative reproductive success

- Four comparisons

- By cross type

- H/H to N/N

- H/N to N/N

- Single parent by sex

- $H_{\text{female}}$  to  $N_{\text{female}}$

- $H_{\text{male}}$  to  $N_{\text{male}}$

$$\frac{\sum_{i=1}^{n_{H \text{ females}}} n_{\text{off}_i} / n_{H \text{ females}}}{\sum_{i=1}^{n_{N \text{ females}}} n_{\text{off}_i} / n_{N \text{ females}}}$$

# Relative reproductive success

- Four comparisons

- By cross type

- H/H to N/N

- H/N to N/N

- Single parent by sex

- $H_{\text{female}}$  to  $N_{\text{female}}$  →

- $H_{\text{male}}$  to  $N_{\text{male}}$   $RRS_{H \text{ to } N_{\text{female}}} = 0.5$

# Relative reproductive success

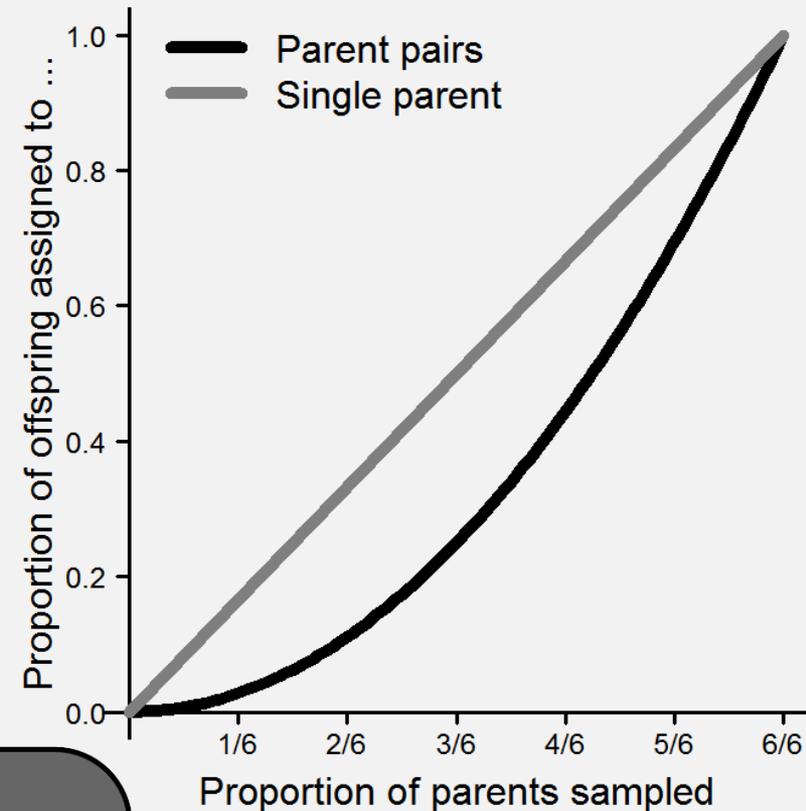
- Four comparisons
  - By cross type
    - H/H to N/N
    - H/N to N/N
  - Single parent by sex

## Caveats

- Min # offspring = 0
- Only  $\frac{1}{2}$  genetic info

# Relative reproductive success

- Four comparisons
  - By cross type
    - H/H to N/N
    - H/N to N/N
  - Single parent by sex



## Caveats

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# Test of fitness differences

- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test

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- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test

Number of offspring	
Hatchery	Natural
1	1
3	2
1	3
0	1
3	5
2	1
0	0
2	2
	2
	3

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  - Permutation test

Number of offspring	
Hatchery	Natural
1	1
3	2
1	3
0	1
3	5
2	1
0	0
2	2
	2
	3
$RS_H = 1.5$	$RS_N = 2.0$

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  - Permutation test

Number of offspring	
Hatchery	Natural
1	1
3	2
1	3
0	1
3	5
2	1
0	0
2	2
	2
	3
$RS_H = 1.5$	$RS_N = 2.0$
$RRS_{H/N} = 0.75$	

# Test of fitness differences

- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test

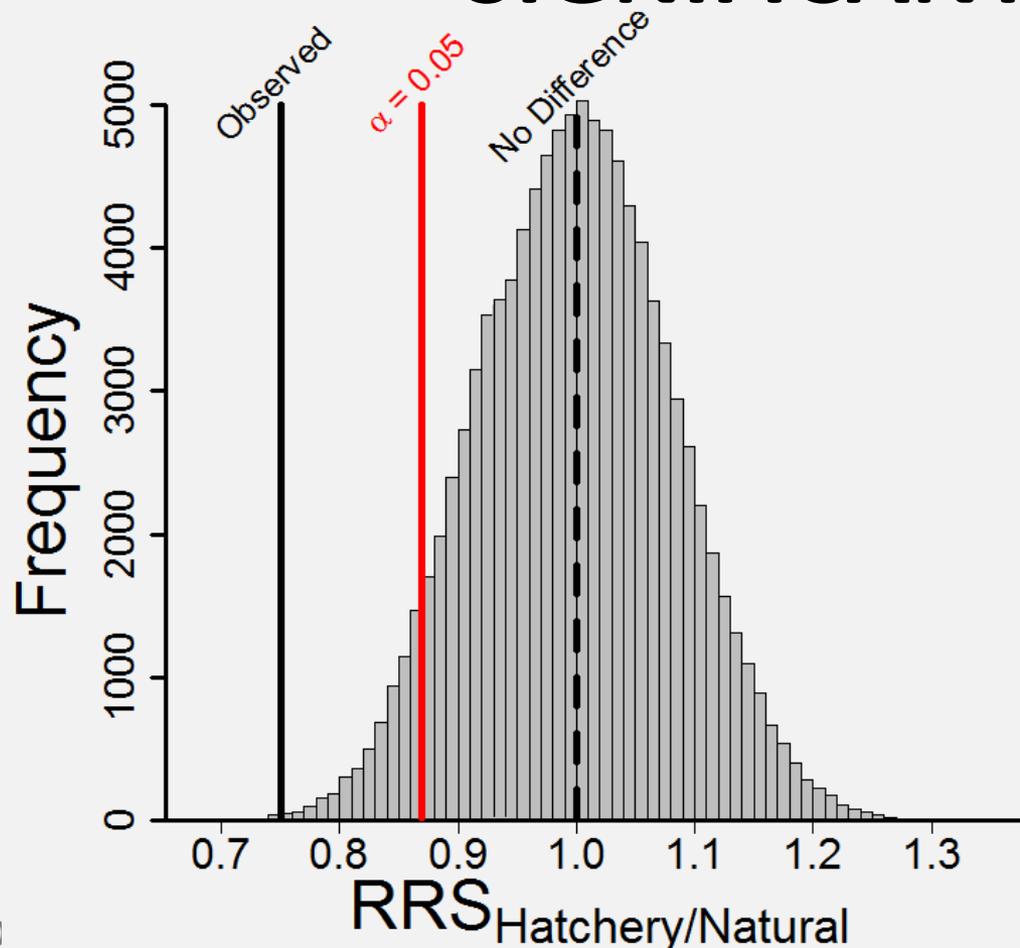
**REPEAT  
100,000  
TIMES**

Number of offspring	
Hatchery	Natural
1	3
1	2
0	0
3	1
3	2
2	1
0	1
5	2
	2
	3
$RS_H = 1.9$	$RS_N = 1.7$
$RRS_{H/N} = 1.10$	

# Test of fitness differences

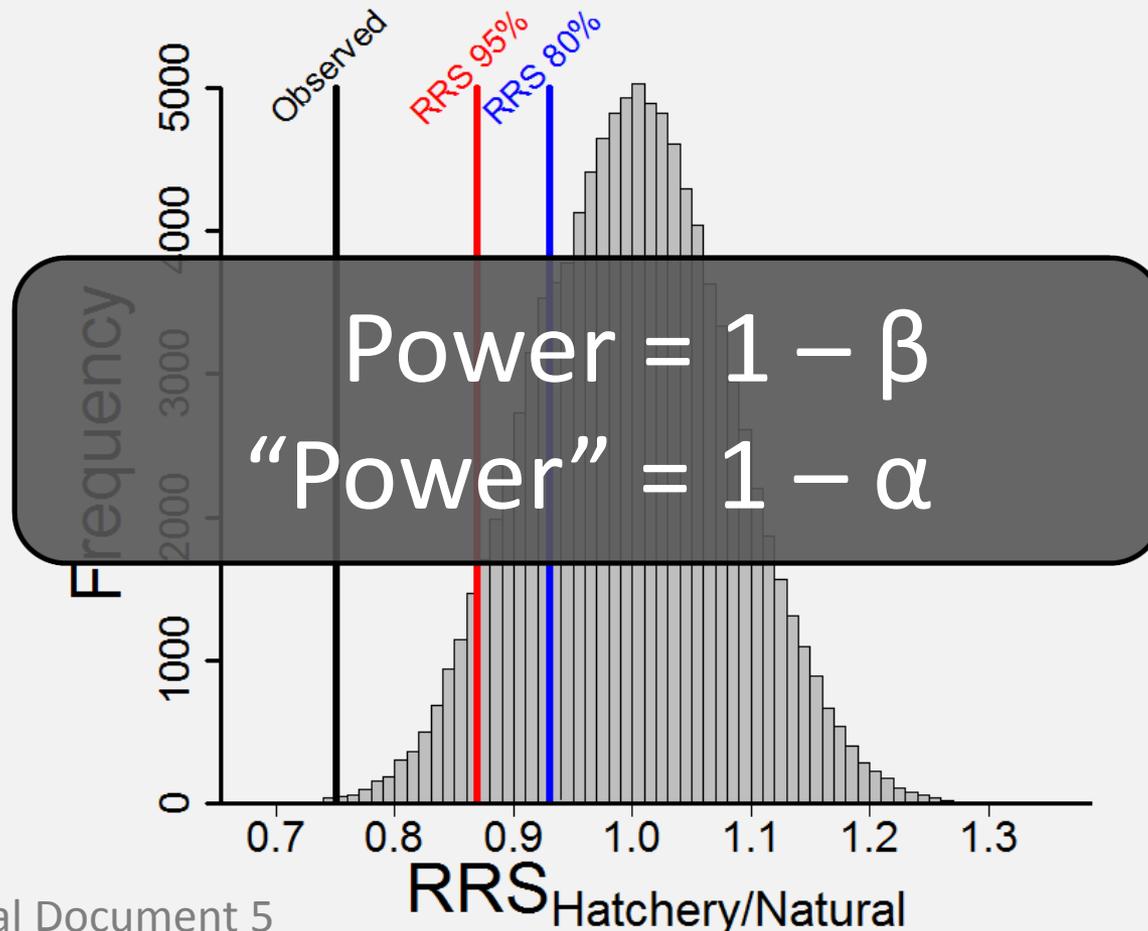
- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test

**SIGNIFICANT**



# Test of fitness differences

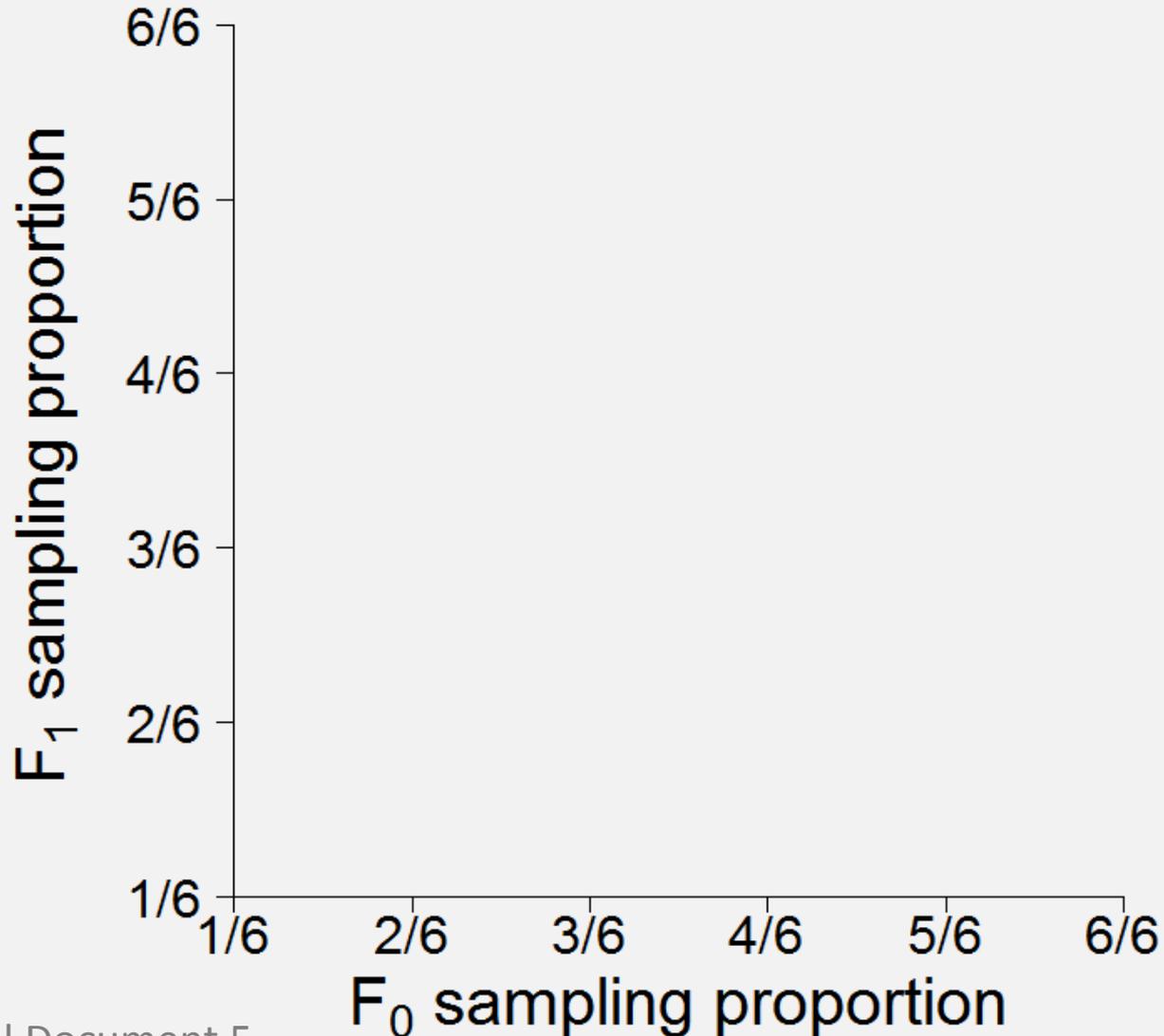
- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test
    - "Power" - 80% & 95% (Thériault et al. 2011 & Hess et al. 2012)



# Single parent results: Permutation test

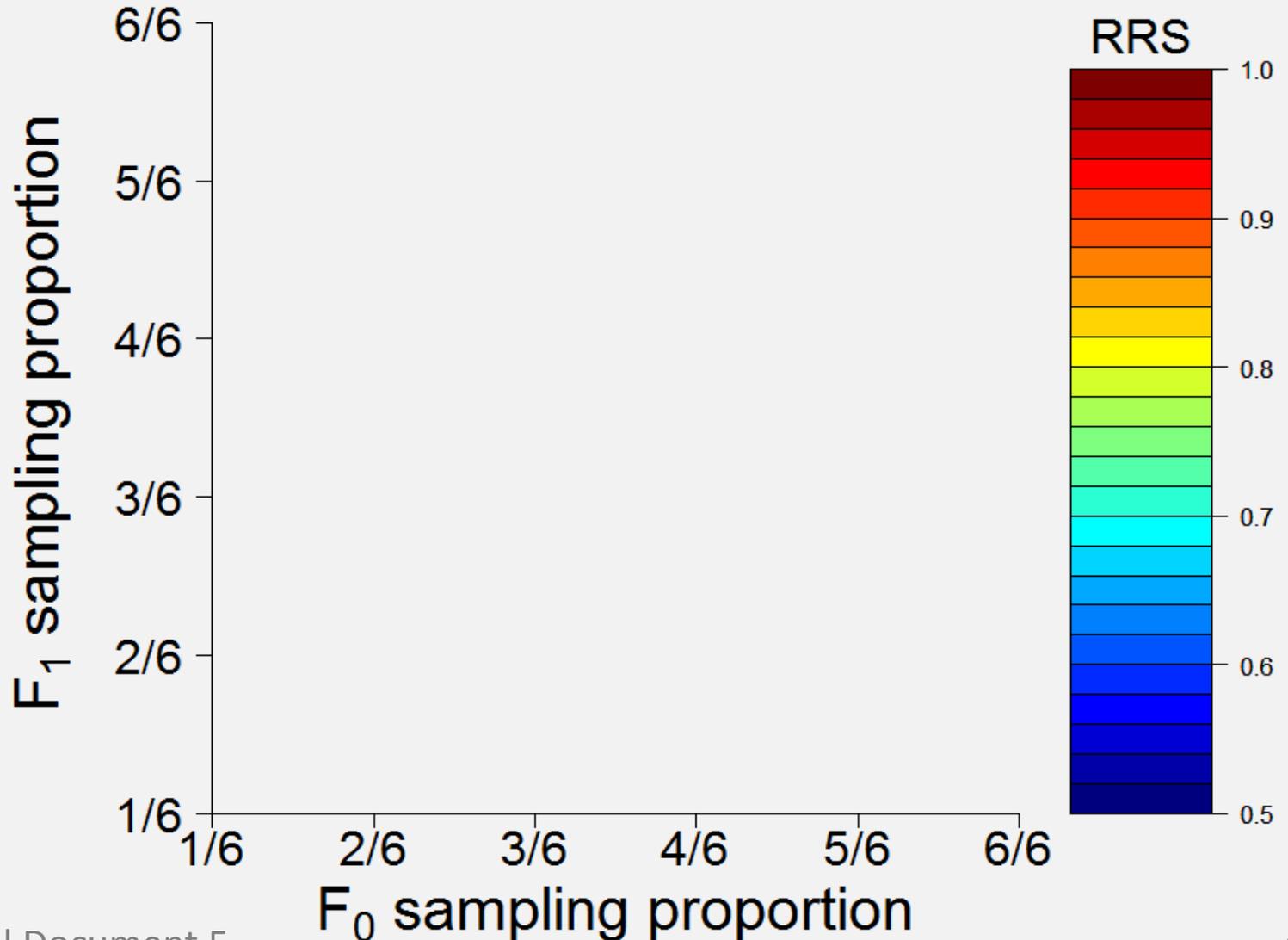
# RRS 95% "Power"

Modeled  $RRS_{HH/NN} = 0.50$



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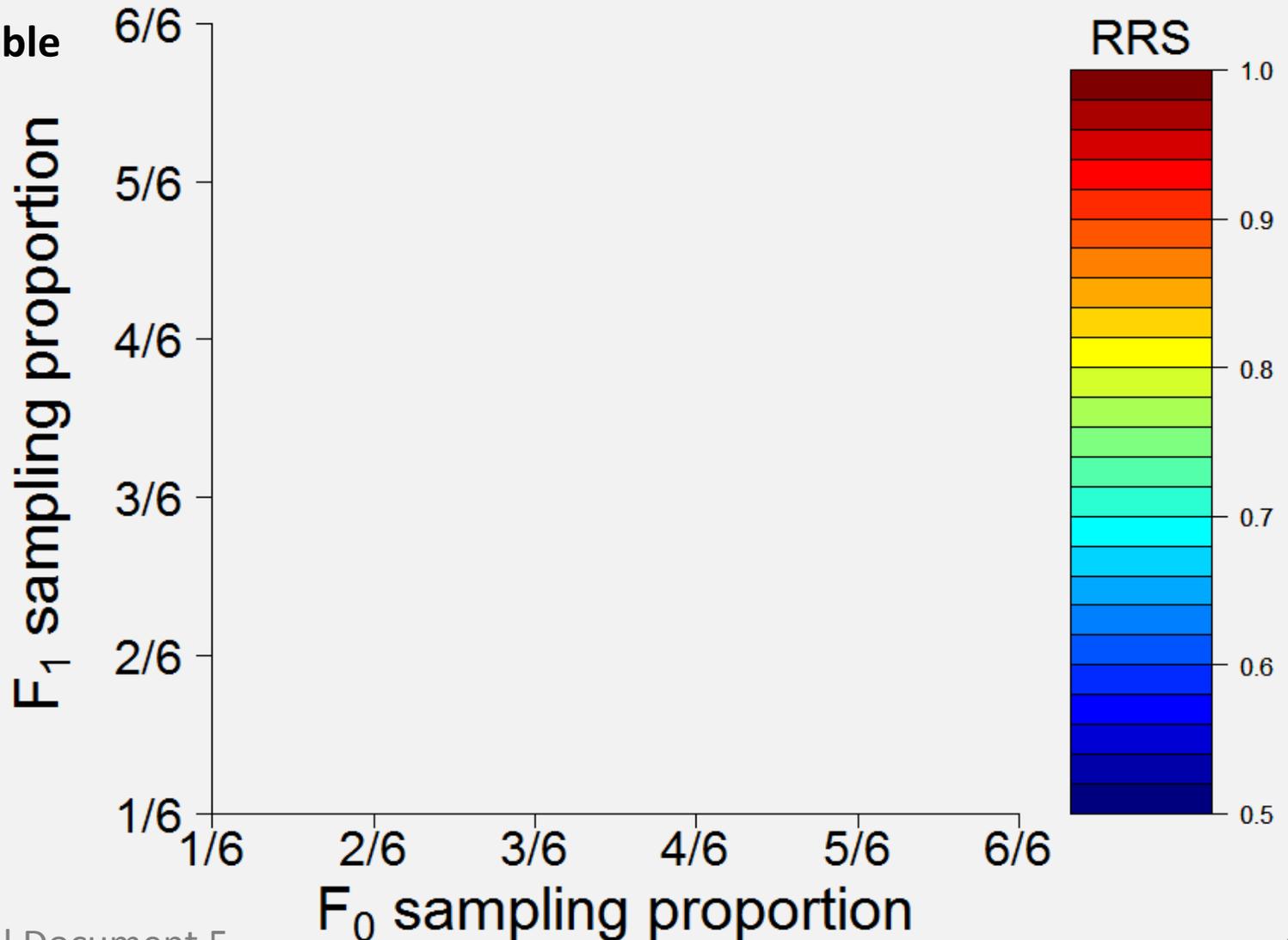


# RRS 95% "Power"

Modeled  $RRS_{HH/NN} = 0.50$

✦ Detectable

✦ Undetectable

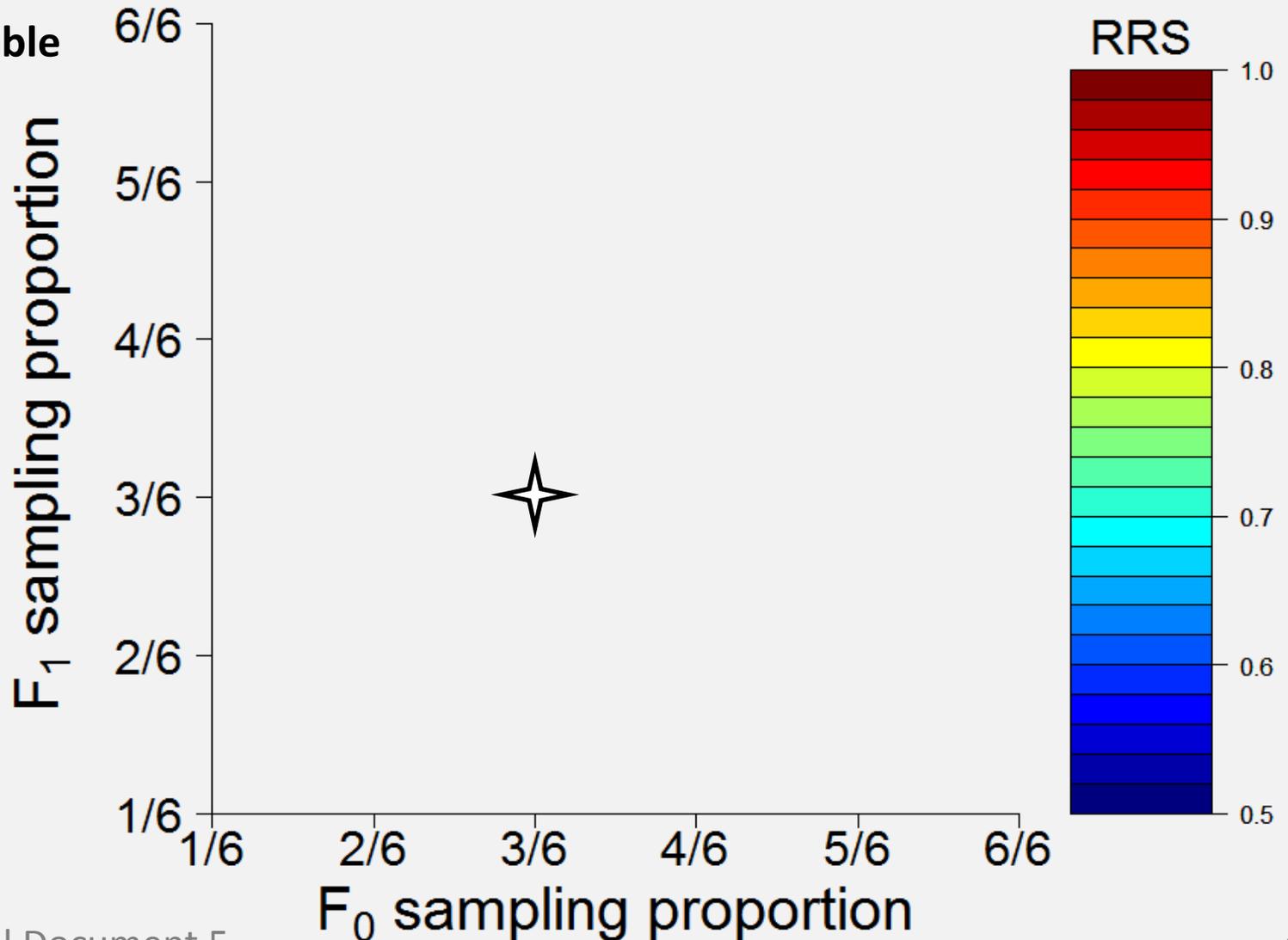


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 Detectable

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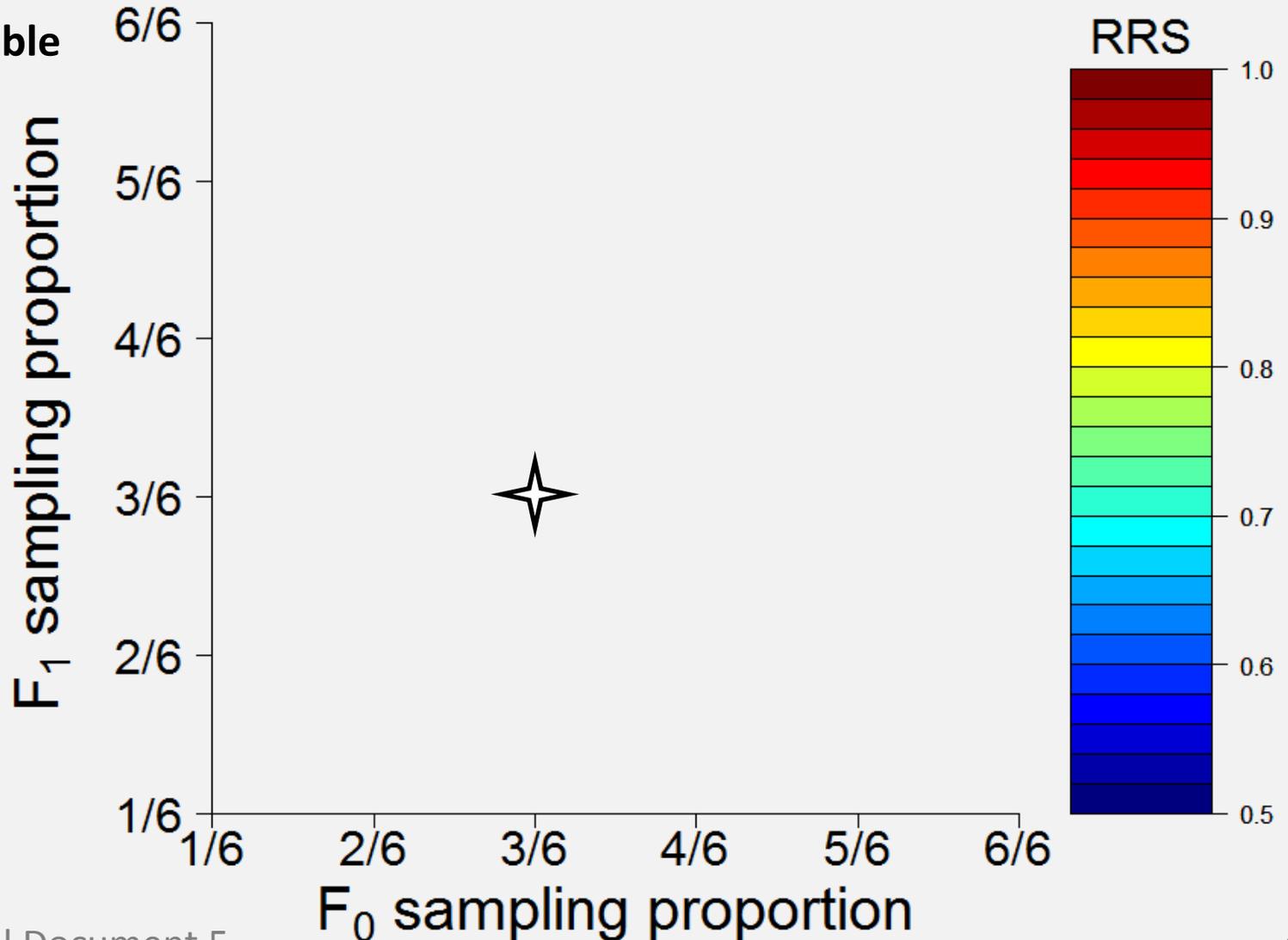


# RRS 95% "Power"

✦ Detectable

✦ Undetectable

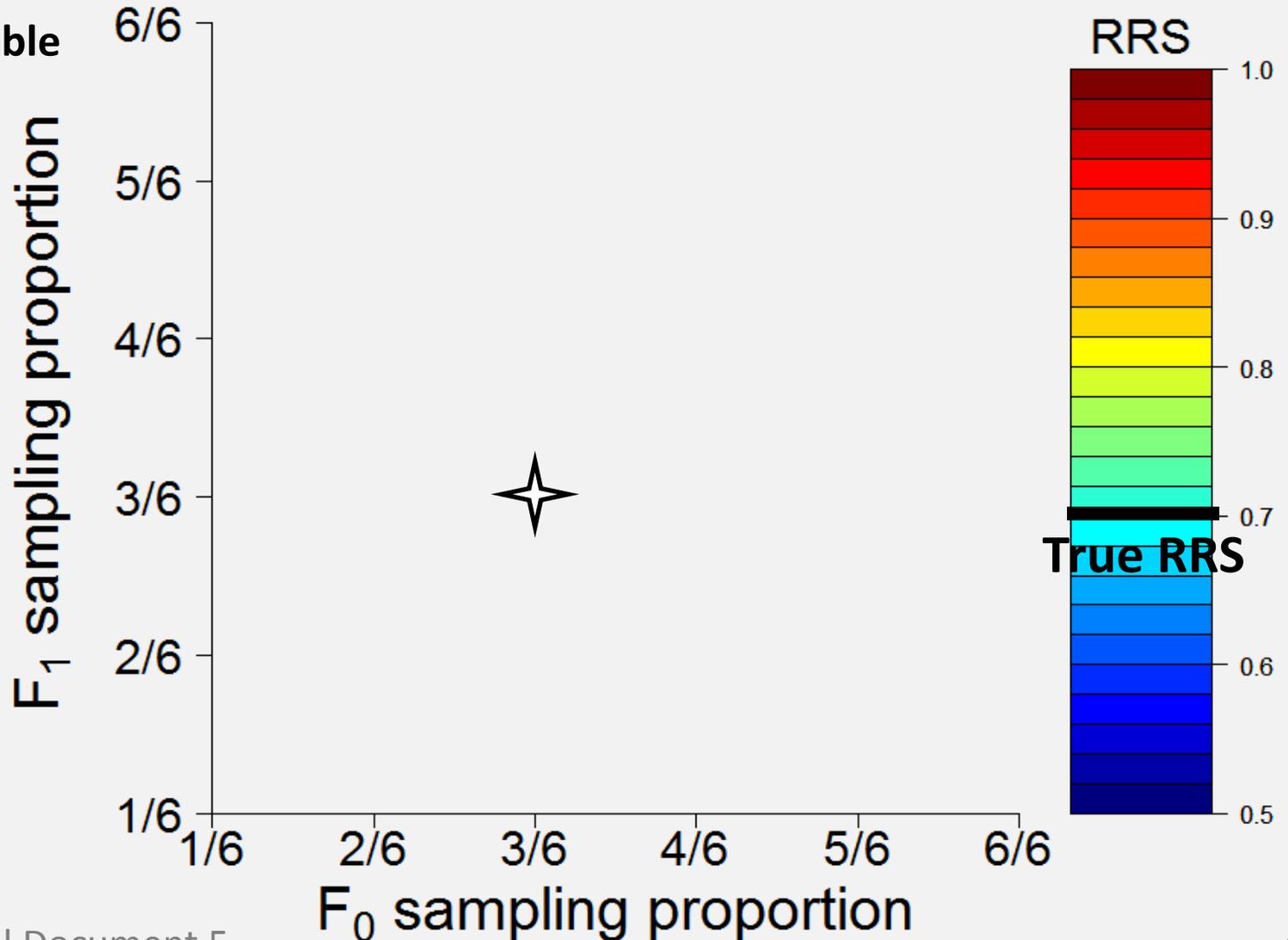
Modeled  $RRS_{HH/NN} = 0.50$ ;  $RRS_{H/N} = 0.70$



# RRS 95% "Power"

✦ Detectable  
✦ Undetectable

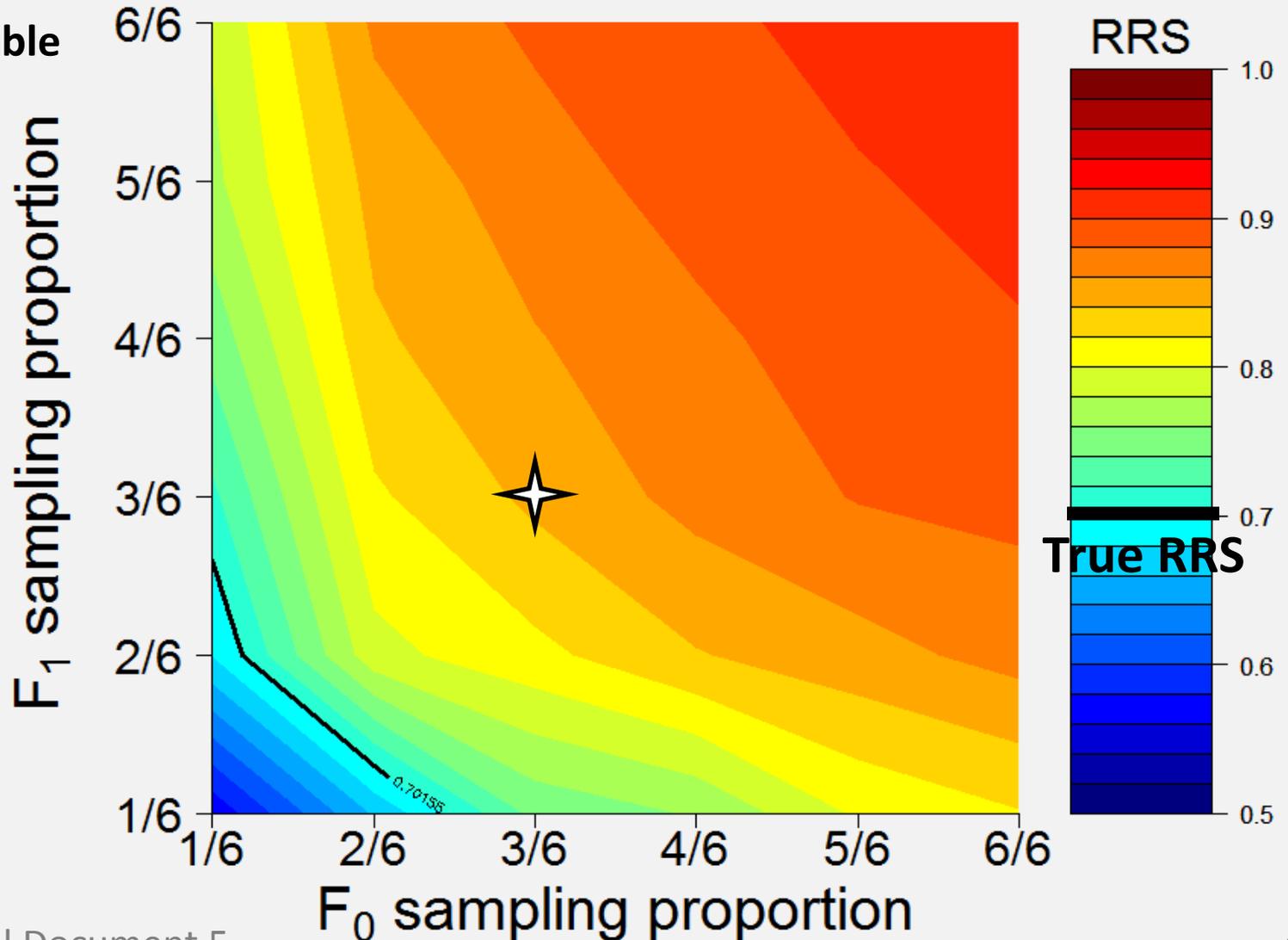
Modeled  $RRS_{HH/NN} = 0.50$ ;  $RRS_{H/N} = 0.70$



# RRS 95% "Power"

- ✦ Detectable
- ✦ Undetectable

Modeled  $RRS_{HH/NN} = 0.50$ ;  $RRS_{H/N} = 0.70$



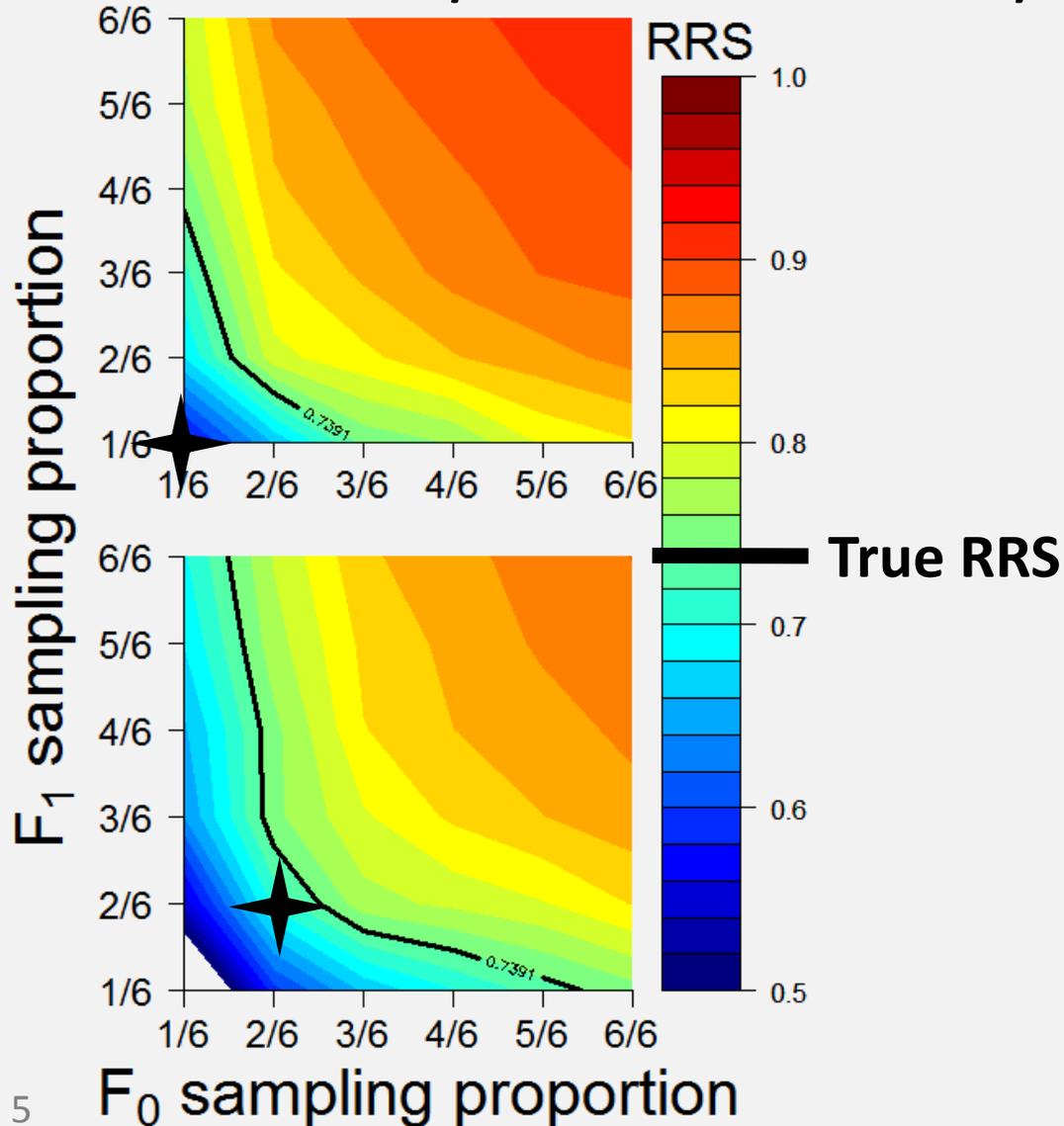
# RRS 95% "Power"

Modeled RRS<sub>HH/NN</sub> = 0.50; RRS<sub>H/N</sub> = 0.74



High  
Stray

Low  
Stray

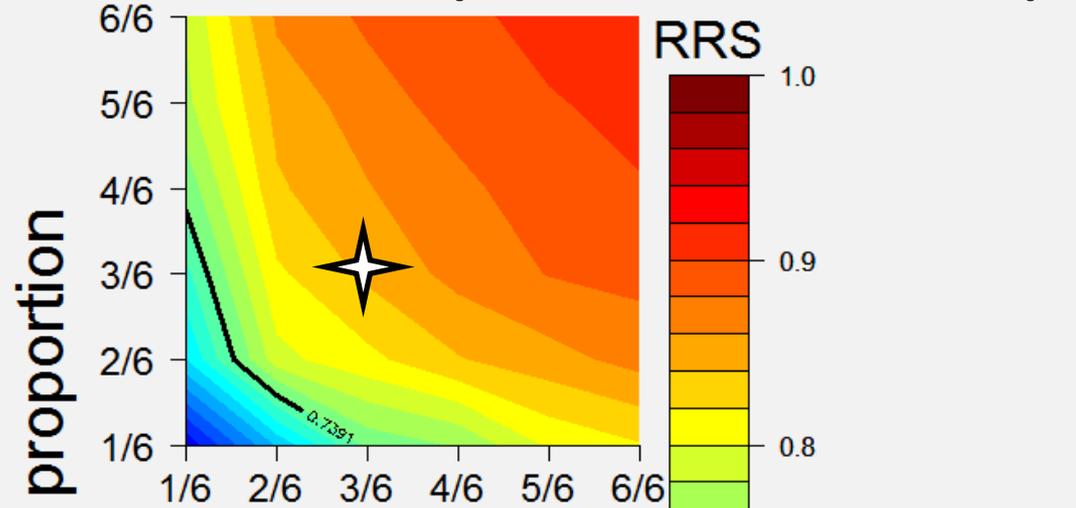


# RRS 95% "Power"

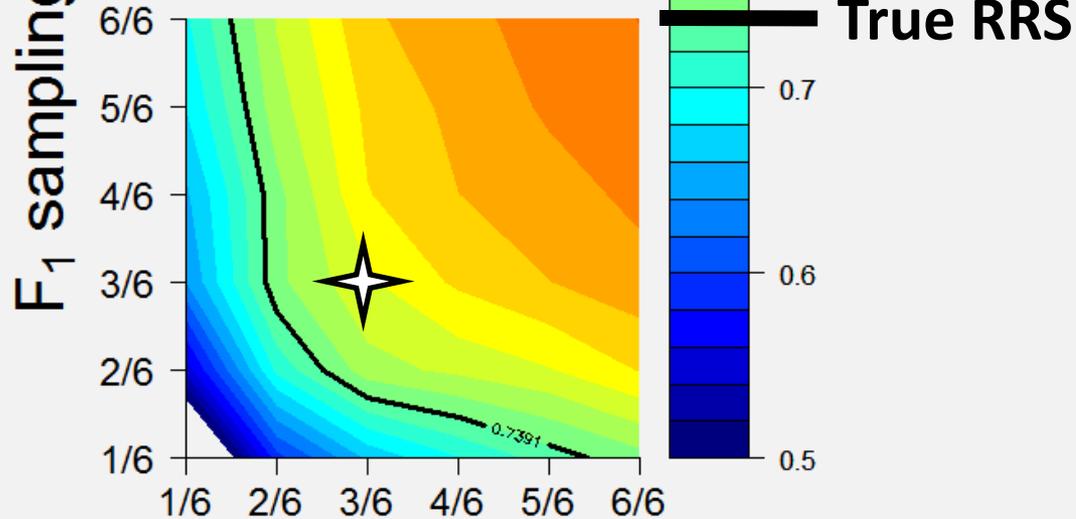
Modeled RRS<sub>HH/NN</sub> = 0.50; RRS<sub>H/N</sub> = 0.74



High  
Stray



Low  
Stray



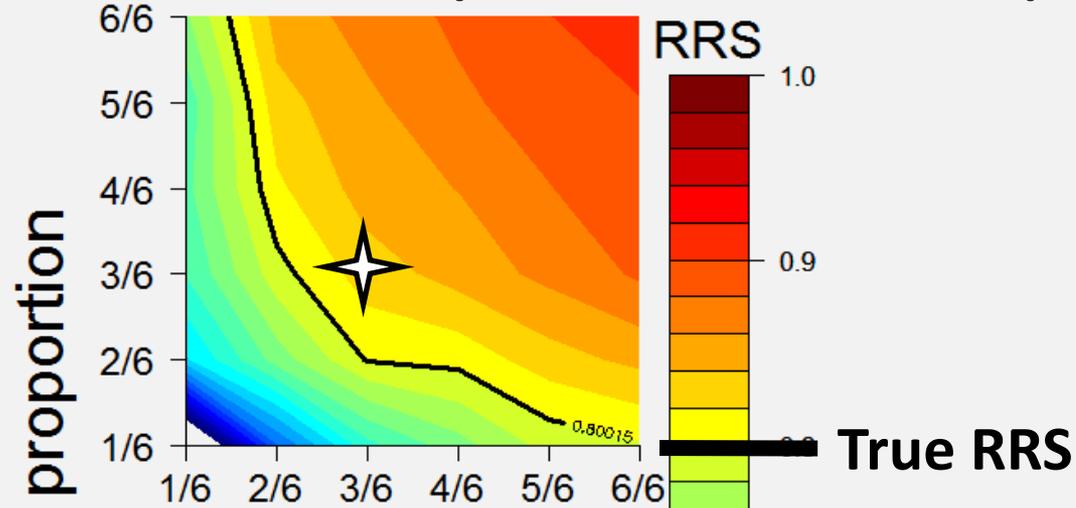
F<sub>0</sub> sampling proportion

# RRS 95% "Power"

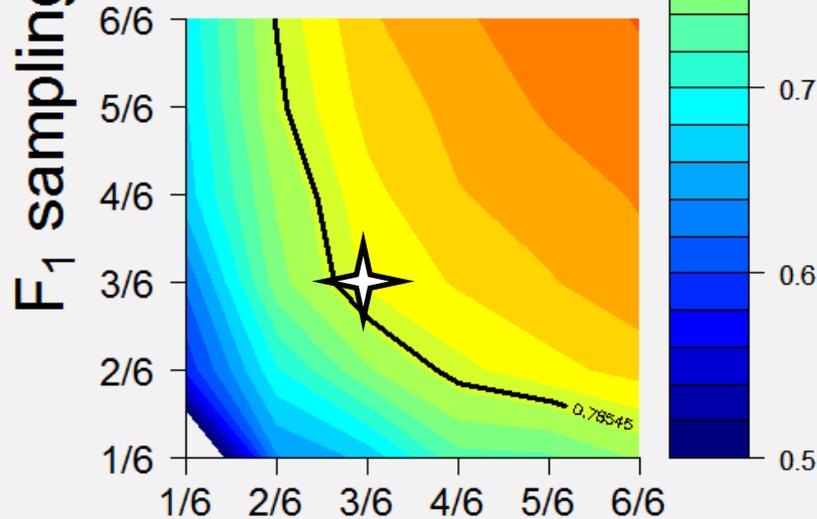
Modeled RRS<sub>HH/NN</sub> = 0.63; RRS<sub>H/N</sub> = 0.79



**High  
Stray**



**Low  
Stray**



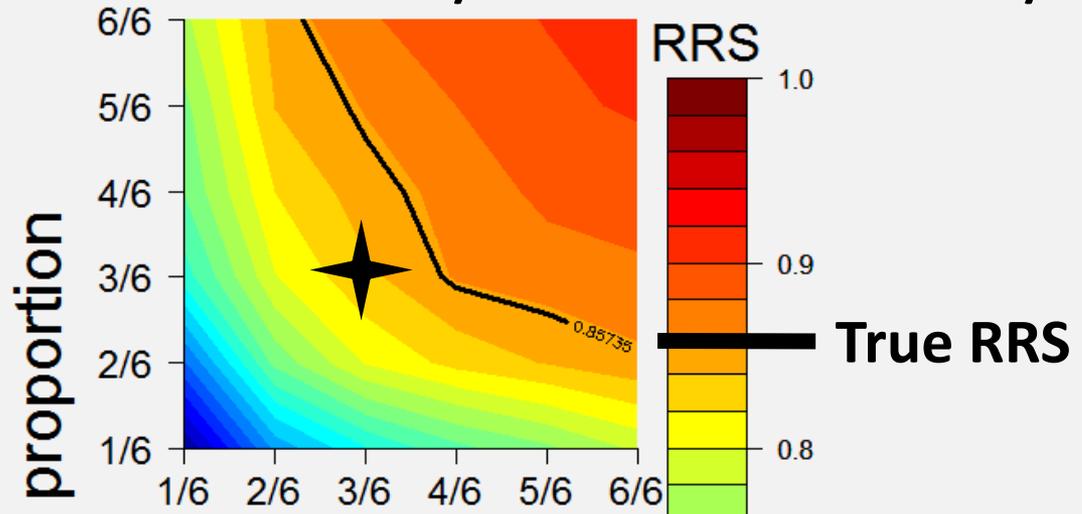
$F_0$  sampling proportion

# RRS 95% "Power"

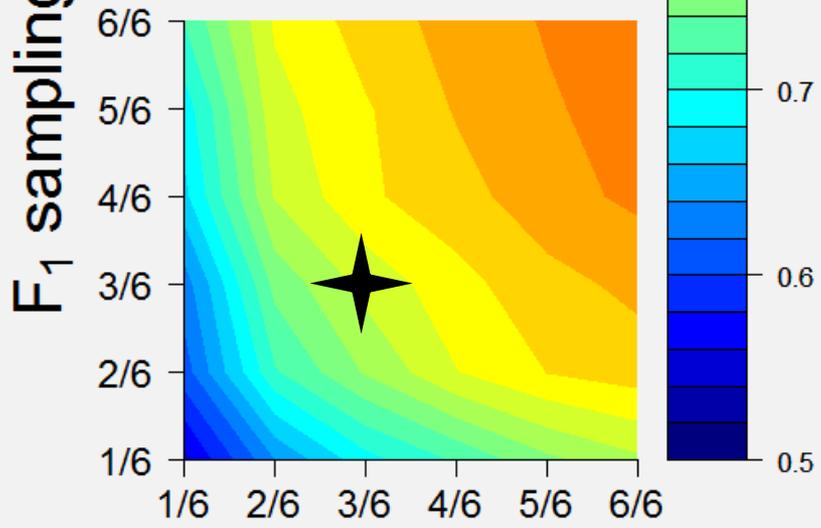
Modeled RRS<sub>HH/NN</sub> = 0.75; RRS<sub>H/N</sub> = 0.86

 Detectable  
 Undetectable

**High  
Stray**



**Low  
Stray**



$F_0$  sampling proportion

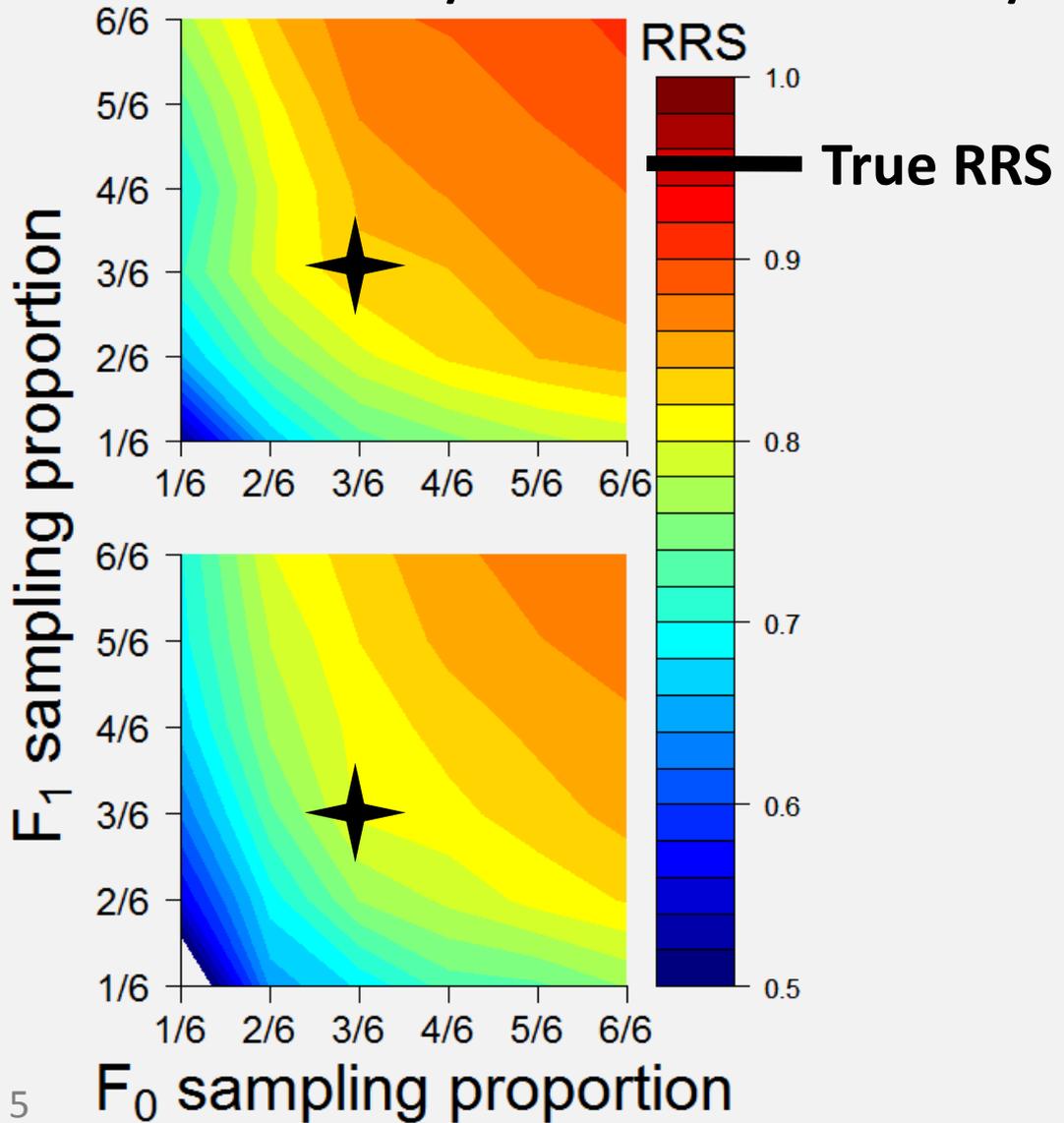
# RRS 95% "Power"

Modeled  $RRS_{HH/NN} = 0.88$ ;  $RRS_{H/N} = 0.95$

★ Detectable  
★ Undetectable

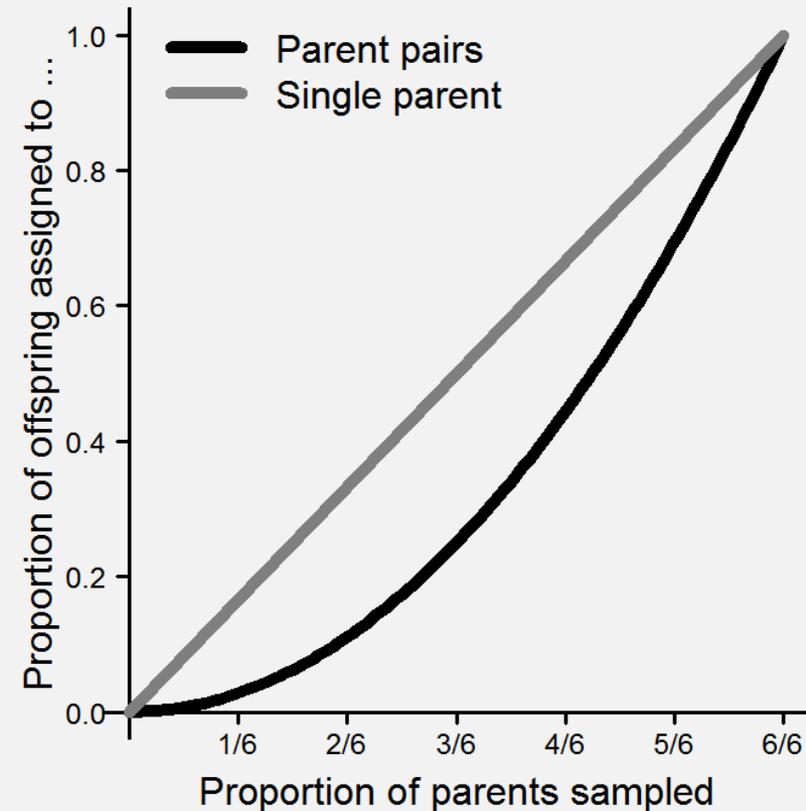
High Stray

Low Stray



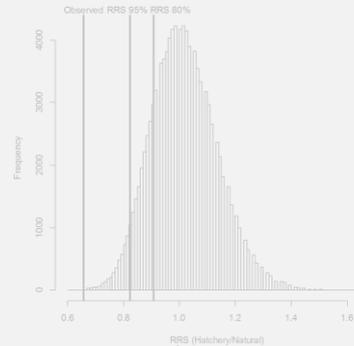
# RRS: Single Parents by Sex

- Described in RFP
- Larger sample size
- Greater power



# Test of fitness differences

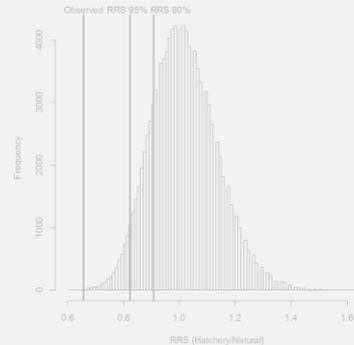
- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test
    - "Power" - 80% & 95% (Thériault et al. 2011 & Hess et al. 2012)



- Parametric approach (Anderson 2013)
  - Negative binomial general linear model (GLM)

# Test of fitness differences

- Non-parametric approach (Araki & Blouin 2005)
  - Permutation test
    - "Power" - 80% & 95% (Thériault et al. 2011 & Hess et al. 2012)

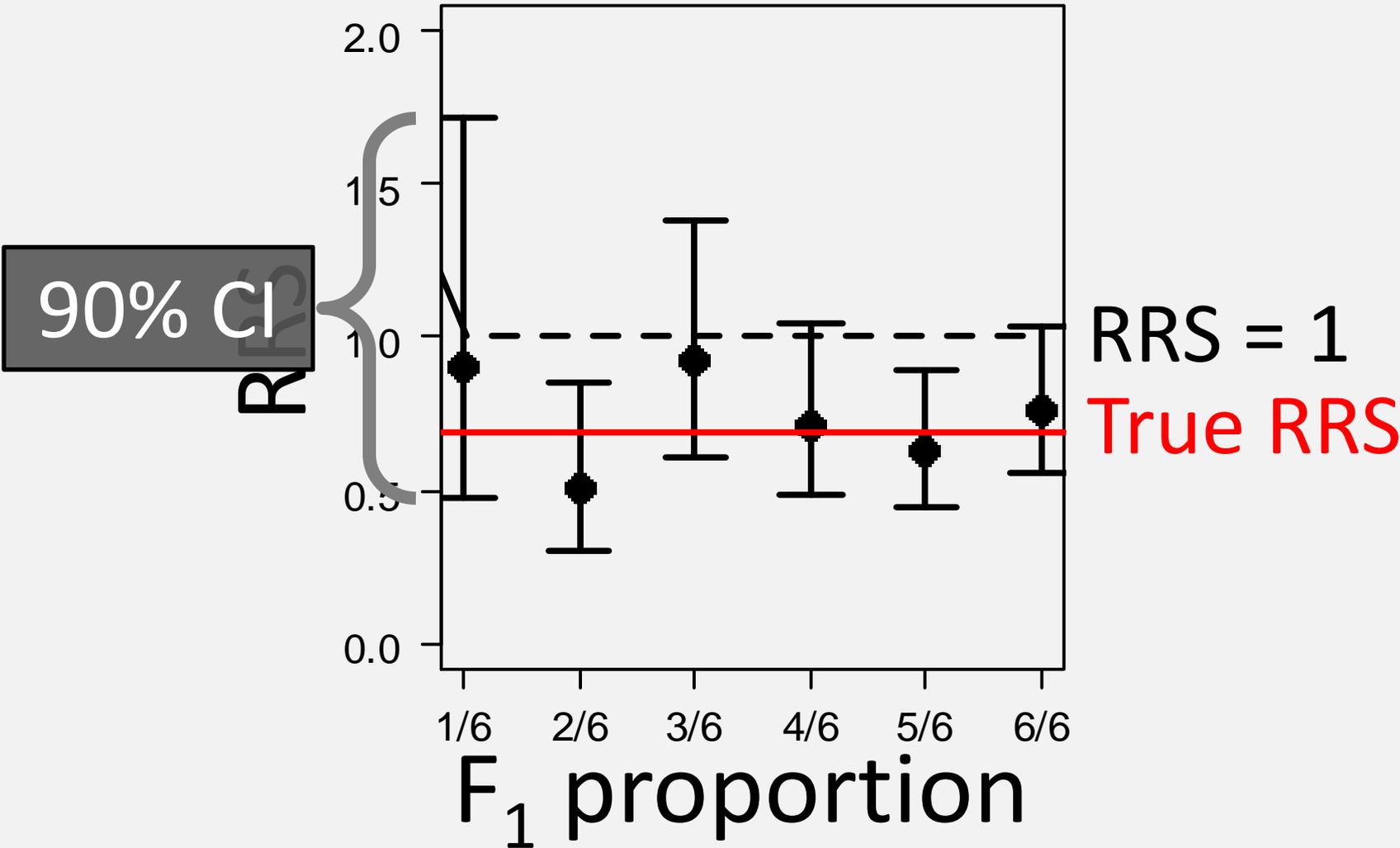


- Parametric approach (Anderson 2013)
  - Negative binomial general linear model (GLM)
    - 90% CI

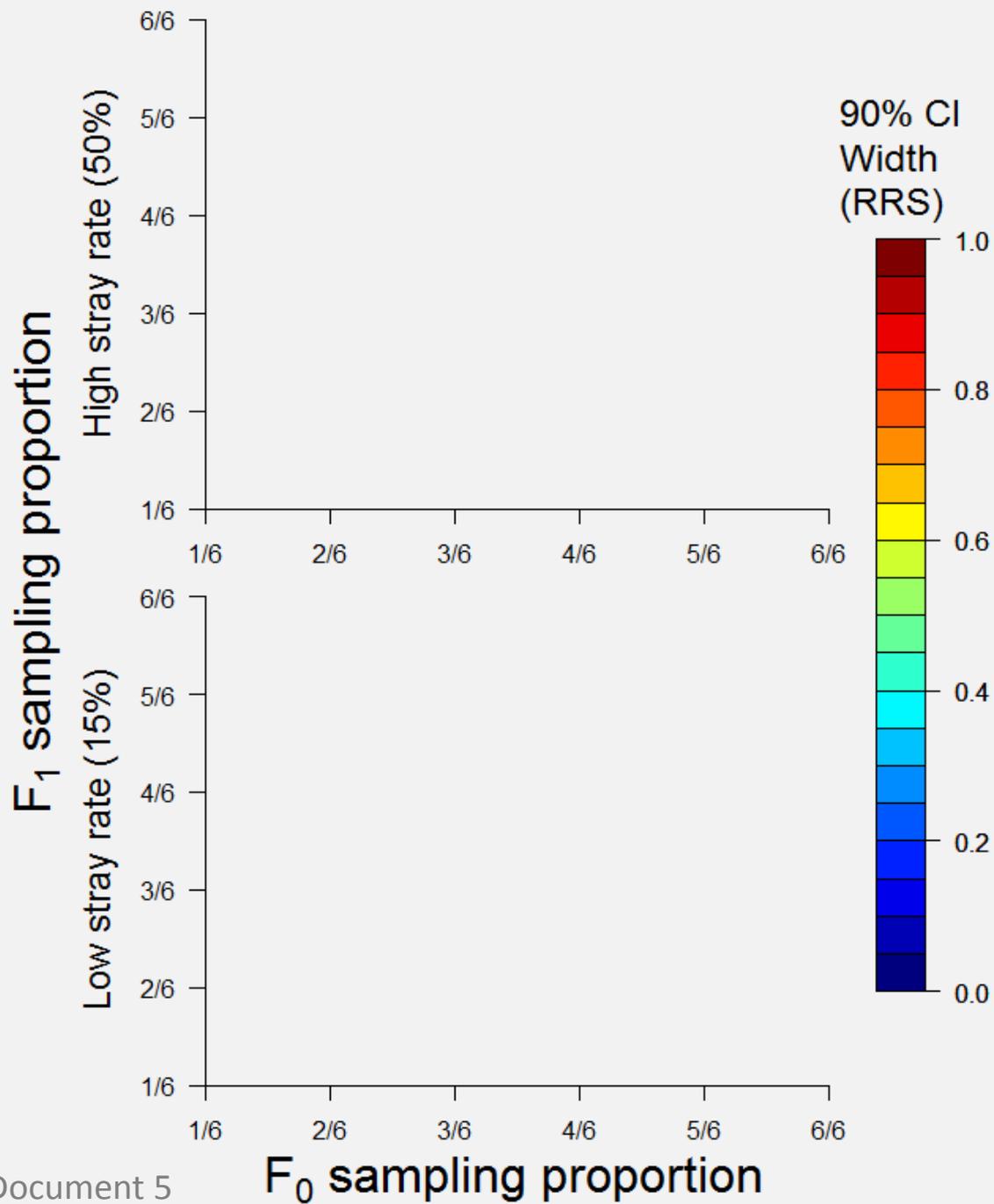
–  $RRS = 1/e^\beta$

```
Coefficients:
              Estimate Std. Error z value Pr(>|z|)
(Intercept) -1.05022    0.09076  -11.571  < 2e-16 ***
OriginW      0.42128    0.11646   3.617  0.000298 ***
---
Signif. codes:  0 '***' 0.001 '**' 0.01 '*' 0.05 '.' 0.1 ' ' 1
```

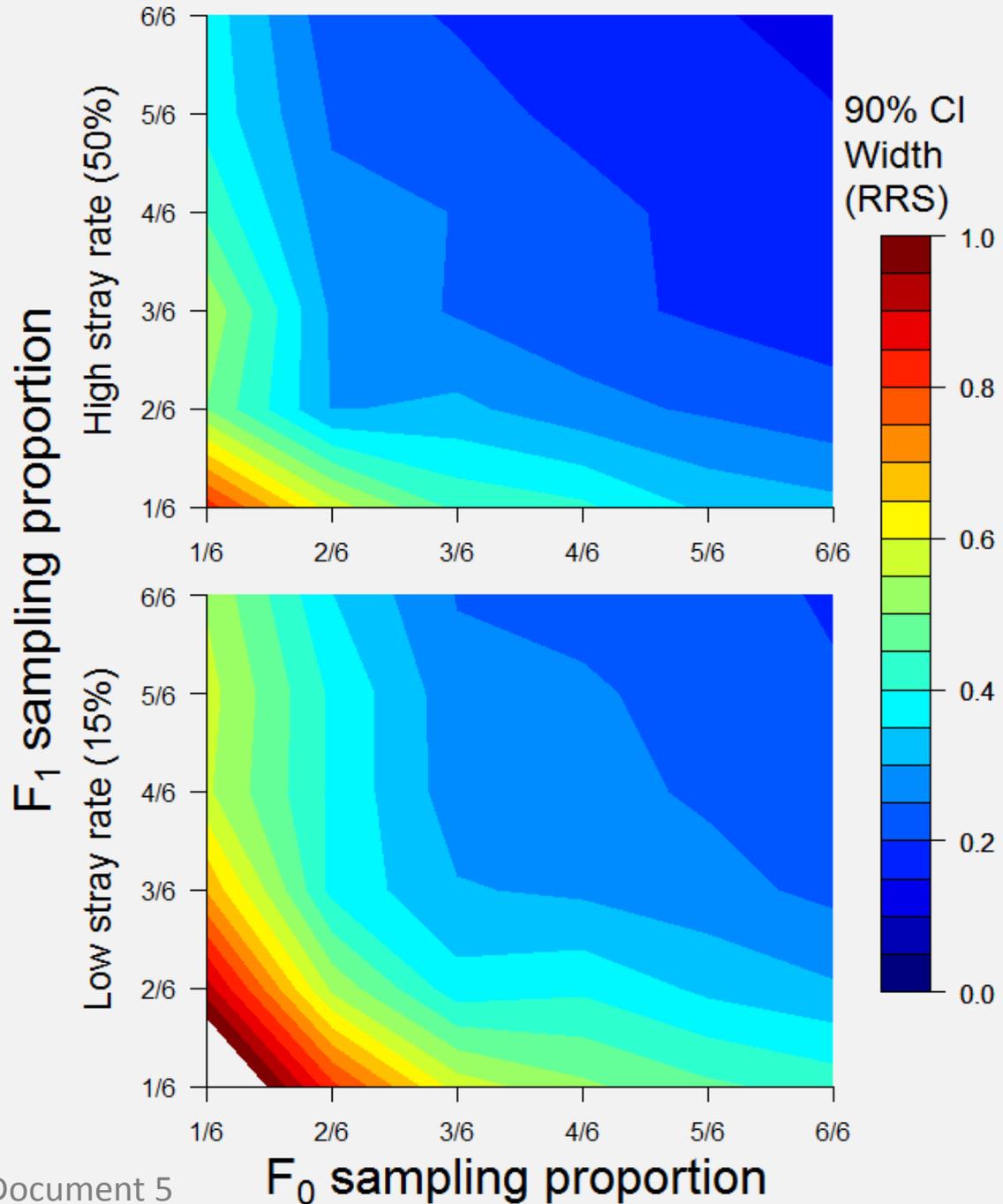
# Precision of RRS estimate



Single parent results:  
Negative binomial 90% CI width

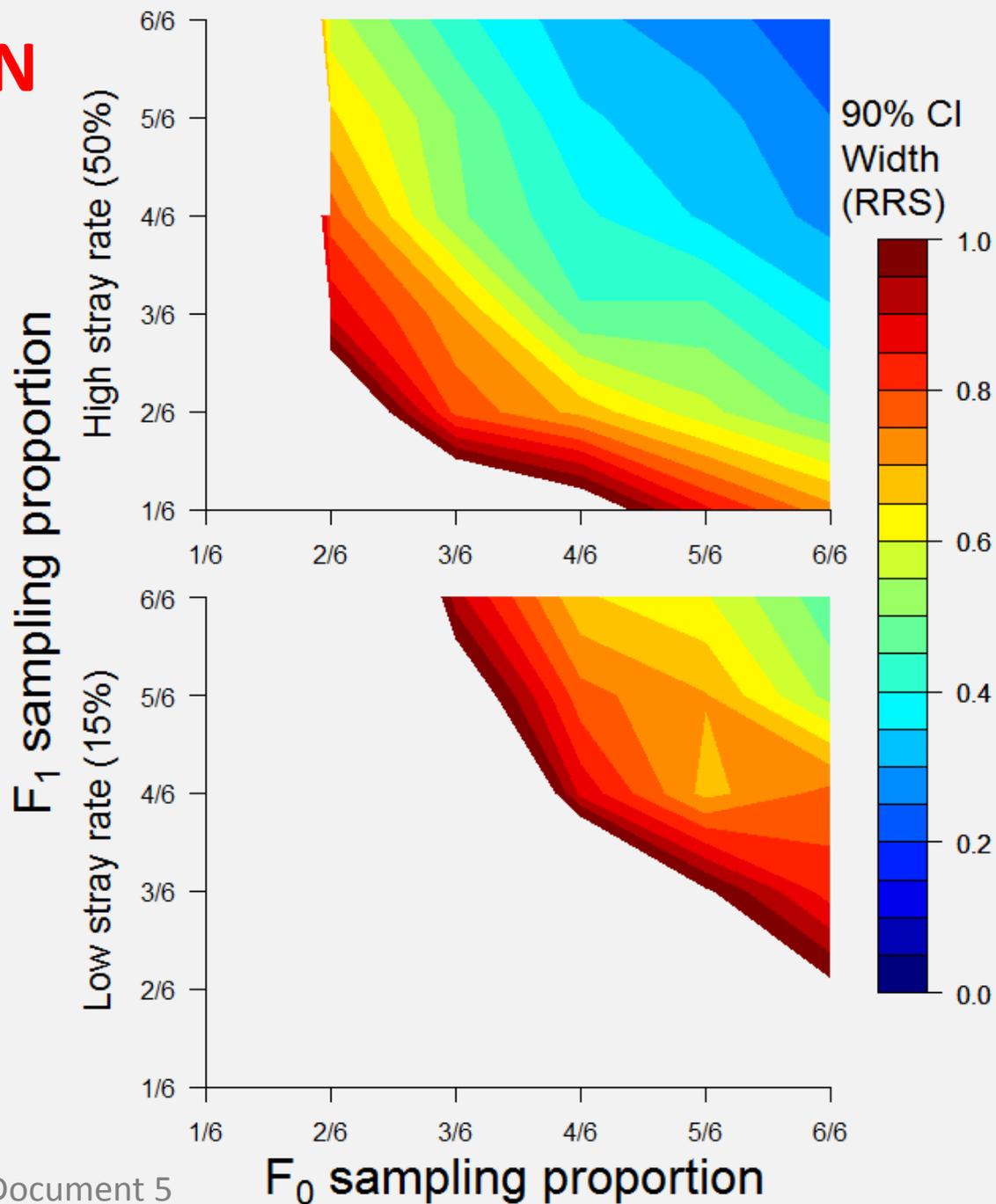


# Single parent



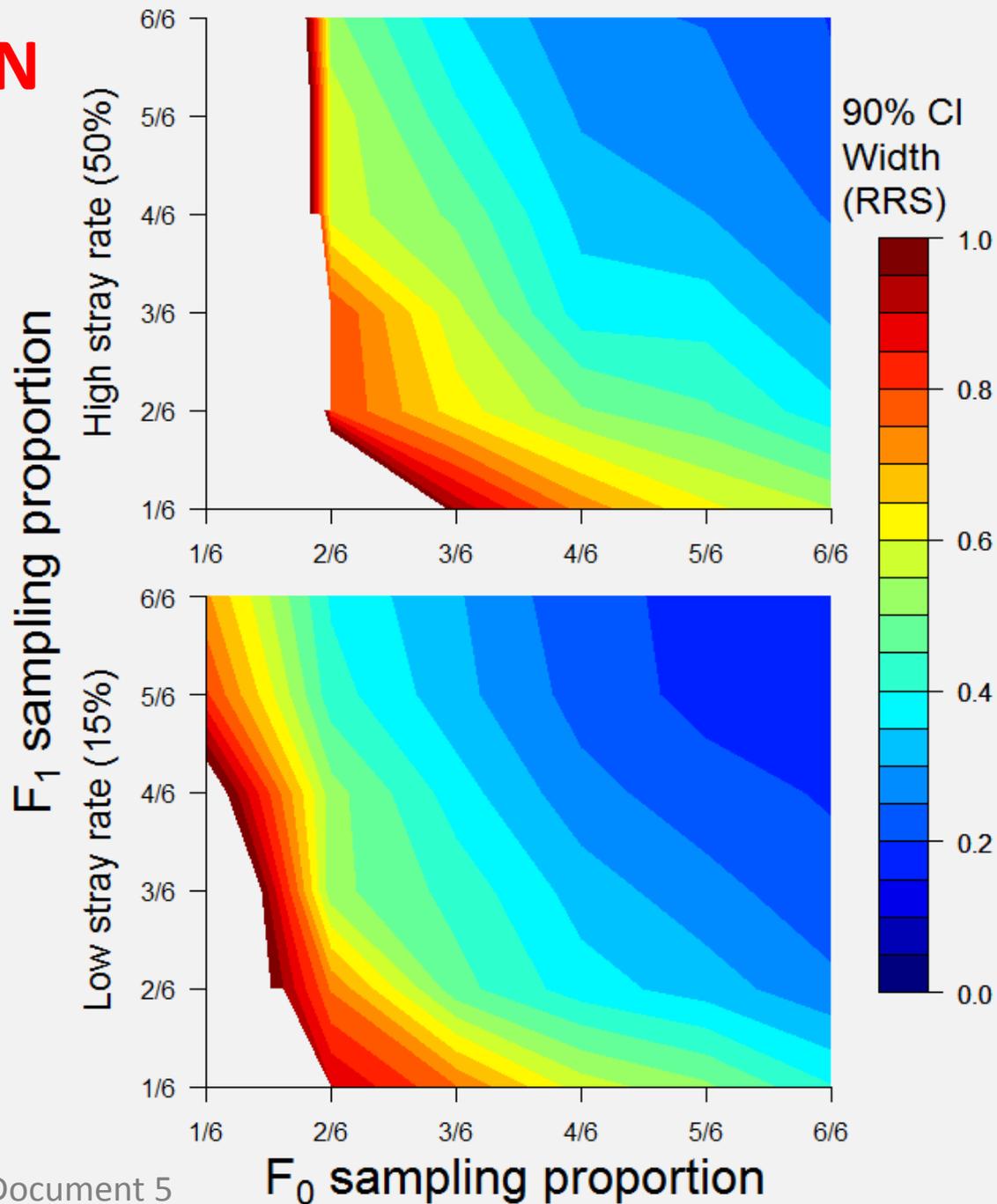
Parent pair results:  
Negative binomial 90% CI width  
**H/H** to **N/N** cross type

# H/H to N/N cross type



Parent pair results:  
Negative binomial 90% CI width  
H/N to N/N cross type

# H/N to N/N cross type



# Conclusions from simulations

- Low power with 1/6 sampling
- Lower power for low stray rate
- Lower power for cross type RRS
- Increases faster with  $F_0$  sampling

**BREAK: Questions so far?**

# Outline

- Background of AHRP
- Parentage and RRS
- Proposed study design
- Simulations
- Power analysis
- **Christie et al. 2014 review**

# Christie et al. 2014

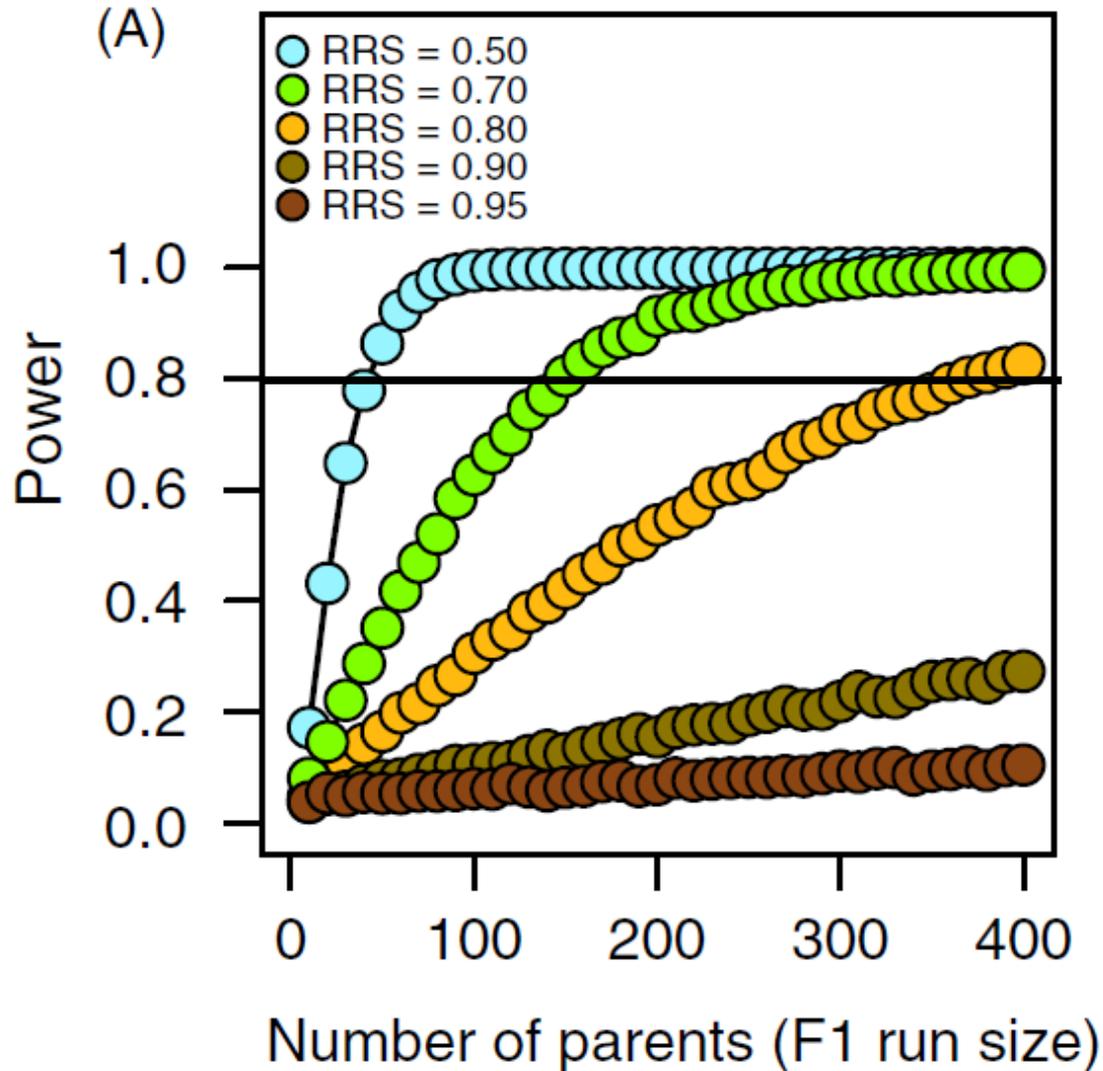
Christie, M. R., M. J. Ford, and M. S. Blouin.  
2014. On the reproductive success of early-generation hatchery fish in the wild.  
*Evolutionary Applications*:7(8).  
<http://dx.doi.org/10.1111/eva.12183>

- Review of early-generation RRS studies
- Examine statistical power of RRS studies

# Christie et al. 2014 Box 2

- Sample size and power for RRS
  - Vary single parent RRS from 0.5-0.95
  - Sample 5-400 parents
  - 50% stray rate
  - Generate offspring from negative binomial
  - Sample ALL offspring
  - Test for KNOWN fitness difference
  - Repeat 5,000 times
  - Power = % of tests that are “significant”

# Christie et al. 2014 Box 2



# Christie et al. 2014 Box 2

- Caveats

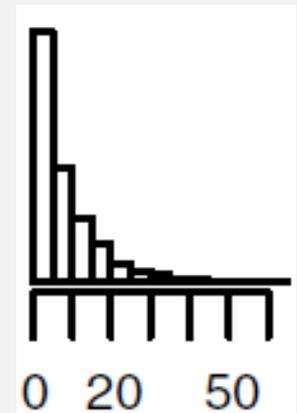
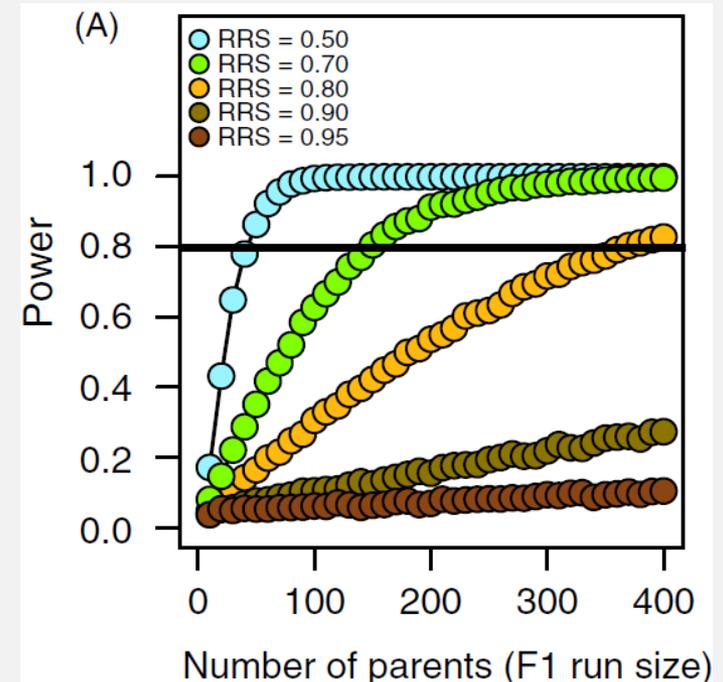
- RRS = 0.5, 0.7, 0.8, 0.9, 0.95

- Stray rate = 50%

- # Parents 5-400

- Sampled ALL offspring

- Negative binomial of offspring had  $\mu = 8$



# Power analysis

## Christie et al. 2014

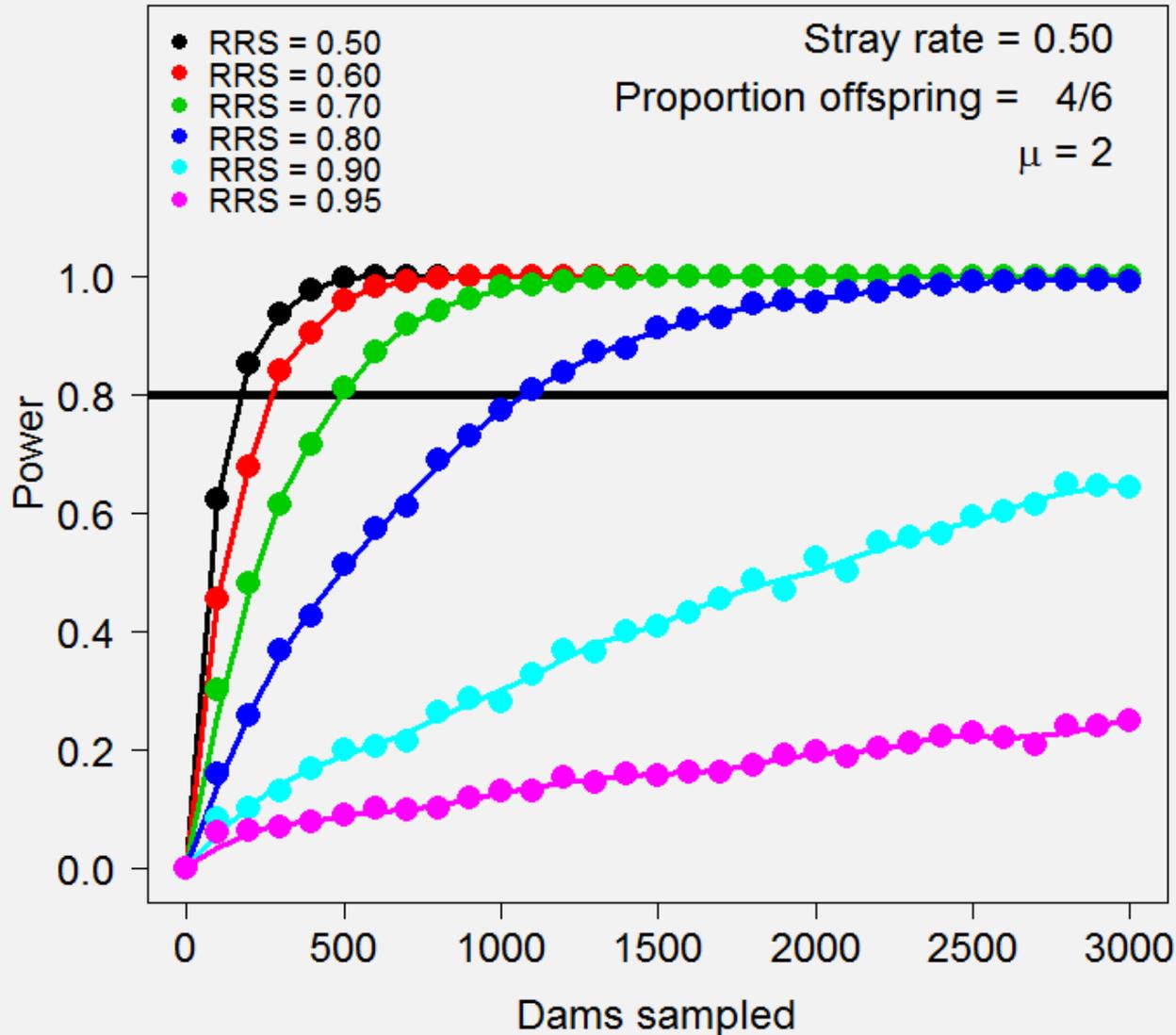
- 5 RRS values
- Sample 5-400 parents
- 50% stray rate
- Negative binomial of offspring  $\mu = 8$
- Sample ALL offspring

## What I did

- Many RRS values
- Sample 0-3000 parents
- 50% & 15% stray rate
- Negative binomial of offspring  $\mu = 2$
- Sample proportions of offspring

# What I did

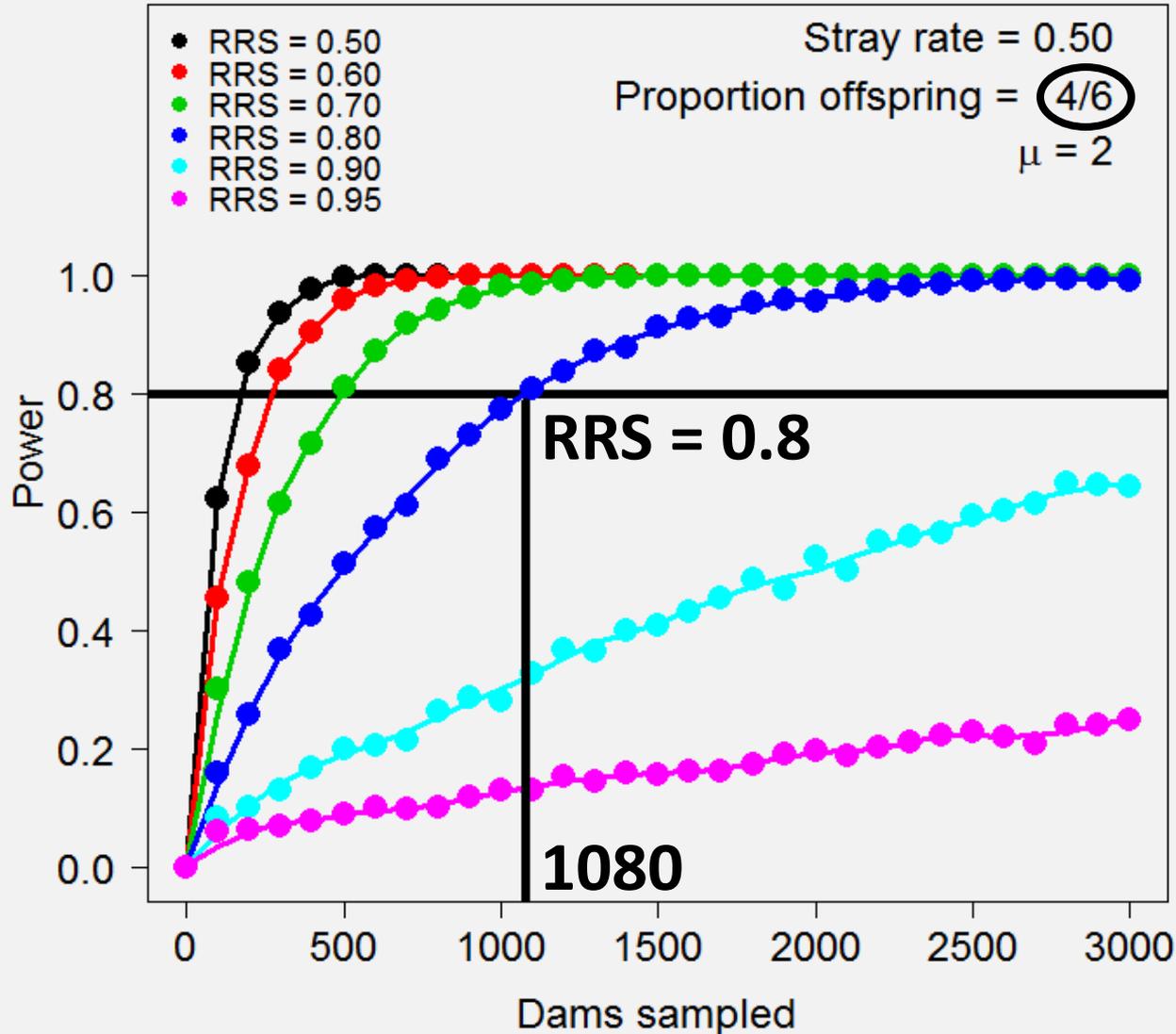
## Permutation Test



See Future AHRP Technical Document

# What I did

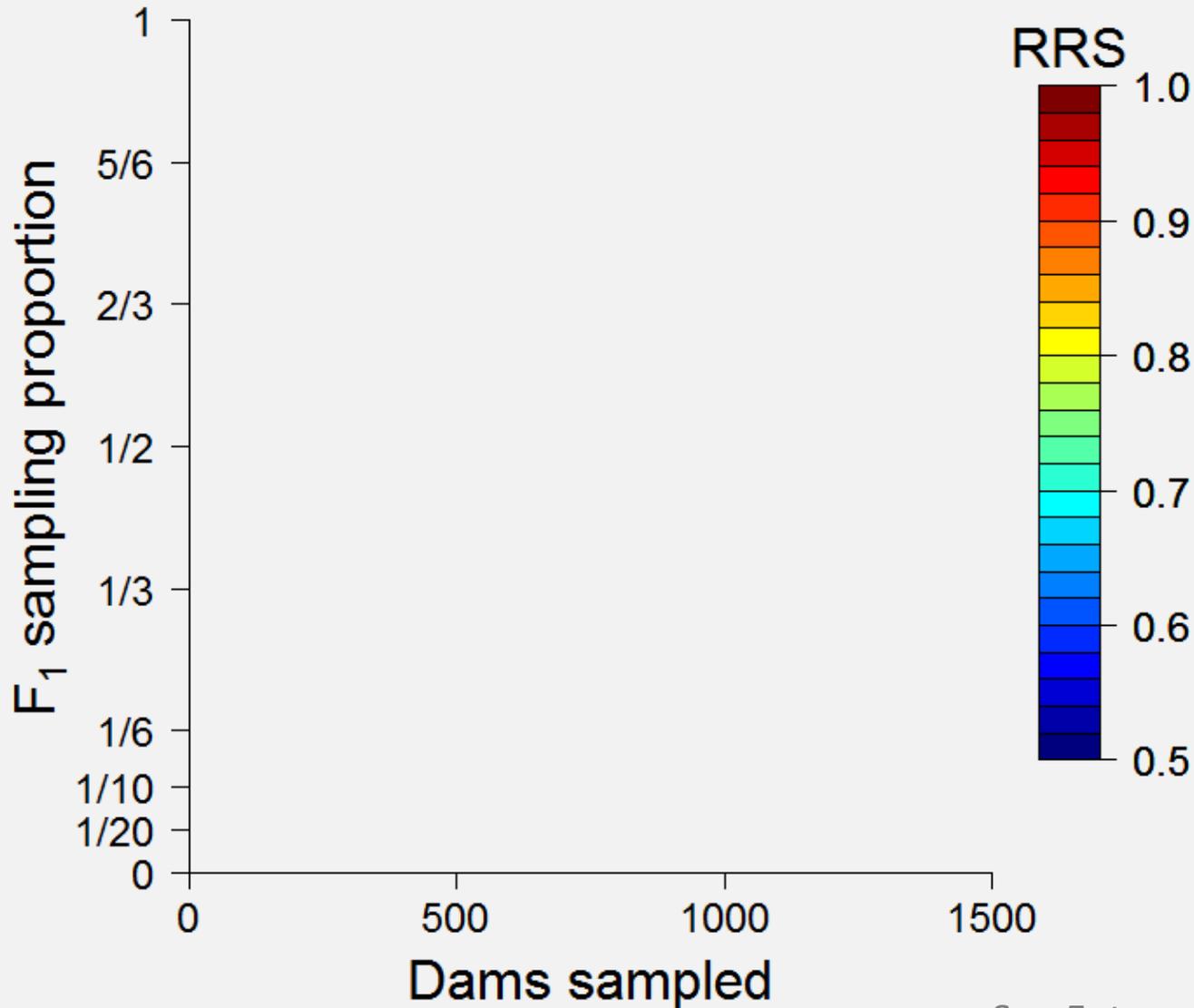
## Permutation Test



See Future AHRP Technical Document

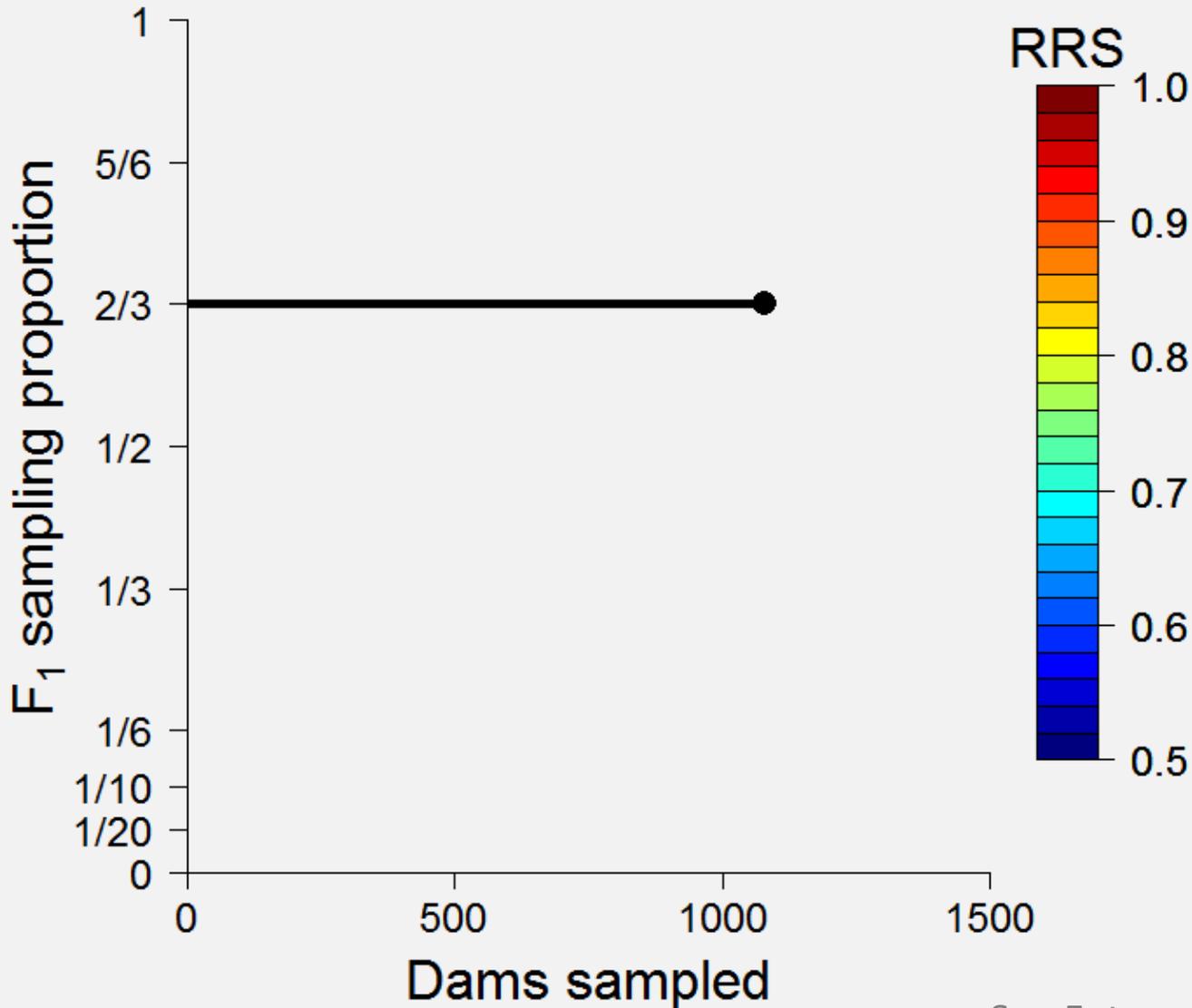
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



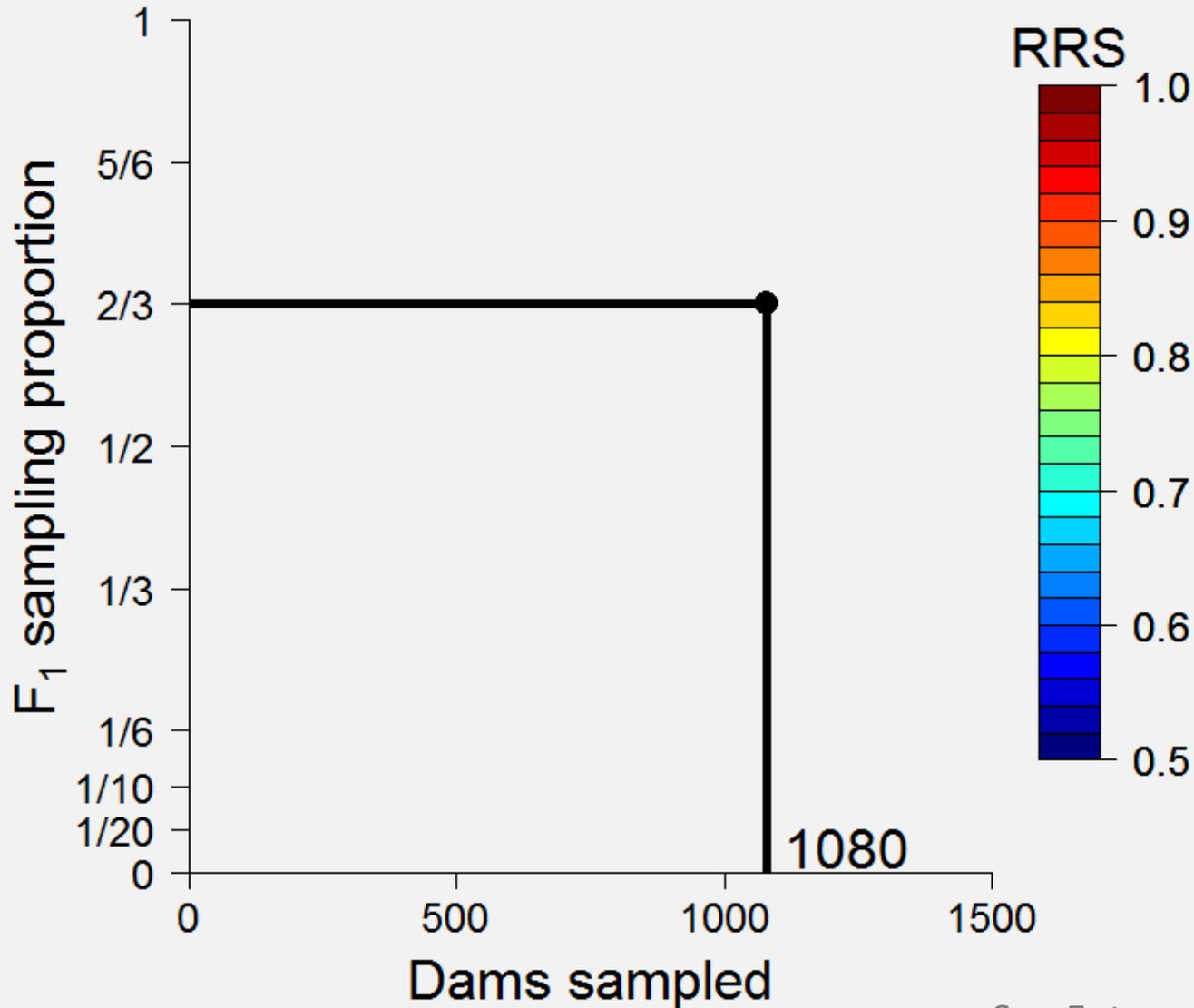
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



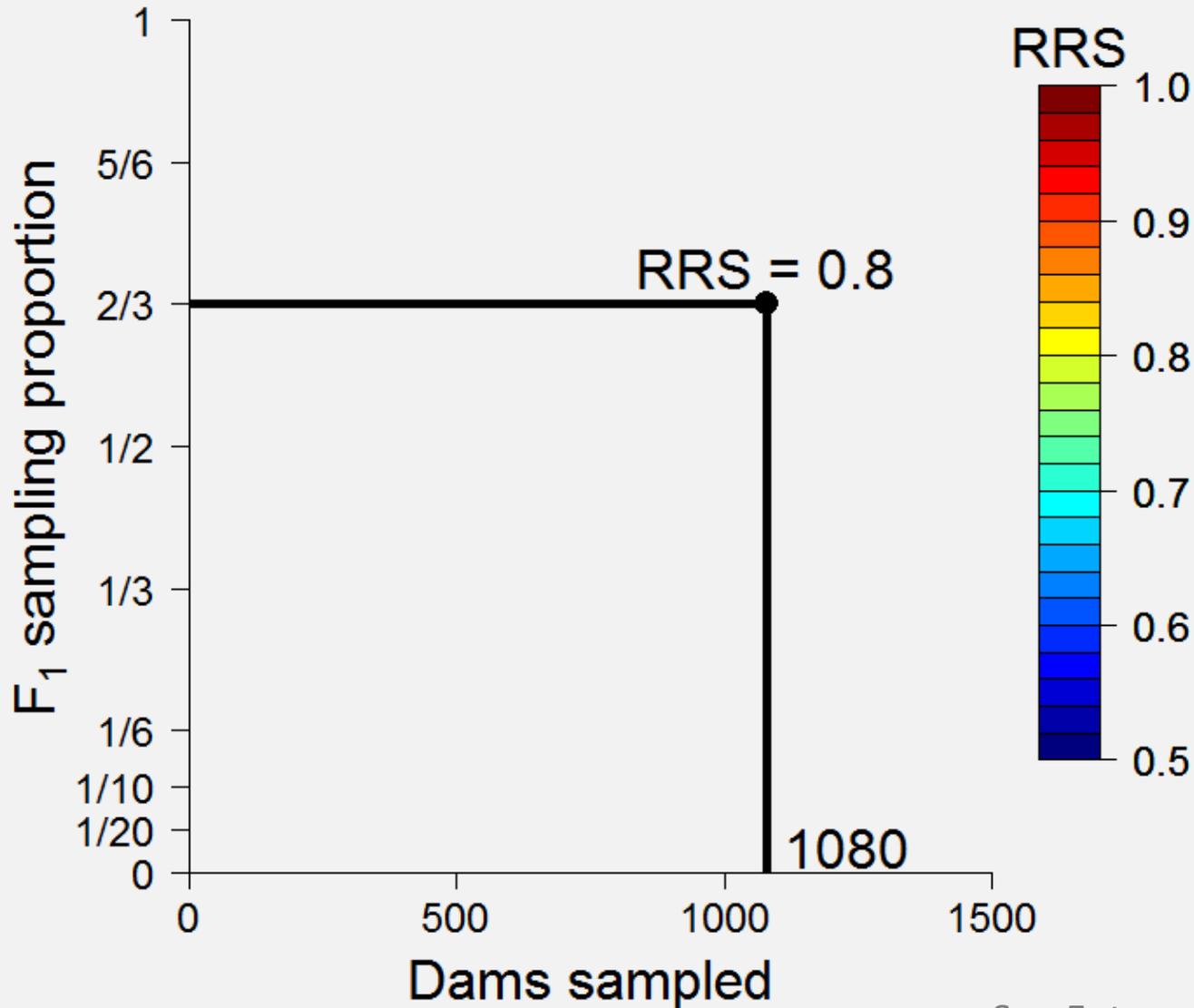
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



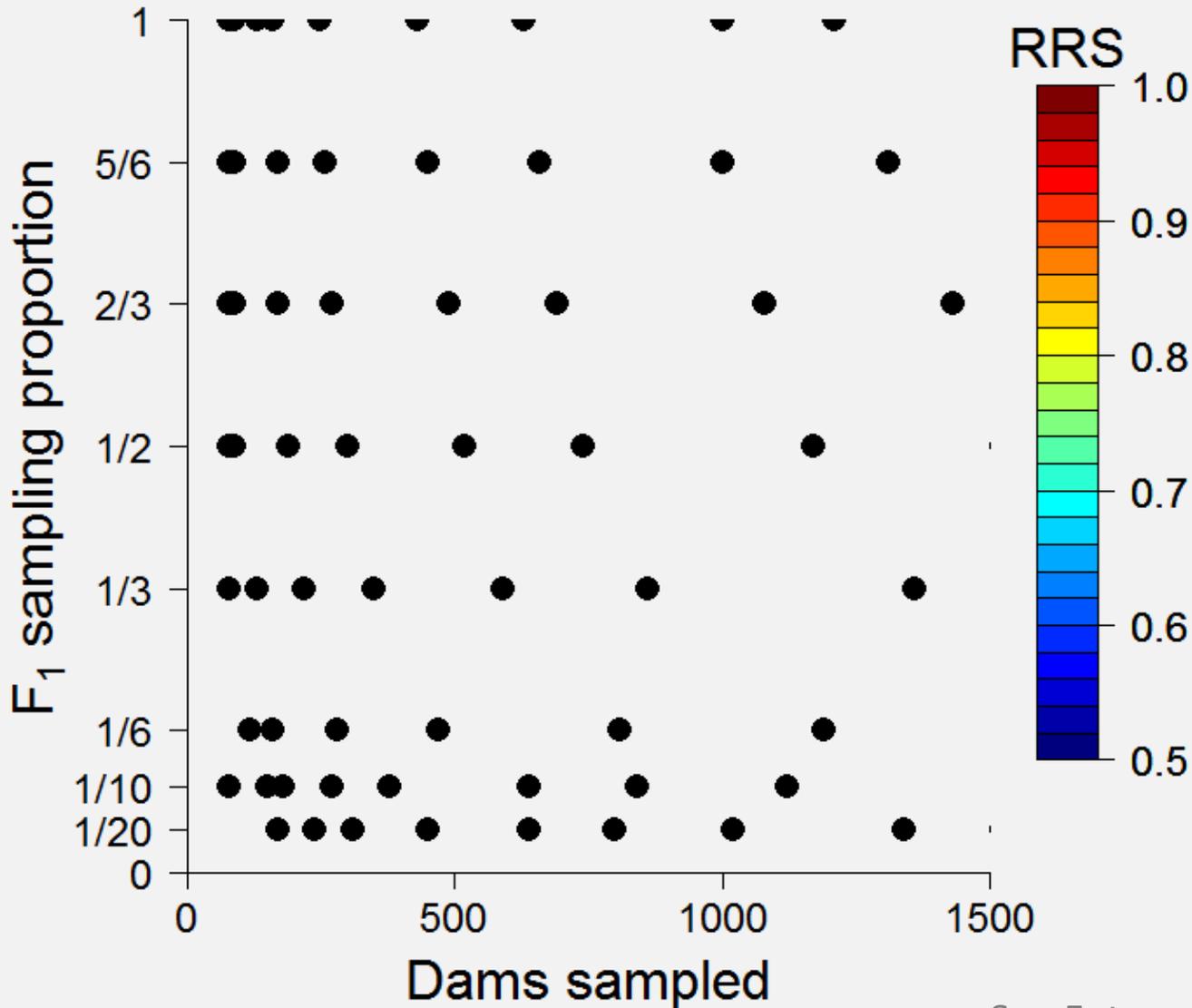
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



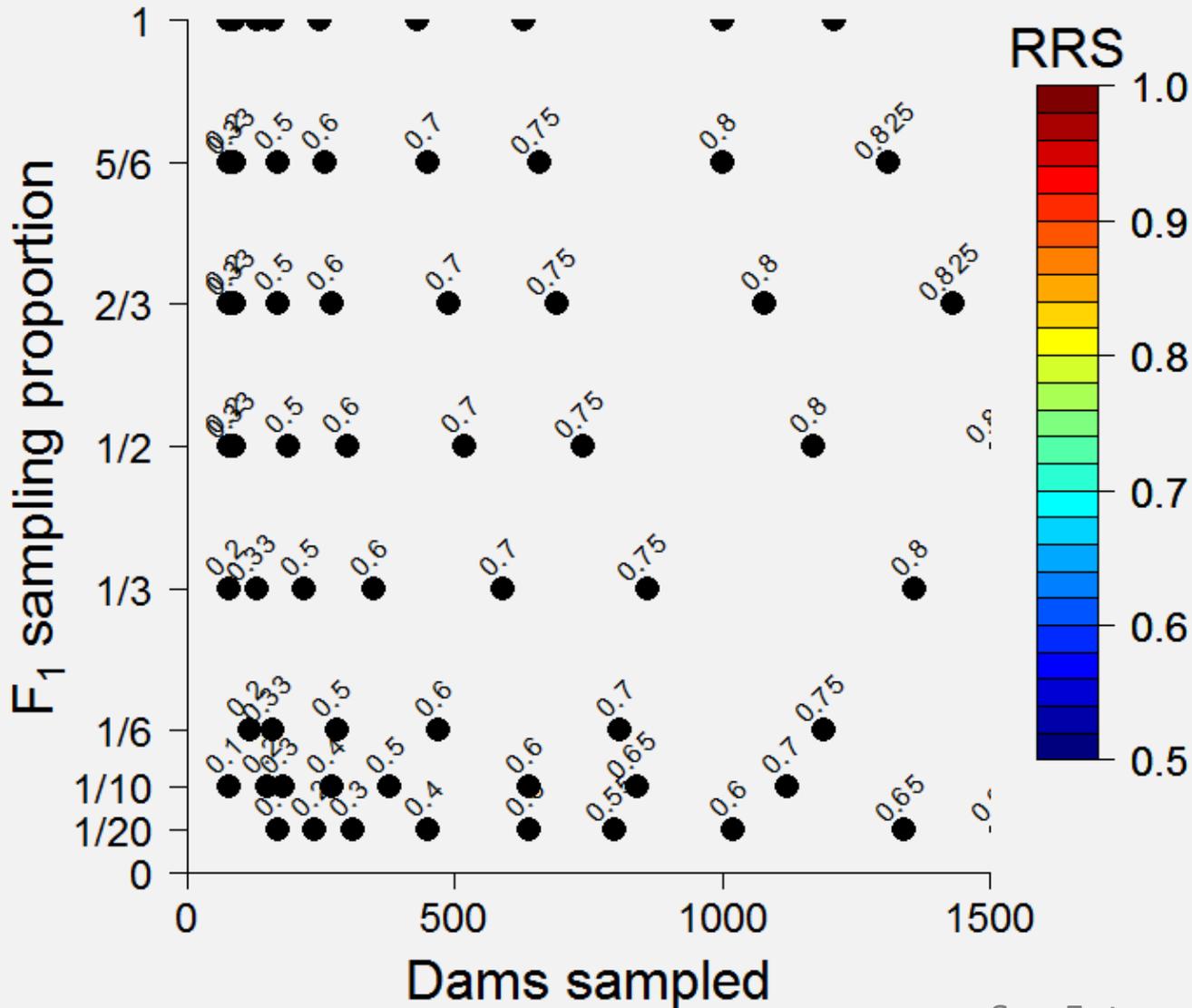
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



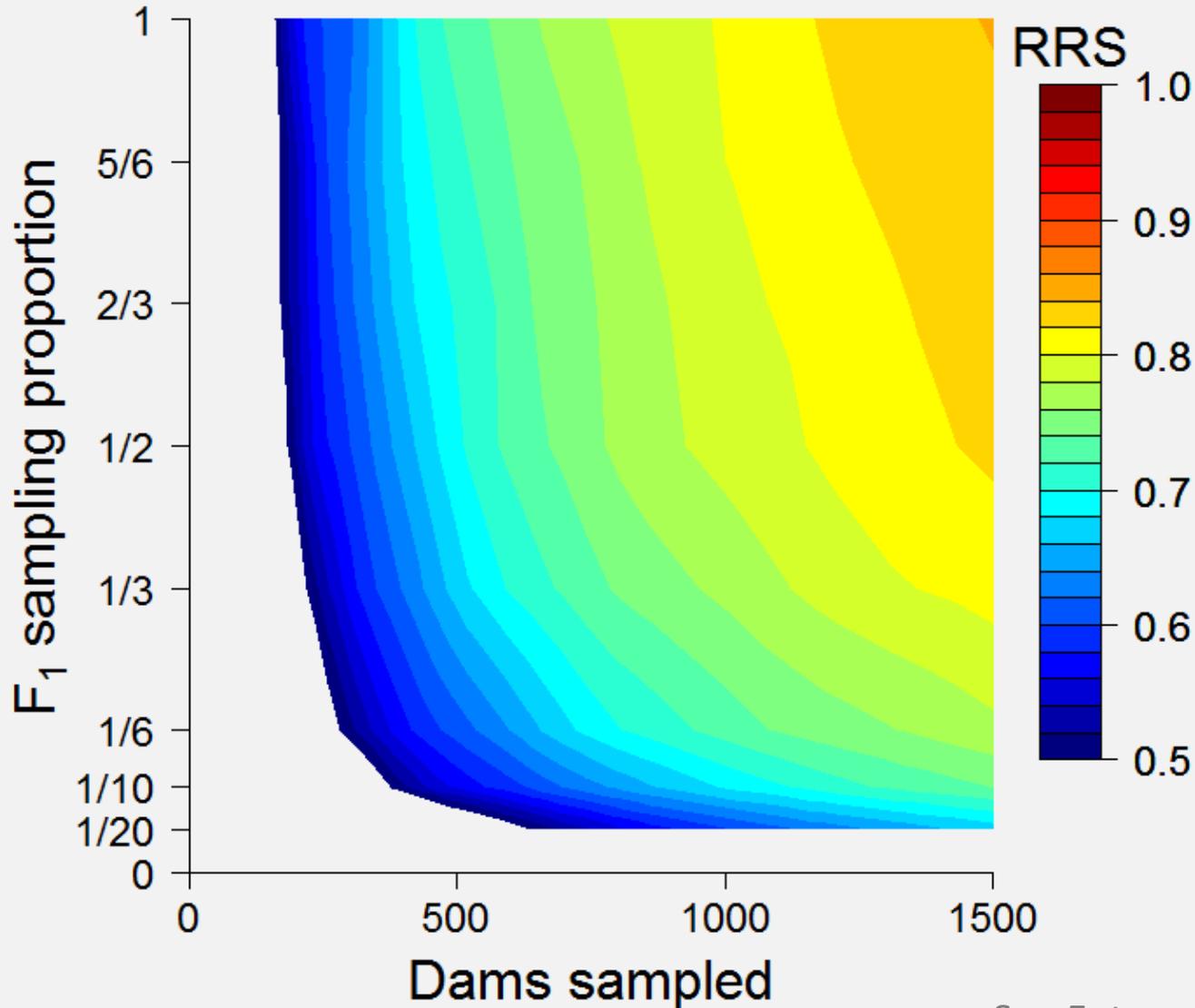
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



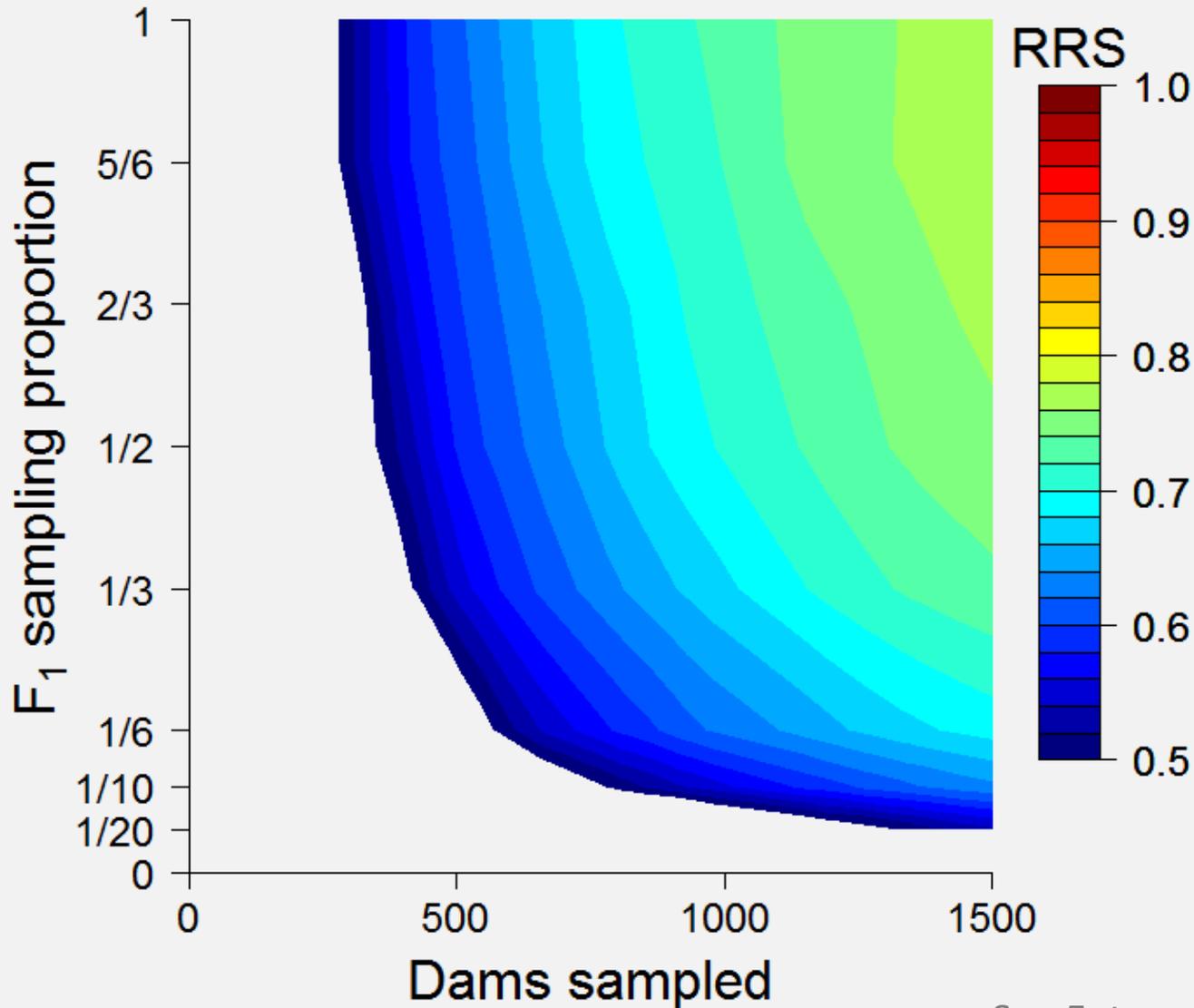
# What I did

Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



# What I did

Stray rate = 0.15;  $\mu = 2$ ; Power = 0.8



# Recommendations

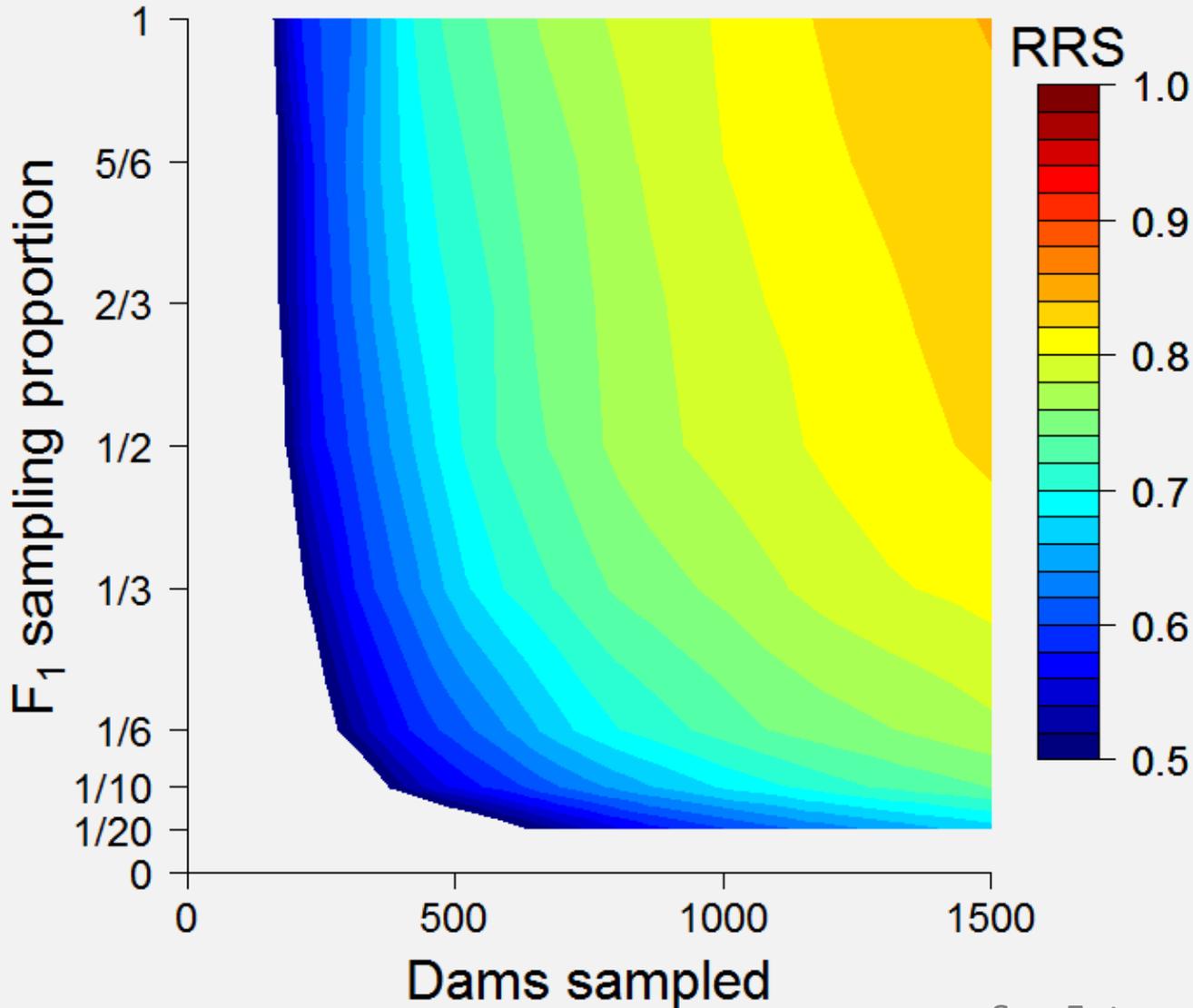
- Where do we go from here?
- Where are we?
  - Stray rate?
  - Sample size  $F_0$ ?
  - Sample proportion of  $F_1$ ?
    - Use sample proportion of  $F_0$  (2013 & 2014 ) as proxy

# Sample sizes of $F_0$ – PWS Pink

Location	Stray	2013	2014
Spring	Low	1,351	151
Stockdale	Low	1,195	1,551
Gilmour	Low	NA	669
Hogan	High	829	2,649
Erb	High	637	1,957
Paddy	High	125	1,158

# Sample sizes of $F_0$ – PWS Pink

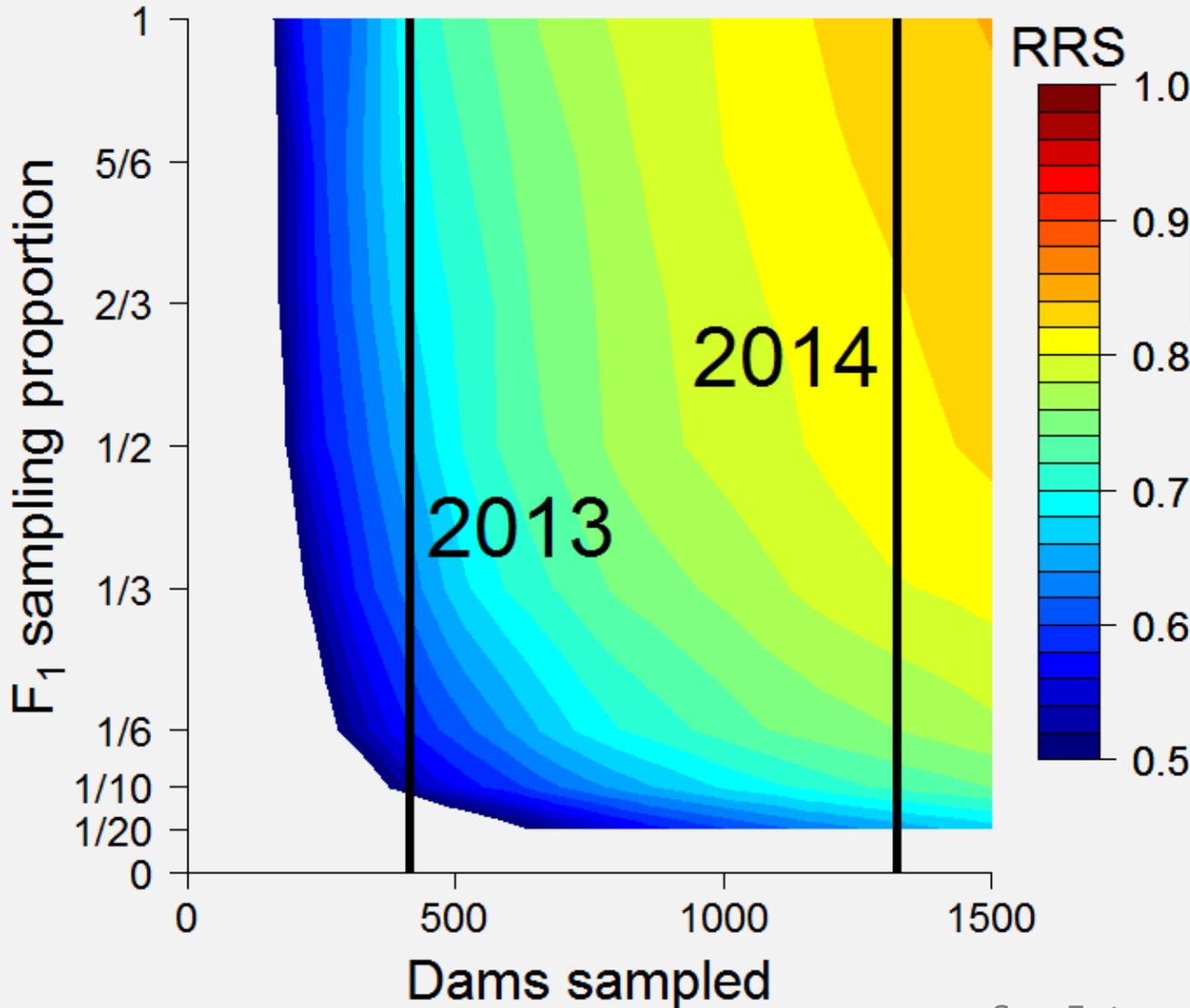
Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



Location	Stray	2013	2014
Spring	Low	1,351	151
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# Sample sizes of $F_0$ – PWS Pink

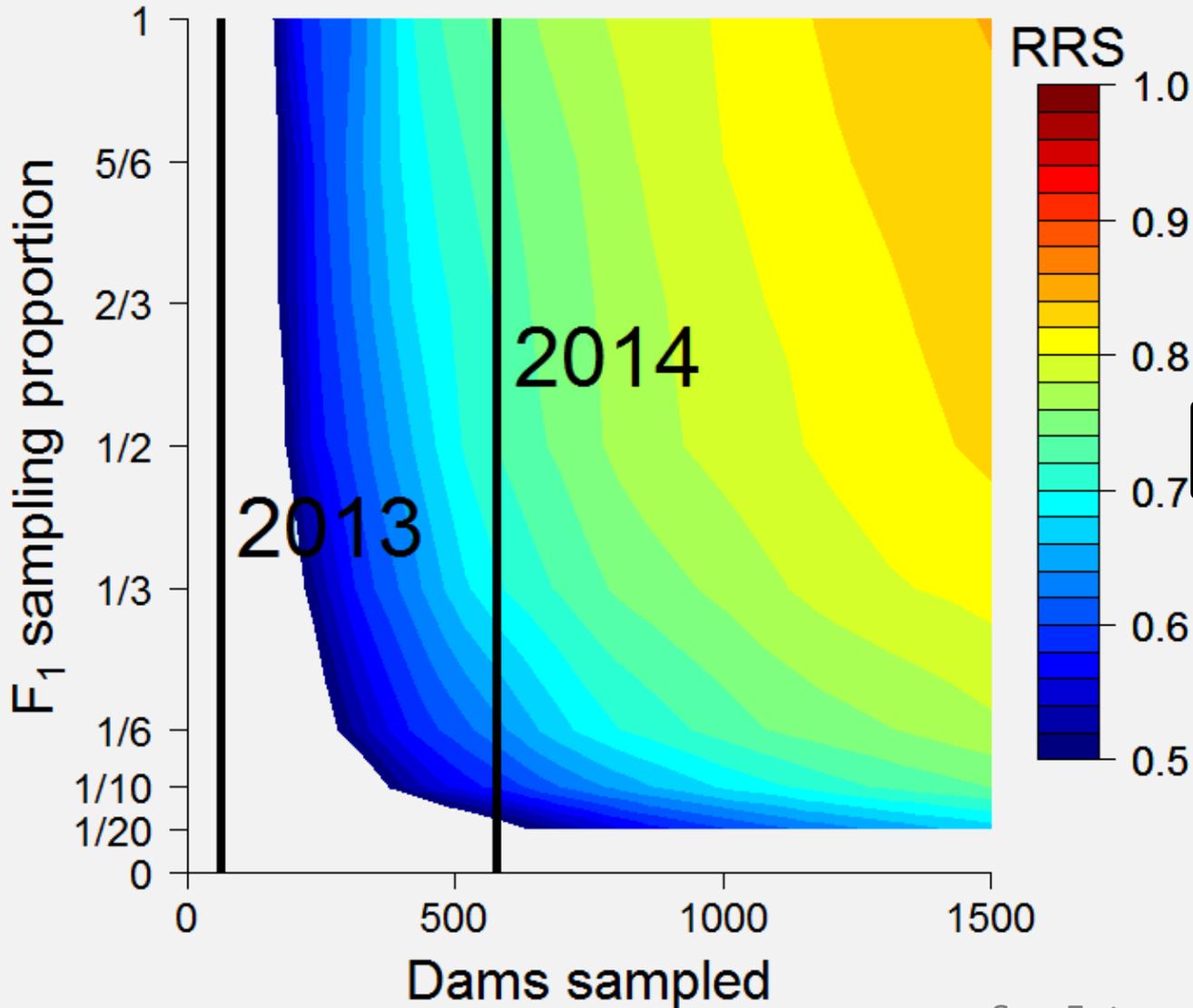
Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



Location	Stray	2013	2014
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# Sample sizes of $F_0$ – PWS Pink

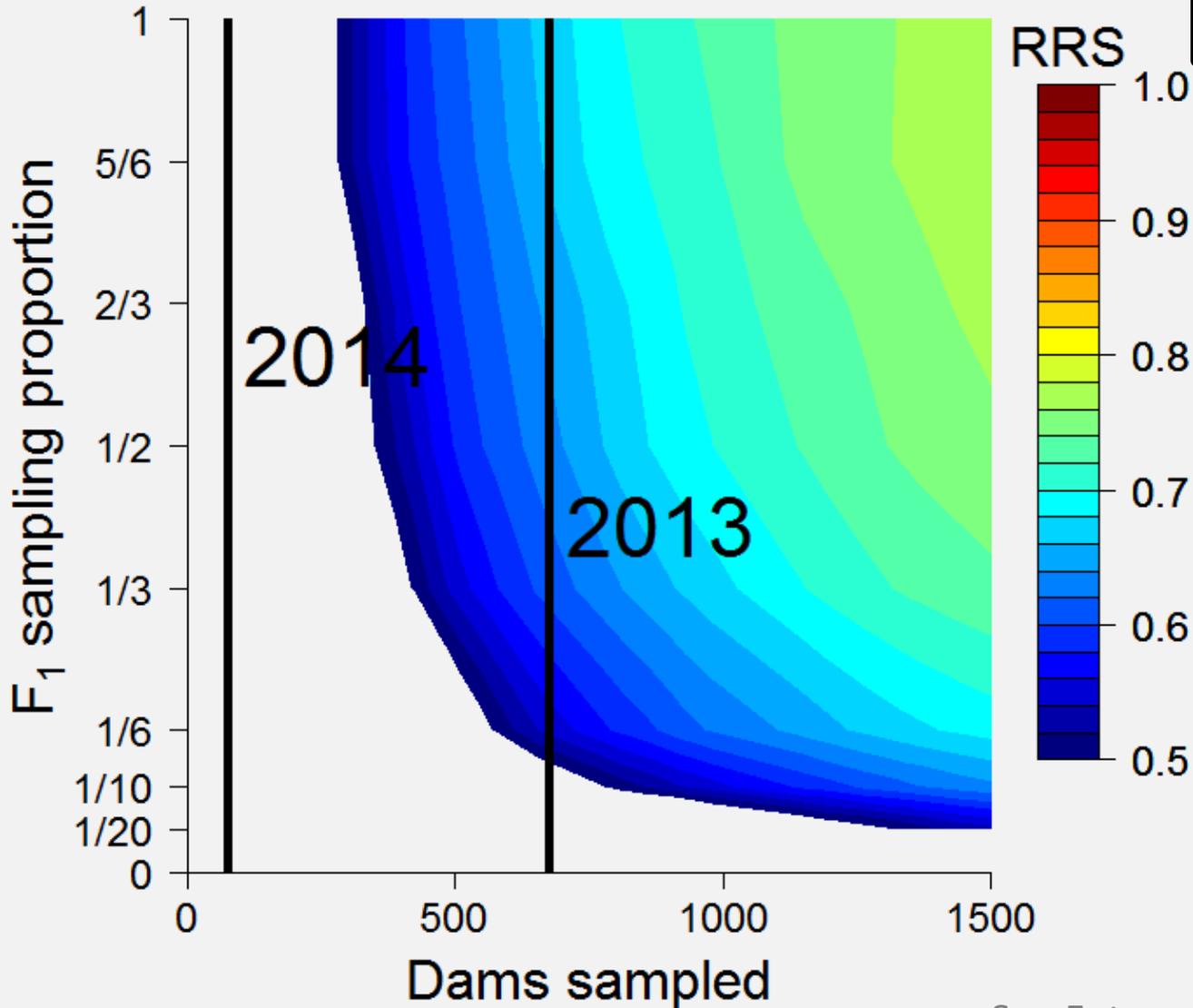
Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



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# Sample sizes of $F_0$ – PWS Pink

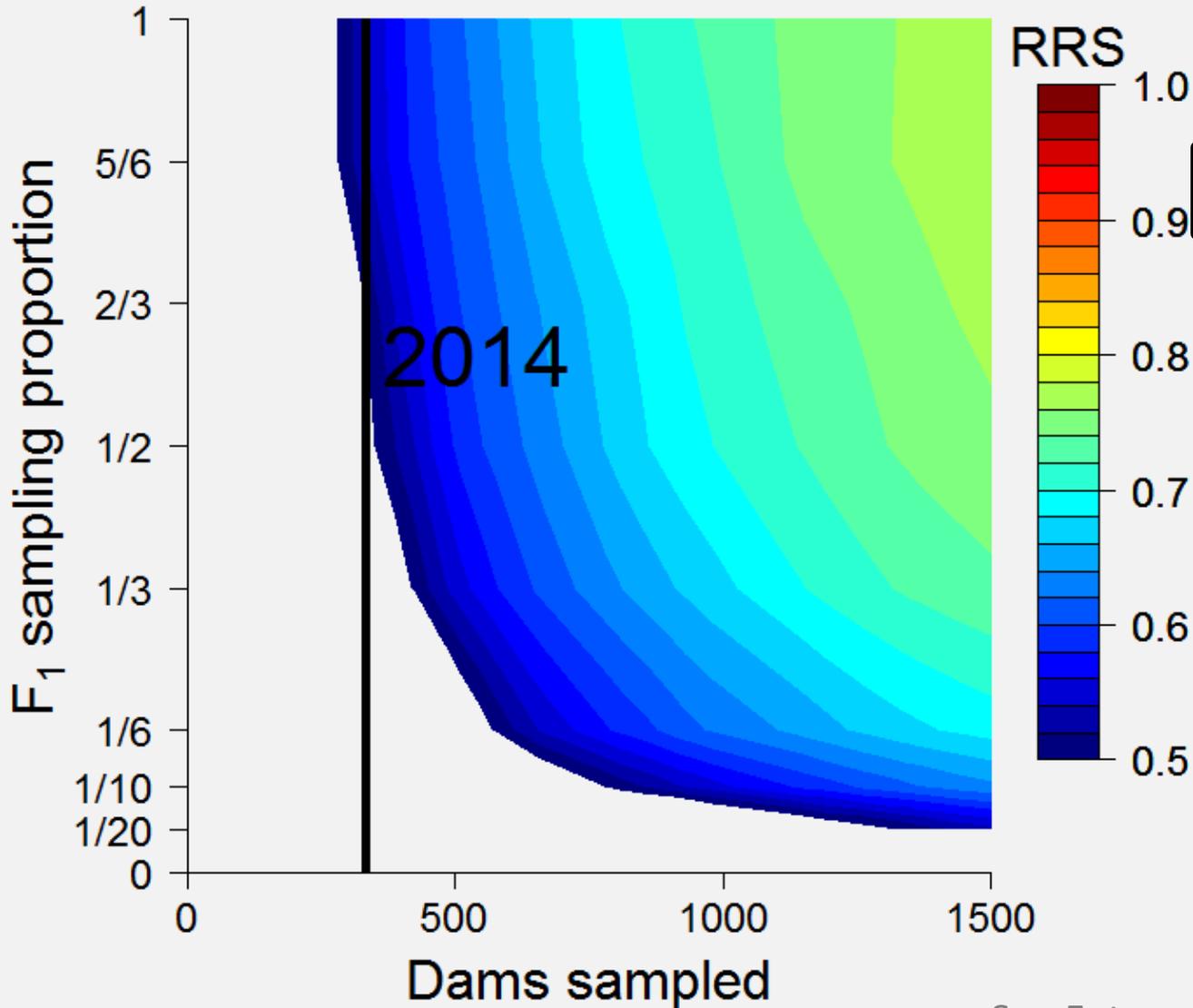
Stray rate = 0.15;  $\mu = 2$ ; Power = 0.8



Location	Stray	2013	2014
Spring	Low	1,351	151
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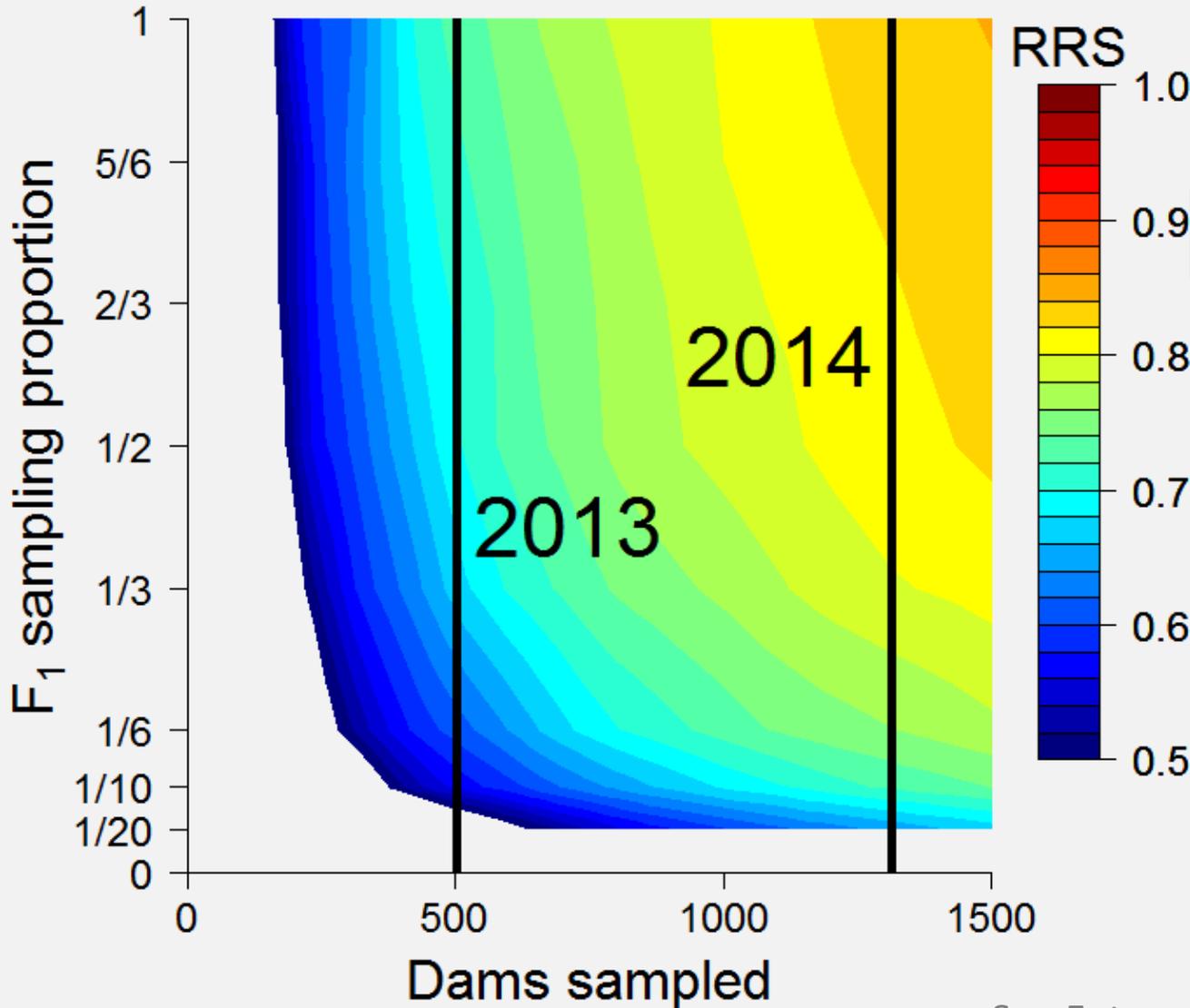
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Erb	High	637	1,957
Paddy	High	125	1,158

# Sample sizes of $F_0$ – SEAK Chum

<b>Location</b>	<b>Stray</b>	<b>2013</b>	<b>2014</b>
Admiralty	Low	421	260
Prospect	Low	487	473
Fish	High	1,008	2,626
Sawmill	High	344	124

# Sample sizes of $F_0$ – SEAK Chum

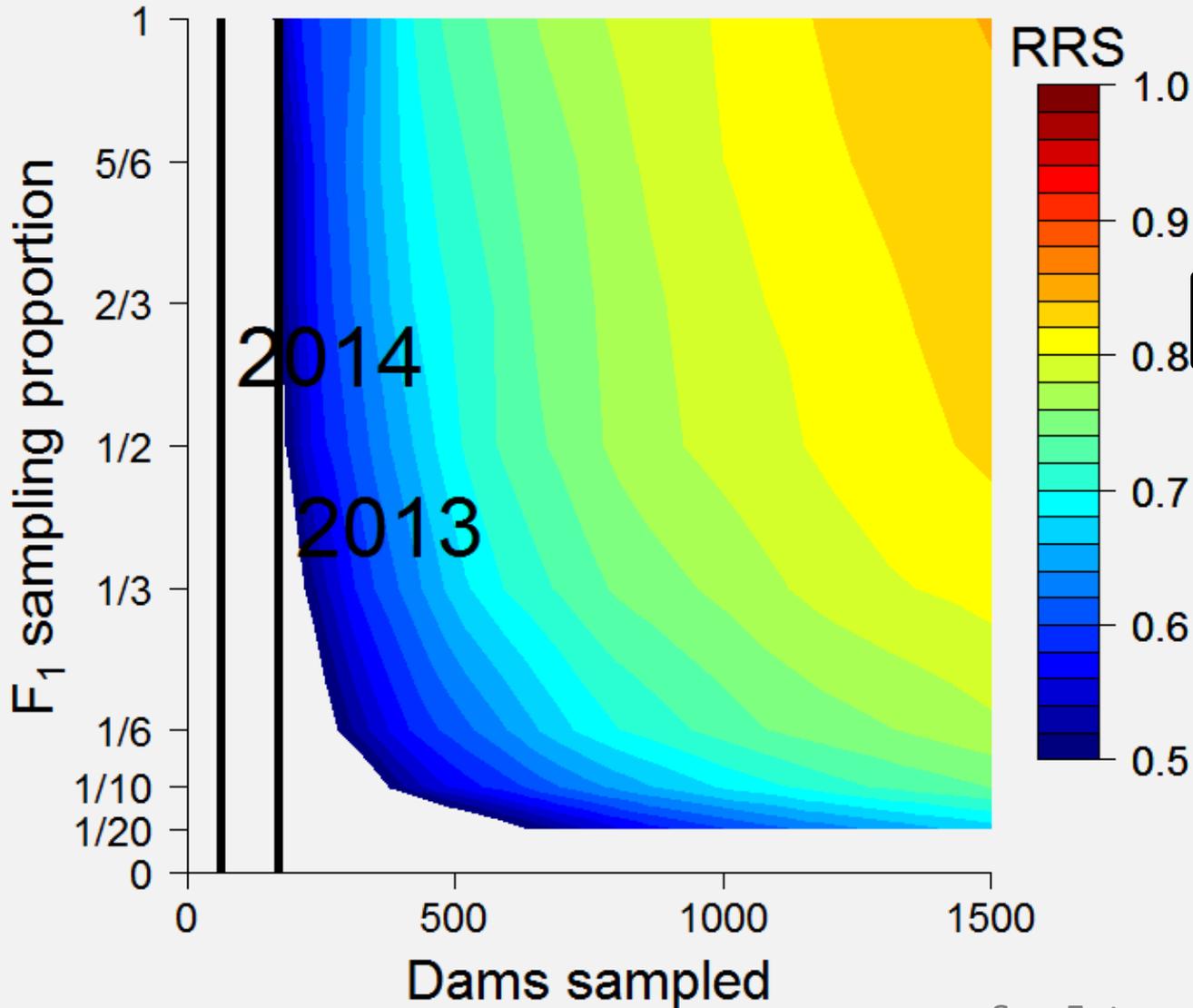
Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



Location	Stray	2013	2014
Admiralty	Low	421	260
Prospect	Low	487	473
Fish	High	1,008	2,626
Sawmill	High	344	124

# Sample sizes of $F_0$ – SEAK Chum

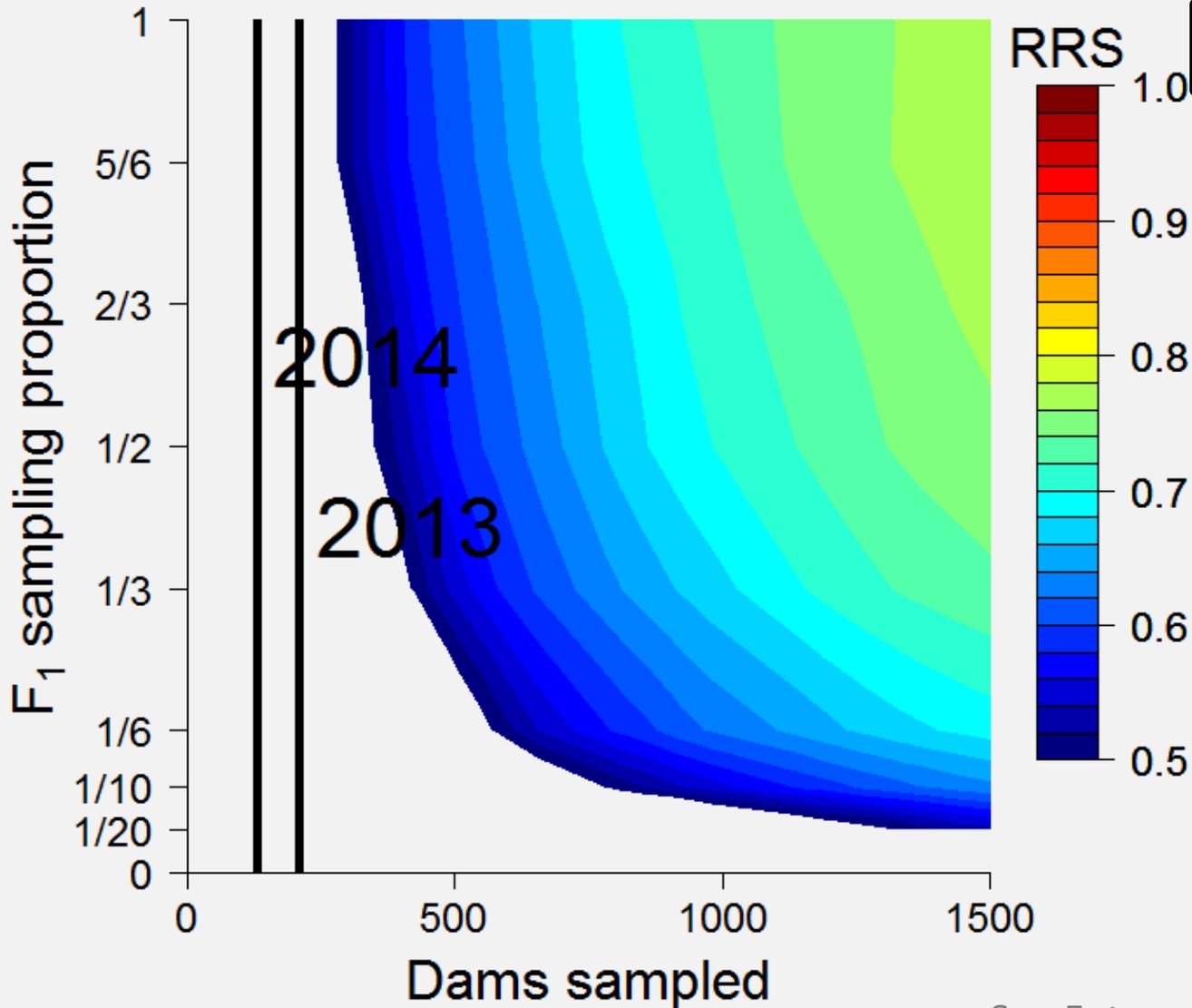
Stray rate = 0.5;  $\mu = 2$ ; Power = 0.8



Location	Stray	2013	2014
Admiralty	Low	421	260
Prospect	Low	487	473
Fish	High	1,008	2,626
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# Sample sizes of $F_0$ – SEAK Chum

Stray rate = 0.15;  $\mu = 2$ ; Power = 0.8



Location	Stray	2013	2014
Admiralty	Low	421	260
Prospect	Low	487	473
Fish	High	1,008	2,626
Sawmill	High	344	124

# Conclusions

## Power depends on

- Stray rate
- Distribution of offspring
  - Shape
  - Mean RS
- Num. families sampled ( $F_0$ )
- Prop. offspring sampled ( $F_1$ )

## Higher power with

50% stray rate

Low variation

High Productivity

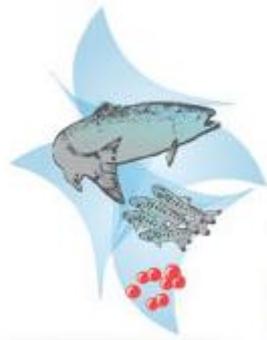
> 500 families

High prop. offspring

# Questions for Science Panel

- What level of power, in terms of maximum RRS that would likely be detectable, is appropriate for this study?
- What are alternative study designs that could increase the proportion of adults and offspring sampled?

# Acknowledgements



# Questions?



